

Keywords: Ginkgo biloba/living fossil/ Rooting/ propagation/ Indole - 3-butyric acid/ Bavistin/ medicinal tree/ conservation

## 1. Introduction

Unfortunately, due to legal and illegal exploitation of important plant species from wild, anthropogenic pressure and lack of knowledge about sustainable harvesting of useful bioresources particularly medicinal ones as many of them has been listed under the categories of rare, threatened, endangered or at the verge of extinction. Although the extinction of species is a natural process, the current speed of extinction of species through human interventions is approximately 100-1000 times faster than the natural speed of extinction. In many groups of organisms $5-20 \%$ of all species are already extinct (Chaplin et al. 2000). The species currently on earth are the result of a natural selection process over the last three billion years, which has lead to a large degree of specialization. Species diversity is therefore a prerequisite for ecosystem to function. But how important are species and ecosystems for society? This can be researched by investigating the functions and associated goods and services of ecosystems that are important for humans. A central problem is a
continuing loss of biodiversity, but our knowledge about human dimension of biodiversity is still relatively limited. However, there are several plant species growing in this earth but has not still given priority for conservation due to lack of awareness about their usefulness, economic potential and applicable multiplication technology package.
The Ginkgo biloba Linn. (MAIDENHAIR TREE; family Ginkgoaceae) is worldôs oldest tree mostly known as living fossils and only surviving member of seed plant groups. It is found growing naturally in very limited localities in the central Himalayan mountain at an elevation of 6000 ft (Anonymous, 1999). It is a handsome, straight, up to 100 ft . high, sparsely branched when young, bearing clusters of fan shaped leaves and dioecious slow growing gymnosperm and known to have huge medicinal, spiritual and horticultural importance worldwide. The Leaves of this species is extensively used in the form of a concentrated, standardized Ginkgo biloba extract (GBE) in different countries (particularly China, Europe, France and Germany) as a source of herbal


Ginkgo is being used to treat circulatory disorders and enhance loss of memory. Its leaves contain two types of chemicals (flavonoids and terpenoids) have potent antioxident properties. It is also widely used for treating dementia, eye problems, intermittent claudicating, memory impairment, tinnitus and a variety of other ailments including altitude sickness, asthma, depression, disorientation, headaches, high blood pressure, erectile dysfunction, and vertigo (http://www.umm.edu/altmed/ConsHerbs/GinkgoB ilobach.html).
In central Himalayan Mountains of India, there are very few spots in Uttarakhand (i.e. Ranikhet, Nainital and Dehradun) where the individuals of this species are found growing naturally and require immediate conservation measures. However, countries like China, Europe, France, and Germany have already undertaken initiatives for large-scale propagation and plantation for its conservation so as to maintain its status and population on one hand and to make use of it properly in the field of herbal medicine in near future. Despite of having huge medicinal properties and ornamental value, this species still has not received much attention as far as conservation is concerned particularly in India. Due to poor regeneration, only few individuals exists in the nature particularly in diverse climatic conditions in different places and facing serious threat of extinction from central Himalayan mountains of India. Since this is a rare species and has got the status of oldest living tree fossils and therefore require urgent propagation protocol for large-scale multiplication. Rooting of stem cuttings provides the advantage of greater genetic uniformity and availability of superior stock in a short period of time for afforestation works. This method has been tried sufficiently in number of gymnosperms as well as angiosperms trees (Nandi et al. 1996, Tamta et al. 2000, Nandi et al. 2002, Purohit et al. 2005). The vegetative propagation studies on Ginkgo biloba has been reported by Dirr et al. 1987, Doran, 1954 and Natalia, 1994). Using auxins, but there are very few studies or reports available in Indian Himalayan region where phenolic compounds either alone or in combination with auxins were applied (Prakash et al. 2002 and Gopichand et al. 2006). In view of this, an experiment on vegetative propagation using rooting of branch cuttings was carried out to examine the effect of IBA and Bavistin under different microclimatic conditions.

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## 2. Material and methods

The experiments were carried out inside the polyhouse, polypit and in the open conditions (details given below) established in the G. B. Pant Institute of Himalayan Environment \& Development, Garhwal, Unit, Srinagar, Garhwal, Uttarakhand, India, ( $30^{\circ} 13^{\prime} 05.8^{\prime \prime} \mathrm{N}$ and $78^{\circ} 46^{\prime}$ $24.5^{\prime \prime} \mathrm{E} ; 584 \mathrm{~m}$ amsl). Branch cuttings (semi hardwood) were excised in the morning and brought to the laboratory in polythene bags in order to prevent desiccation, during the first week of January 2006 from mature single male and female tree of G.biloba (of more than 50 years old trees) growing in Kalika ( 1750 m amsl) and Chaubatia ( 1850 m amsl ) areas of Ranikhet (Distt. Almora, Uttarakhand, India), respectively. The final cuttings (average length $10.52 \pm 0.88 \mathrm{~cm}$ male and $11.14 \pm 0.32 \mathrm{~cm}$ female, dia. $8.82 \pm 0.34 \mathrm{~mm}$ male and $9.21 \pm 0.41 \mathrm{~mm}$ female, with atleast 3-4 nodes per cutting) were clipped from the branches. The basal 2.0 cm portion of cuttings was dipped in various concentrations of test solutions for 24 h at $22^{\circ} \mathrm{C}$. The treatments consisted of Indole - 3 - butyric acid, IBA 100, 250 \& $500 \mu \mathrm{M}$; HiMedia Laboratories Pvt. Ltd, Mumbai) and Bavistin ( $0.1,0.5,1.0 \%$; a systemic fungicide; containing 50\% Carbendazim, a. i.; from BASF India Ltd., Mumbai, India); one untreated set served as control. The IBA was dissolved in $1.5 \%$ ( $\mathrm{v} / \mathrm{v}$ ) aqueous ethanol; control cuttings were dipped in aqueous ethanol ( $1.5 \%, \mathrm{v} / \mathrm{v}$ ). In each treatment only 24 cuttings were applied.
Following treatment, cuttings were planted vertically in polythene bags $(16.0 \mathrm{~cm} \mathrm{~h} \mathrm{x} 8.0 \mathrm{~cm}$ dia; one cutting per bag) containing sieved rooting mixture (equal parts of sand, soil and farmyard manure) and placed in following microclimatic conditions.
2.1. Polyhouse: A small polyhouse of 20 ft length x 9.0 ft width and 7.0 ft height (in the middle) was erected with the help of bamboo poles and covered with a thin semi-transparent polyethylene sheet (thickness: $162.5 \mu \mathrm{M}$, UV stabilized) from all the sides. The door could be opened at the front end to access the polyhouse.
2.2 Polypit: A pit dug in the ground (size 10.0 ft L x 6.0 ft W x 3.0 ft D ), covered on the top with a semi ï transparent polyethylene sheet (thickness: $162.5 \mu \mathrm{M}$, UV stabilized) supported by a bamboo frame. On one side, a small mud wall (about 30 cm high from the ground level, sloping on the two sides) was raised. The polyethylene sheet was sealed on higher side with mud, leaving three sides


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the open were dried up after four months of planting, therefore, details data of both the conditions are not presented. The first root initiation was observed at six months in majority of the cuttings planted inside polypit (average temperature and relative humidity ( RH ) during experiment period; $25-30^{\circ} \mathrm{C}$, with 60 to $80 \%$ ). The cuttings under control sets failed to root in both male \& female even planted under polypit condition. LSD ( $\mathrm{P}=0.05$ ) indicates significant differences in all the attributes measured for different treatments. IBA $(500 \mu \mathrm{M})$ and Bavistin (1.0\%) were found very effective for root formation. A maximum $50.0 \%$ rooting success was achieved in male cuttings (Table 1 and Figure 1C) with the application of $500 \mu \mathrm{M}$ IBA treatment on an average of 6.71 roots per cuttings and average root length of 3.27 cm , while Bavistin $1.0 \%$ treatment produce maximum rooting success $41.66 \%$ (with an average of 4.83 roots per cuttings and average root length 3.19; Table 1) in male cuttings and $58.33 \%$ (with an average of 4.66 roots per cuttings and average root length 4.0 cm ) in female cuttings of G. biloba (Table 2 and Figure 1D). Other concentration tried $100 \mu \mathrm{M}$ IBA was found to be slightly effective with rooting success of $33.33 \%$ in female cuttings with an average of 5.0 roots per cuttings and average root length of 3.25 cm . In male cuttings average length of longest root 7.5 cm and 7.32 cm while in female cuttings 6.83 cm and 6.72 cm was recorded with $500 \mu \mathrm{M}$ IBA and $1.0 \%$ Bavistin treatment, respectively. The maximum root diameter was recorded 1.26 mm for male (Table 1) and 1.53 mm for female cuttings (Table 2).

## 3. Results

Rooting ability of male and female branch cuttings of G.biloba was examined inside polyhouse, polypit and in the open conditions. Cuttings planted in all the conditions exhibited $70-80 \%$ bud
plastering (even with mud) was carried out. As a routine polyethylene covers from the top of the polypit were partially opened during the daytime after midday for a few hours (see Vyas et al. 1999 for details about its function and benefit).
2.3 Open: Open field space without any cover or modifications.

Observations were taken six months after treatment and planting for estimation of percent rooting, number of roots formed per cutting, root diameter and the length of individual roots. Only those cuttings with one or more clearly visible root initials ( $\geq 2 \mathrm{~mm}$ ) and/ or roots were classified as having rooted (Nandi et. al. 1996). Following this the well rooted cuttings were again planted in polybags and moved into the shade house, receiving $50 \%$ sunlight for hardening and the plants were watered periodically (Fig. 1E).
Least significant differences (LSD), standard error (SE) were calculated for comparison among the treatments following the methods as described by (Snedecor and Cochran, 1967). Analysis of variance (ANOVA) was performed using the Microsoft excel programme. sprouting (Figure 1A and B). Based on the random observations, cuttings planted in polyhouse and in

| Table 1. Effect of IB |
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| condition |

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Rooting | No. of |
| :---: |
| roots/ |
| cutting |
| $($ mean $\pm$ SE $)$ |

| Avg. Root length | Length of longest | Avg. root diameter |
| :---: | :---: | :---: |
| (mean $\pm$ SE, cm) | root (mean $\pm$ SE, | (mean $\pm$ SE, mm) |

Treatments

| IBA $(100 \mu \mathrm{M})$ | 20.83 | $3.77 \pm 0.88$ | $2.61 \pm 0.57$ | $5.0 \pm 0.73$ | $1.22 \pm 0.26$ |
| :--- | :---: | :---: | :---: | :---: | :---: |
| IBA $(250 \mu \mathrm{M})$ | 25.00 | $2.5 \pm 0.35$ | $3.48 \pm 1.03$ | $5.35 \pm 1.52$ | $0.94 \pm 0.21$ |
| IBA $(500(\mu \mathrm{M})$ | 50.00 | $6.71 \pm 1.30$ | $3.27 \pm 0.36$ | $7.5 \pm 1.02$ | $1.26 \pm 0.06$ |
| Bavistin $(0.1 \%)$ | - | - | - | - | - |
| Bavistin $(0.5 \%)$ | 29.0 | $5.28 \pm 0.37$ | $2.90 \pm 0.39$ | $5.41 \pm 0.33$ | $1.04 \pm 0.15$ |
| Bavistin $(1.0 \%)$ | 41.66 | $4.83 \pm 0.60$ | $3.19 \pm 0.41$ | $7.32 \pm 1.25$ | $1.08 \pm 0.08$ |
| LSD (p=0.05) | $\mathbf{3 0 . 6 7}$ | $\mathbf{4 . 1 8}$ | $\mathbf{2 . 0 8}$ | $\mathbf{4 . 6 3}$ | $\mathbf{0 . 7 3}$ |

$\mathrm{SE}=$ Standard error of mean. All values are an average of 24 cuttings.
A dash (-) indicates cuttings did not root.

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| Click Here to upgrade to Unlimited Pages and Expanded |  |  |  |
| Source of variation | DF | Mean Square | F-ratio |
| Treatments | 5 | 112.6579 | $2.194888^{\text {ns }}$ |
| Parameters | 4 | 742.7473 | 14.47077** |
| Error | 20 | 51.32741 |  |

**Level of significance at 0.05 ; ns: not significant
Table 2. Effect of IBA and Bavistin on rooting response of G. biloba female cuttings inside the polypit

| Treatments | $\begin{gathered} \% \\ \text { Rooting } \end{gathered}$ | No. of roots/ cutting (mean $\pm$ SE) | Avg. Root length (mean $\pm$ SE, cm) | $\begin{gathered} \text { Length of } \\ \text { longest root } \\ (\text { mean } \pm \text { SE, } \mathrm{cm}) \end{gathered}$ | Avg. root diameter (mean $\pm$ SE, mm) |
| :---: | :---: | :---: | :---: | :---: | :---: |
| IBA ( $100 \mu \mathrm{M}$ ) | 33.33 | $5.0 \pm 2.0$ | $3.25 \pm 0.76$ | $4.96 \pm 1.56$ | $1.46 \pm 0.12$ |
| IBA ( $250 \mu \mathrm{M}$ ) | 20.83 | $2.8 \pm 0.86$ | $3.77 \pm 1.13$ | $4.96 \pm 1.56$ | $1.46 \pm 0.12$ |
| IBA (500 ( $\mu \mathrm{M}$ ) | 12.5 | $4.0 \pm 1.52$ | $5.14 \pm 1.13$ | $6.83 \pm 1.96$ | $1.35 \pm 0.13$ |
| Bavistin (0.1\%) | 12.5 | $4.33 \pm 1.85$ | $3.33 \pm 0.63$ | $4.83 \pm 1.58$ | $1.12 \pm 0.13$ |
| Bavistin (0.5\%) | 20.83 | $1.5 \pm 0.35$ | $1.56 \pm 0.23$ | $2.0 \pm 0.0$ | $1.34 \pm 0.22$ |
| Bavistin (1.0\%) | 58.33 | $4.66 \pm 1.33$ | $4.0 \pm 0.58$ | $6.72 \pm 0.84$ | $1.53 \pm 0.11$ |
| LSD ( $\mathrm{p}=0.05$ ) | 30.67 | 2.35 | 1.83 | 2.94 | 0.24 |

$\mathrm{SE}=$ Standard error of mean. All values are an average of 24 cuttings.
ANOVA Summary Table

| Source of Variation | DF | Mean Square | F-Ratio |
| :--- | :---: | :---: | :---: |
| Treatments | 5 | 72.14717 | $1.216312^{\text {ns }}$ |
| Parameters | 4 | 643.7649 | $10.85308^{* *}$ |
| Error | 20 | 59.31633 |  |

**Level of significance at 0.05 ; ns: not significant.


Figure 1. Propagation of Ginkgo biloba: (A) A mature tree of G. biloba, (B) A sprouted male and female cuttings, (C\&D) well rooted male and
female cuttings and (E) Cuttings raised plants kept inside shade house for hardening.

## 4. Discussion

The stimulation of adventitious root formation in stem cuttings with auxins and commercial rooting mixtures is well known in many species those are difficult-to-root (Morsink and Smith, 1974, Blazich, 1998, Nandi et al. 1996, Purohit, 2002). The effect of growth hormone and fungicide on adventitious root formation in semi - hardwood cuttings of G.biloba in different microclimatic conditions, was examined for the first time. Although there are several studies available on the propagation of gymnosperms and angiosperms trees using stem cuttings (Nandi et al. 1996, 1997, 2002, Tamta et al. 2000, Purohit et al. 2005). The rooting efficiency observed in G. biloba showed satisfactory result within six months of planting inside the polypit conditions whereas reports available reveals that cuttings taking two years to root in natural conditions (Annonymous, 1999),

in the present study the application of IBA $(500 \mu \mathrm{M})$ significantly improved the rooting up to $50.0 \%$ in male cuttings whereas the application of Bavistin ( $1.0 \%$ ) was found more effective in both the cuttings (male and female) with highest rooting response was observed in female cuttings (58.33\%). However, on the other hand, the application of growth regulators (catechin and gallic acid) produced $53.3 \%$ and $56.7 \%$ rooting respectively, in the semi hard woodcuttings of $G$. biloba (Gopichand et al. 2006). Besides, the applications of different phenolic compounds were applied to assess the rooting response in $G$. biloba (Prakash et al. 2002) and they reported that the combination of IBA ( $500 \mathrm{mg} / \mathrm{l}$ ) and catechin (5 $\mathrm{mg} / \mathrm{l}$ ) enhanced the rooting upto $96.0 \%$.

IBA and Bavistin have also been reported to be more effective in inducing rooting in stem cutting of Taxus baccata (Nandi et al. 1996, 1997) and Cedrus deodara (Nandi et al. 2002). The rooting percentage displayed a positive trend with increasing concentrations of chemicals in this study. Applications of auxins enhanced rooting and root quality in many tree species (Hartman and Kester, 1983). The application of IBA may have an indirect influence by enhancing the speed of translocation and movement of sugar to the base of cuttings and consequently stimulate rooting (Haissig, 1974). Treatments with a systematic fungicide, Bavistin were also found effective in this study, however, the mechanism of stimulation of rooting by Bavistin is not clear. It may be related to auxin ï like activity of benomyl, which is known to be converted into carbendazim in water and in contact with plants (Thurston et al. 1979). Application of $500 \mu \mathrm{M}$ IBA and $1.0 \%$ Bavistin also enhanced the number of roots developed on each rooted cuttings as compared with another concentrations applied. This may have an advantage by enhancing good anchorage when planted in the field. Besides the effect of IBA and Bavistin, the diameter of cuttings may also have influenced the root formation on the cuttings whereas most of the thin cuttings either dried up or could not develop root even after six months.

In the present study rooting of stem cuttings was carried out in the months of January to June because of the deciduous nature of tree, best sprouting of dormant buds, active growth season which has been reported to favor rooting of cuttings (Loach, 1988). High humidity environments created by means of mist systems or plastic covers are commonly used in vegetative propagation experiments to reduce the

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risk of water stress (Hartmann et al.1990). Generally, spring season when the fresh flush starts is considered to be the best period for rooting.

## 5. Conclusions

The results of this study are important and low natural regeneration as reported in this species can be supplemented with clonally propagated plants raised through rooting of stem cuttings. Moreover, it is important to note that the cuttings were taken from mature trees from different sources; further trials in different seasons may result in better rooting efficiency and genetically specificdifferences. Due to poor economic condition and tough terrain in the mountains the construction of Green houses are difficult, therefore, a low cost polypit (poor man growth chamber) can meet out the requirement of rooting of cuttings and raising maximum number of plants. Keeping in view the present status, importance and conservation value of G. biloba a multi-faceted efforts is required while involving local communities, scientific institutions and NGOs for its nursery raising and afforestation programmes. Further, well-rooted plants could be obtained within a short time; the method is also inexpensive and easy to perform. It is hoped that it will be acceptable to the involved in the forestry sector.

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