

Insect and fungal pests of some mushrooms collected from university of Ibadan, Nigeria campusJonathan S.G.¹, Popoola K.O.K.², Olawuyi OJ¹, Ajiboye M.¹ and Oyelakan A. O.¹¹Department of Botany and Microbiology University of Ibadan, Ibadan, Nigeria²Department of Zoology, University of Ibadan, Ibadan, Nigeria.sg.jonathan@mail.ui.edu.ng

Abstract: Ten mushrooms species collected within the premises of University of Ibadan were examined for infestation of various insect and fungal pests. Insects belonging to the orders; Coleoptera, Hymenoptera, Diptera, and Collembolla were encountered both at the larval and adult stages of life on the collected mushroom samples. Infestation by the order Coleoptera (adult beetle) on *Pleurotus squar-rosulus* was found to be higher in incidence, with a total number of 17 species which were found at the adult stage of life; but the larva stage were found on *Lycoperdon gigantum*. Fungal species identified to be *Aspergillus niger*, *Aspergillus terreus*, *Fusarium redolens*, *Trichoderma viride*, *Rhizopus stolonifer* and *Mucor piriformis* were found to be associated with several species of mushrooms.

[Jonathan S.G., Popoola K.O.K., Olawuyi OJ, Ajiboye M. and Oyelakan A. O. **Insect and fungal pests of some mushrooms collected from university of Ibadan, Nigeria campus.** Nature and Science 2012; 10(9): 142-147]. (ISSN: 1545-0740). <http://www.sciencepub.net>. 20

Keywords: Mushrooms, fungal pathogens, insects, collection, pollution

1. Introduction

Morphologically, mushrooms have a fruiting body which can be easily distinguished by the sporocarps. A typical mushroom is made up of the pileus or cap; it is an expanded portion which may be thick, fleshy, membranous and also with varied shape (Zoberi, 1973; Jonathan and Adeoyo, 2011a). The lamellae or gill which is leaf-like radiating from the edge inward towards the stem and the stipe or stalk supporting the pileus. (Atkin, 1982, Jonathan, 2002). Mushrooms are the richest source of vegetable proteins. They contain 31-40% of protein. Mushrooms contain minerals like calcium, potassium, sodium, phosphorus and vitamins like B, C, D and K. mushroom contains niacin which is ten times higher than other vegetables (Jonathan *et al.*, 2012). The fruit bodies of mushrooms are used to produce suede-like material from which hand bags, hats, clothing, and picture frames are made. (Chang and Hayes 1978).

Mushrooms have very less calories and contain approximately 80 to 90 percent water (Aina *et al.*, 2012). At the same time, they have low sodium, carbohydrate and fat content and high fibre content. This is the reason why mushrooms are considered good for those aiming to loose weight. Mushrooms are valuable health foods that is low in calories, high in vegetable proteins chitin iron zinc fibre essential amino acids, vitamins, and minerals, such as copper that help the body to produce red blood cells (Esminger and Esminger 1986, Jonathan *et al.*, 2006; Aina *et al.*, 2012).

Mushrooms are excellent source of potassium. In fact, it is said that there is more potassium in a mushrooms than in banana. Since potassium helps lower blood pressure and diminish the risk of stroke, mushrooms are recommended to people suffering from hypertension (Chang *et al.*, 1989; Gbolagade, 2005). Mushrooms are rich in copper, a mineral that has cardio-protective properties. A single serving of mushrooms is said to provide about 20 to 40 percent of the daily needs of copper. They are excellent source of selenium, an antioxidant that works with vitamin E to protect cells from the damaging effects of free radicals. Researchers have suggested that white button mushrooms could reduce the risk of breast and prostate cancer. In fact, extract of white button mushrooms has been found to help in diminishing cell proliferation as well as tumour size. It has been found that mushroom extract helps stop migraine headaches and is beneficial for people suffering from mental illnesses, like obsessive-compulsive disorder (Jonathan, 2002; Aina *et al.*, 2012). Oyster mushrooms are said to be useful in strengthening of veins and relaxation of the tendons.

Despite nutritional and medicinal importance of mushrooms, they are being face with many pests and diseases. The various insects pests associated with mushrooms include; flies such as sciarids, phorids and cecids (Ajayi and Jonathan, 2004; Fasidi *et al.*, 2008). The flies belong to the order Diptera. Sciarid flies also known as fungus gnats belong to the family: Sciaridae and Species include; *Sciara multiselta*, *Sciara agaris*. Cecid flies also known as gall midges belong to the family: *Cecidomyidae* and

Species include; *Mycophila speyeri*, *Mycophila borresi*. Phorid flies belong to the family: *Phoridae* and Species include *Megaselia nigra*, *Megaselia halterata*.

Mites which are found in straw and manure include; small mushroom mites (*Tarsonemus sp*), straw or hay mites (*Tyrophagus sp*), Red pepper mites/pygmy mites (*Pygmephorus sp*). Eelworms or nematodes, they are tiny and transparent, they include the parasitic eelworms which are directly harmful such as; *Composticola*, *Ditylenchus muceliophagus*, and also the saprophytic nematodes which are indirectly harmful such as the Rhabdit types. The springtails which are also tiny insects include species such as *Isotoma simplex*, *Lepidocryptus cyaneus*. (Keil, 1996). Fungal diseases of cultivated mushrooms include; Dry bubble disease caused by *Verticillium fungicola*, wet bubble disease caused by *Mycogone perniciosa*, Cobweb or Dactylium mildew caused by *Cladobotryum dendroides* (*Hypomyces rosellus*), Green mould caused by *Trichoderma*. (Gbolagade, 2005, Fasidi *et al.*, 2008). There are certain abnormalities that occur in mushrooms and these disorders have several abiotic origins. Such abnormalities include; formation of stroma, formation of scales or crocodile skins, changes in the colour of fruit bodies, outgrowth on mushroom cap, long stipe, small cap on a normal stipe, rosecomb and scaling. (Singh *et al.*, 1991; Ajayi and Jonathan, 2004; Gbolagade, 2006). The objectives of this research work were to Identify various insect and fungal pests found on wild edible mushrooms and their features of damage and suggest possible control measure for the insect and fungal pests of mushrooms.

2. Materials and Methods

2.1 Study area

This study was conducted at the University of Ibadan, Oyo state. Ibadan is located in the South western Nigeria approximately between Latitude N 7° 26' Longitude E 3° 53' and an Altitude of 190m. The city ranges in elevation from 150m in the valley area to 275m above sea level. Ibadan has a tropical wet and dry climate with mean monthly temperatures fluctuating between 23° C to 30° C and humidity is usually from 55% to 75%.

2.2 Mushroom collection

The sample collection site for this research work was the University of Ibadan premises including Ibadan University Botanical Gardens. Between the month of April and August 2011. Survey trips and inventory of mushrooms in these areas were taken at seven days intervals. Ten species of mushrooms were

collected from the sample areas and each of the specie was replicated ten times. Collections were made in the morning. Mushrooms were collected using a shovel for obtaining part of the substratum (wood) on which mushrooms were growing, following the procedure of Jonathan and Adeoyo (2011b). They were identified using the standard procedures of Zoberi (1973).

2.3 Insect collection

Insect pests were removed from the mushroom samples by hand picking method. Insects were picked from each species of mushrooms at the point of collection and kept in specimen bottles. After collection, mushrooms were brought to the laboratory and part of the sporophores were carefully opened up using a dissecting knife in order to bring out the insects that had bored into the mushroom tissues. Pests were brought out and placed in labelled specimen bottles; insects were then preserved in 4% formalin. (Kim and Hwang, 1996).

2.3 Identification of insects

They were identified using the procedures of Kim and Hwang (1996). Accuracy of identification were carried out using the method and Bartlett. (1996). They were authenticated by Dr K.O.K Poopoola an Entomologist in the Department of Zoology, University of Ibadan. The identified arthropod species were stored in the Entomology laboratory, Department of Zoology, University of Ibadan, Ibadan, Nigeria for reference purpose.

2.4. Fungal isolation and characterization

Fungal infected mushrooms were collected and brought to the laboratory for isolation. Excised portions of the infected portion of *Pleurotus squarrosulus*, *Pleurotus pulmonarius* and *Pleurotus tuber-regium* were plated using potato dextrose agar (Oxoid). Streptomycin sulphate (0.05g/1000cm⁻³) was added to prevent bacterial contamination (Jonathan and Fsid, 2001). The isolates were plated in triplicates and incubated at room temperature (25 ± 2°C) for 7 days. At the end of the incubation period, the plates were observed for fungal growth and different colonies were sub-cultured on fresh plates of potato dextrose agar. Wet mount was done on grease free slides using 0.1% lactophenol cotton blue and were observed under the microscope (Domsh *et al.*, 1980). Cultural features observed on isolated fungi and characterization were carried out using the descriptions of Alexopolous (1996).

3. Results and Discussion

Results from preliminary studies revealed that the species of mushrooms (*Pleurotus squarrosulus*, *Volvariella esculenta*, *Termitomyces robustus*, *Pleurotus tuber-regium*, *Coprinus commatus*, *Lycoperdon giganteum*, *Boletus edulis*, *Macrolepiota*

sp., *Agaricus campestris* and *Psathyrella hydrophila*) were infested with various arthropod pests. *Boletus edulis* was found with the highest number, having a percentage composition of 10.1% and *Polyporus melanopus* was found with the lowest number and a percentage composition of 1.4%.

Table 1. List of Mushrooms by Families on University of Ibadan Campus.

Families	Mushrooms	Numbers	%Composition
<i>Agaricaceae</i>	<i>Agaricus campestris</i>	12	8.1
	<i>Pleurotus tuber-regium</i>	10	6.8
	<i>Pholiota terrestris</i>	5	3.4
	<i>Pleurotus squarrosulus</i>	11	7.4
<i>Polyporaceae</i>	<i>Polyporus melanopus</i>	2	1.4
	<i>Ganoderma lucidium</i>	7	4.7
<i>Coprinaceae</i>	<i>Coprinus commatus</i>	8	5.4
	<i>Psathyrella hydrophila</i>	13	8.8
<i>Amanitaceae</i>	<i>Amanita verna</i>	5	3.4
<i>Lycoperdaceae</i>	<i>Lycoperdon germinatum</i>	5	3.4
<i>Clavariaceae</i>	<i>Clavaria vermiculuris</i>	3	2.0
<i>Boletaceae</i>	<i>Boletus edulis</i>	15	10.1
<i>Tricholomalaceae</i>	<i>Tricholoma aurantium</i>	6	4.1
	<i>Chlorophyllum molybdites</i>	10	6.8
	<i>Macrolepiota sp</i>	7	4.7
	<i>Volvariella esculenta</i>	10	6.8
	<i>Termitomyces robustus</i>	9	6.1
	<i>Cylocybe dilate</i>	10	6.8

Table 2. Mean number of Edible mushrooms from sample areas in the University of Ibadan

Sample Areas	Numbers	Means
Botanical garden		
<i>Agaricus campestris</i>	12	1.2
<i>Volvariella esculenta</i>	10	1.0
<i>Boletus edulis</i>	15	1.5
Nursery (Botany dept.)		
<i>Termitomyces robustus</i>	9	0.9
<i>Macrolepiota sp</i>	7	0.7
<i>Pleurotus tuber-regium</i>	10	1.0
Jaja		
<i>Coprinus commatus</i>	8	0.8
<i>Lycoperdon germinatum</i>	5	0.5
Balewa road		
<i>Pleurotus squarrosulus</i>	11	1.1
<i>Psathyrella hydrophila</i>	13	1.3

Table 3. Insect Pests Encountered on Mushrooms, order, common name, life stage and number collected.

Sample areas	Mushroom	Insect order	Common name	Life stage	Number collected
Botanical garden	<i>Agaricus campestris</i> <i>Volvariella esculenta</i> <i>Boletus edulis</i>	Coleoptera	Beetle	Larval	4
				Adult	11
Nursery	<i>Termitomyces robustus</i> <i>Macrolepiota sp</i> <i>Pleurotus tuberigium</i>	Coleoptera	Beetle	Larva	4
		Coleoptera	Beetle	Adult	1
		Hymenoptera	Ant	Adult	1
		Collembola	Springtail	Adult	5
		Diptera	True fly	Adult	1
Jaja	<i>Coprinus commatus</i> <i>Lycoperdon germinatum</i>	Hymenoptera	Ant	Adult	12
		Coleoptera	Beetle	Larva	1
Balewa	<i>Pleurotus squarrosulus</i> <i>Psathyrella hydrophila</i>	Coleoptera	Adult Beetle	Larval	17
				Adult	2

Table 4. Distinguishing features of pests and damages done on different part of mushroom.

Order	Distinguishing features	Damages done on Mushroom
Collembola	Moderate sized, elongated body which is silvery in colour	Eats up the edges of the pileus and lamella, ingestion of mycelium
Diptera	Presence of a shiny black head capsule usually elongated and vermiform in shape	Feeds on lamellae, loss of mycelium and pileus, reduction of stipe.
Hymenoptera	Comparatively large, with a pair of antennae of 4-13 segments, abdomen distinctly constructed at the base, No wings.	Loss of pileus, lamella and reduction of stipe.
Coleoptera	Elongated body with dark elliptical body, covered with setae	Bore hole into the stipe of the mushrooms

Table 5- Identified isolates from infected mushroom samples

Isolate code	Isolate	Surface colour	Reverse colour
SQ1	<i>Aspergillus niger</i>	Blackish brown	Creamish yellow
FL1	<i>Aspergillus niger</i>	Blackish brown	Creamish yellow
SAJ1	<i>Mucor piriformis</i>	Black	Milky
SQ2	<i>Fusarium redolens</i>	Orange	Creamish
FL2	<i>Aspergillus terreus</i>	Cinnamon(brownish)	Yellowish brown
SAJ2	<i>Rhizopus stolonifer</i>	Reddish brown	Milky
FL3	<i>Trichoderma viride</i>	Green	Creamy

Table 6. Morphology and cultural characteristics of fungal isolates obtained from infected mushroom samples

Isolates	Mycelia colour	Reverse colour	Growth pattern	Microscopic examination
<i>Aspergillus terreus</i>	Cinnamon (Brownish)	Yellowish brown	Rapid	Conidial head showing metulae and phialides
<i>Aspergillus niger</i>	Blackish brown	Creamish yellow	Rapid	Conidial head with metulae and phialides
<i>Mucor piriformis</i>	Whitish mycelia with blackish sporangia	Milky	Grows very fast	Sporangiophore tips with columellae
<i>Rhizopus stolonifer</i>	Reddish brown	Milky	Rapid	straight dark brown sporangiophore with collumellae
<i>Fusarium redolens</i>	Orange	Creamish	Rapid	Macro conidia formed with chlamydospores arising in the mycelium and conidia
<i>Trichoderma viride</i>	Green	Creamy	rapid	Conidiophore pyramidally branched, phialides slender and irregularly bent.
<i>Aspergillus niger</i>	Blackish brown	Creamish yellow	Rapid	Conidial head with metulae and phialides

Insect pests such as ants, beetles and true flies were encountered on the mushrooms, they were found at the larval and adult stages. Insect orders

such as Coleoptera, Hymenoptera, Collembola and Diptera were present. Infestation by Coleoptera (Adult beetle) in *Pleurotus squarrosulus* was found

to be high, with a total number of 17 which were found at the adult stage of life and the number found in *Lycoperdon gigantum* was low also found at its larval stage. Their population was observed to be high in the lamellae of *Pleurotus squarrosulus* due to their feeding habits and protection derived from the lamell. The distinguishing features of the Coleopterans were also recorded; they have elongated body with dark elliptical body which are covered with setae. Their features of damage were also observed, they bore holes into the stipe of the mushroom.

In this study, beetle larvae also caused damages to the mushrooms, this type of damage was found to be related to those reported by Jonathan (2008). This work also showed that they were responsible for mycelium damage by feeding on the hypha and also transmits fungal infection which can be related to the report of (Fasidi *et al.*, 2008). The Collembola (Springtails) were found present at the Adult stage, they are moderately sized, with elongated body which is silvery in colour. They eat up the edges of the pileus, lamella and also ingest the mycelium. The Diptera (True fly) were also found on *Termitomyces robustus* at the Adult stage, they possess a shiny black head capsule usually elongated and vermiform in shape, they damage the mushroom by feeding on the lamella, they cause loss of mycelium and pileus, they also reduce the stipe. The Hymenoptera (Ants) were present on *Pleurotus tuber-regium* and *Coprinus commatus* at the Adult stages of life. The Ants having a blue-black with brown stripes, the abdomen distinctly constricted at the base, they also lack wings. The damages caused includes; loss of pileus, lamella and reduction of stipe. Since it is generally known that ants eat almost anything sugary and the major constituent of mushroom is sugar-alcohol mannitol. This justifies their presence in mushrooms.

However, this study provides useful information on how the various Arthropod pests have caused damage to these mushrooms. Also, insects have been found to infest mushrooms for them to be able to complete their life cycle and, in this process, they reduce the growth rate of mushroom; they nibble holes on different parts of the mushroom thereby reducing the market value of the mushroom. As described by Cantelo (1980), reducing fly numbers without using insecticides require a good understanding of fly biology and behaviour, therefore non-toxic chemicals such as Diflubenzuron may be applied in order to arrest the development of insect larvae (Fasidi *et al.*, 2008)

Fungal isolates such as *Aspergillus niger*, *Fusarium redolens*, *Mucor piriformis*, *Aspergillus terreus*, *Rhizopus stolonifer* and *Trichoderma viride*

were isolated from the fungal infected mushrooms (*Pleurotus pulmonarius*, *Pleurotus tuber-regium* and *Pleurotus squarrosulus*). Similar fungal species were reported by Ajayi and Jonathan (2004). The morphology and cultural characteristics of fungal isolates obtained from infected mushroom samples were observed. Identification of each genus was based on morphological and cultural characteristics compared to compendium of soil fungi (Domsh *et al.*, 1980). They were further characterised using the standard descriptions of Alexopolous *et al.* (1996). Confirmed identification were carried out using illustrated manual of Singh *et al.*, (1991).

Aspergillus niger was a fast growing fungus which appeared dark brown at first and later turned black, with conidia heads which were globose and later spilled to conidia chain which were brownish and smooth. *Rhizopus stolonifer*, was reddish brown in colour had a rapid growth with straight dark brown sporangiophore and collumellae. *Mucor piriformis* which had whitish mycelia with blackish sporangia covered the plates after 48 hours, produced sporangiophore tips with collumellae. *Aspergillus terreus* had colonies with cinnamon (brownish) colour, the conidia was globose, conidia head showed metulae and phialides. *Fusarium redolens*, appeared orange in colour, formed macro conidia with chlamydo-spores arising in the mycelium and conidia. *Trichoderma viride*, the growth greenish in colour, and the growth was much after 48 hours. Microscopically, the conidiophores was pyramidally branched, phialides slender and irregularly bent.

Banmet and Hunter 1972, suggested that fungal diseases can be managed physically by steaming at 54.4°C for 15 minutes which will eliminate the disease from casing soil. Kim and Hwang (1996), suggested three methods of prevention of fungal diseases which includes; steam sterilization of mushroom beds, formaldehyde fumigation and fungicidal application. Jonathan (2008) suggested that fungal diseases could be best controlled by a complete careful farm management and hygiene, also recommended fungicides such .He also suggested application of benomyl and chlorothanil at a recommended dosage. It should be noted that these control measures could only be applied to cultivated or domesticated mushrooms. The diseases in wild mushrooms may be difficult for treatment and control unless if they grow together in a specific habitat.

Correspondence to:

Dr. S. G. Jonathan
Department of Botany and Microbiology,

University of Ibadan

E-mail: sg.jonathan@mail.uui.edu.ng

Tel: +2348164746758

References

1. Aina DA, Oloke JK, Jonathan SG and Olawuyi OJ 2012. Comparative assessment of mycelia biomass and exo-polysaccharide production in wild type and mutant strains of *Schizophyllum commune* grown in Submerged liquid medium. *Nature and Science* 10(10):82-89.
2. Ajayi E.J and Jonathan S.G 2004. Plant Pests Diseases: An approach to control methods. Jacob Ojo and Sons. 152 pp.
3. Alexopolous C.J, Mims C.W and Blackwell M. 1996. *Introductory Mycology*. 4th Edition, John Wiley, New York.
4. Banrnet H.L and Hunter B.B 1972. Illustrated Genera of Imperfect Fungi. Minneapolis Burgess Publishing Company. Minneapolis, MN. 241pp.
5. Cantelo W.W 1980. Control of mushroom flies without chemicals. *Mushroom news* 28 (4):9-17.
6. Chang S.T and W.A Hayes, 1978. The biology and cultivation of edible mushroom. Academic press; Inc, New York and London 81 9pp.
7. Chang, Shu-Ting; Phillip G. Miles (1989). Mushrooms: cultivation, nutritional value, medicinal effect, and Environmental Impact. CRC Press. pp.4-6.
8. Domsh K.H, Gam W, Anderson T.H, 1980 Compedium of soil fungi vol.1, institute of soil biology, federal agriculture research centre Braunschweig. Academic press London.
9. Esminger A.A and Esminger M.K. 1986. Food for health, a nutrition encyclopedia clovis califonia.
10. Fasidi I.O, Kadiri. M. Jonathan SG. Adenipekun C.O. Kuforiji C.O 2008. Cultivation on edible tropical mushrooms. Ibadan University press. 81pp.
11. Gbolagade J.S (2005). Bacteria associated with cultures of *Psathyrella atrombonata* (Pegler), and *Schizophyllum commune* (Fr.Ex.Fr), Nigerian edible mushrooms *Acta Phytopathologica. Et Entomologica Hungarica*, 40 : (2-3), 333-340.
12. Gbolagade, S.J. 2006. Bacteria associated with compost used for cultivation of Nigerian edible mushrooms *Pleurotus tuber-regium* (Fr.) Singer and *Lentinus squarrosulus* Berk. *African Journal of Biotechnology* 5: 338-342.
13. Jonathan, S.G and I.O Fasidi (2001). Effect of Carbon, nitrogen and mineral sources on growth of *Psathyrella atroumbonata*, Pegler. A Nigeria edible mushroom. *Food chemistry*, 72:479-48.
14. Jonathan S.G (2002). Studies on vegetative growth requirements and antimicrobial activities of higher fungi from Nigeria. Ph.D thesis University of Ibadan, Nigeria.
15. Jonathan SG (2008). Mushroom pests and diseases. In: Fasidi *et al*, 2008: Cultivation of Tropical mushrooms. University Press. Ibadan 81pp.
16. Jonathan SG and Adeoyo O.R 2011a. Collection, morphological characterization and nutrient profile of some wild mushrooms from Akoko, Ondo state, Nigeria. *Natural products*: 7(3):128-136.
17. Jonathan SG and Adeoyo O.R 2011b. Evaluation of ten wild Nigerian mushrooms for amylase and cellulose activities. *Mycobiology* 39(2):103-108
18. Jonathan SG, Okorie AN Garuba EO and Babayemi OJ (.2012). Bioconversion of sorghum stalk and rice straw into value added ruminant feed using *Pleurotus pulmonarius*. *Nature and Science*; 10(4):1016.
19. Keil, C. B. O. and G. R. Bartlett. 1996. Permethrin resistance in *Lycoriella mali* (Fitch) (Diptera: Sciaridae) on commercial mushroom farms. *Mushroom News* 44: 8-13.
20. Kim, K.C. & Hwang, C.Y. 1996. An investigation of insect pest on the mushroom (*Lentinus edodes*, *Pleurotus ostreatus*) in south region of Korea. *Korean Journal of Applied Entomology* 35 (1), 45-51.
21. Singh K, Frisvad J.C, Thraneus U, Mathew S.B 1991. An illustrated manual on the identification of some seed borne Aspergilli, Fusaria, Penicillia and their mycotoxins. Danish government institute of seed pathology for developing countries, Hellerup pp31-69.
22. Smith, S.E. and Read, D.J. (1997) Mycorrhizal Symbiosis, Academic Press.
23. Zoberi, M.H. 1973. Some edible mushrooms from Nigeria. *Nigerian Fields*. 38:81-90.

7/28/2012