Prevalence, Risk Factors and Major Bacterial Causes of Bovine Mastitis in West Arsi Zone of Oromia Region, Southern Ethiopia

Umer Seid, Tilahun Zenebe, Gizat Almaw, Abdela Edao, Haimanot Disassa, Tadele Kabet, Firmaye Gerbi and Girma Kebede

1Wollega University, School of Veterinary Medicine, P.O. Box 395, Nekemte, Ethiopia
2National Animal Health Diagnostic and Investigation Center, P.O.Box 04, Sebeta, Ethiopia
3Adami Tulu Agricultural Research Center, P.O. Box 35, Ziway, Ethiopia
Email: tilahun.zenebe@yahoo.com

Abstract: A cross-sectional study was conducted from October 2014 to April 2015 in Shashemene and Kofale, West Arsi zone of Oromia regional state with the objectives of determination of the prevalence of bovine mastitis, isolate the predominant bacterial agents involved in causing mastitis and identify associated risk factors. A total of 358 lactating cows (169 local and 189 cross) were examined for mastitis using clinical examination and California mastitis test (CMT). An overall prevalence of mastitis was recorded in the area 38% (136/358), of which 7.3% (26/358) were clinical and 30.7% (110/358) subclinical cases. Total animal examined in Shashemane was 165, out of these 35.8% (59/165) animals were positive for mastitis, from these animals, 8.5% (14/165) clinical and 27.3% (45/165) subclinical mastitis while 193 animals were examined from Kofale, out of these 39.9% (77/193) animals were positive for mastitis, from these animals, 6.2% (12/193) clinical and 33.7% (65/193) subclinical mastitis. Bacteriological methods were also employed to isolate the causative bacteria. About 83 bacterial isolates belonging to 7 species were identified from mastitic milk samples. The predominant isolated bacteria were Staphylococcus aureus (44.6%) followed by Streptococcus agalactiae (18.1%), Pseudomonas aeruginosa and Klebsiella pneumoniae were the least isolate which accounts 3.6% for each. There was no statically significant variation (P>0.05) between breeds, parity, age, but the prevalence of mastitis was found to be statistically significant among different hygiene of milking groups and lactation stages (p<0.05). The study also shows that mastitis is significant problem of dairy cows in the study area and the major isolated bacteria were contagious pathogens. The farmers should also be aware of the impact of the disease and practice hygienic milking, culling of chronic mastitis carriers and treating of clinically infected cows.

Key words: Bovine, Mastitis, Major Pathogens, Prevalence, West Arsi Zone

1. Introduction

Ethiopia, located in tropical region, is one of the most populous countries in Africa, having an estimated population of more than 80 million. The country is very much dependent on agriculture. Livestock represent a major national resource and form an integral part of the agricultural production system. Ethiopia has the largest cattle population in Africa with an estimated population of 56.71 million. Cow represents the biggest portion of cattle population of the country, around 20.7% of the total cattle heads are milking cows (CSA, 2014). However, milk production often does not satisfy the country’s requirements due to a multitude of factors. Mastitis is among the various factors contributing to reduced milk production (Biffa et al., 2005). Dairy enterprise is very gradual in countries of sub Saharan Africa like Ethiopia. In this region, the low local milk production is as result of many factors including low genetic potential for milk production of indigenous breeds, the extensive and low inputs husbandry practices under which they are reared and wide spread livestock diseases (Mohamed et al., 2004). Accordingly, few improved exotic breed animals that mostly limited in urban and peri-urban areas are yet not in position to satisfy the growing demand for milk of nation (Felleke and Geda, 2001). Conversely, low annual per capital consumption of milk in Ethiopia is 17-19 litters witnesses the gap between supply and basic demand of milk in urban areas (Vallezarate, 2000). Consequently, adequacy of the domestic milk production to cover the local demand has resulted in improving a considerable amount of dairy products. According to FAO (2003) in Ethiopia 42% of total cattle for private holdings are milking cows, however, milk production often doesn’t satisfy the countries milk requirement due to multitude factors. There are several types of diseases which potentially infect and affect the wellbeing of livestock population among which mastitis is the common and
costly disease causing loss in milk yield, treatment cost for dairy farmers and culling of animals at unacceptable age (Vaarst and Envoldsen, 1997). It is considered as the most complex disease because of its multi factorial causation (Nibret, 2009).

Mastitis is an inflammation of the mammary gland and udder tissue, and is a major endemic disease of dairy cattle that follows a number of factors including the cow, the pathogen and the environment (Radostits et al., 2007). It usually occurs as an immune response to bacterial invasion of the teat canal by variety of bacterial sources present on the farm, and can also occur as a result of chemical, mechanical and thermal injury to the udder. Mastitis as a disease has received little attention in Ethiopia, especially the subclinical form even though it is by far higher than clinical mastitis (Robertson, 1985). Efforts have only been concentrated on the treatment of clinical cases. In Ethiopia, the available information indicates that bovine mastitis is one of the most frequently encountered diseases of dairy cows (Stephen et al., 2001). It is a major and prevalent disease of dairy cows causing huge economic loss as a result of reduced milk yield, early culling of productive cows (Fekadu, 1995) and a serious problem in the dairy industry of Ethiopia (Mekonnen et al., 2005). The prevalence of clinical and sub clinical mastitis in different parts of Ethiopia is 1.2% and 21.5% respectively (Kassa et al., 1999; Lemma et al., 2001; Mungube, 2001; Kerro and Tareke, 2003; Werkineh et al., 2002).

In Ethiopia, mastitis has long been known (Biffa et al., 2005; Tamirat, 2007; Almaw et al., 2009; Bitew et al., 2010), however, the information on the magnitude, risk factors and causative agent of the disease is inadequate. Such information is important when designing appropriate strategies that would help to reduce its prevalence and effects (Biffa et al., 2005). Most studies in Ethiopia were carried out in Addis Ababa and its surrounding, which are not representative of other regions of the country (Almaw et al., 2009). In west Arsi zone, mastitis is not well considered. There is very few published material on the current status of mastitis in Shashename (Abera et al., 2012).

Hence this study was initiated with the objectives of:
- To estimate the prevalence of bovine mastitis in the selected districts of west Arsi zone,
- To assess the major risk factors associated with the occurrence of the bovine mastitis in the selected districts of west Arsi zone,
- To isolate and identify the major bacterial causative agents of bovine mastitis.

2. Materials and Methods

2.1. Study Area: The study was conducted in selected districts of West Arsi zone from October 2014 up to April 2015 in Kofale and Shashemene districts. Shashemene is located 250 km south of the capital Addis Ababa, and 25 km north of Awassa. The area lies within the Rift Valley, with altitudes ranging from 1700 to 2600 meters above sea level (m.a.s.l) and located at 7° 05’N to 7° 19’N and 38° 23’E to 38° 41’E. It receives an annual rainfall of 700–950 mm, and has an annual temperature range of 12–27°C. Out of the total area of 76,888 hectares, crop land accounts for 48,975 hectares, and the rest 7440, 5160, and 1320 hectares are forest land, grazing land and land for other purposes, respectively. The urban settlement accounts for 1733 hectares. The cattle population in the districts is 184,549 (SDARDO, 2010). While Kofale districts is located at 280 km south of Addis Ababa and located at 7° 19’N to 7° 40’N and 38° 30’E to 38° 53’E. Kofale is a highland, agro-pastoral area. Temperatures are moderate to hot. Some forests, including Arsi State forest, are found in Shashemene and Kofele districts. Rain fall is sufficient, which Kofale has erratic type of bimodal rainfall (KDARDO, 2010).

The human population of Shashemene districts is projected to be 100,454, directly living on agriculture and associated activities also by supplying its produce to the neighboring urban dwellers. While human population in Kofale districts is estimated around 178,950. The people of both districts belong to the Oromo ethnic community and others. Afan Oromo (the Oromo language) is the widely spoken language in the area (CSA, 2008).

2.2. Study Population: According to the recent census (CSA, 2014) Animal population of the west Arsi zone is 1,957,066 cattle, while 7.8% and 8.4% cattle population where found in Shashemene and Kofale, respectively. However, the study populations were all local and cross breeds of dairy cows that are managed under extensive, semi intensive and intensive farming system. The study was conducted on 358 local and cross breed from selected districtsin west Arsi zone of Oromia regional state namely; Kofale and Shashemene. Out of these animals, 165 were from Shashemene, 193 were from Kofale. Breed, age, parity, hygiene of udder, tick infestation of udder, lactation stage, floor, husbandry system, sequence of milking of cows were explanatory variables used to associate with prevalence rate.

2.3. Study Design: A cross-sectional study was conducted from October 2014 to April 2015 to estimate the prevalence of mastitis, isolate and identify major bacterial pathogens and to assess the association of some potential risk factors with
occurrence of mastitis in lactating cows in Shashemene and Kofale, west Arsi zone of Oromia regional state.

2.4. Sampling and Sample Size Determination: Purposive sampling technique was employed to selected two districts from 12 districts of the zone based on dairy cow population. The peasant association and lactating cows were also selected purposefully based on their accessible for transportation. The sample size was determined based on the formula given by Thrusfield (2007) considering 5% absolute precision, 95% confidence interval and from previous studies in the study area (Abera et al., 2012), with an expected prevalence of 37%. Therefore, the total sample size was 358 based on the given formula below.

\[
\frac{d^2}{n} = \frac{1.96^2 \cdot P_{exp} \cdot (1-P_{exp})}{d^2}
\]

Where, \(n\) = required sample size
\(P_{exp}\) = expected prevalence
\(d\) = desired absolute precision

2.5. Study Methodology

Based on the clinical inspection and CMT result cases were categorized as positive or negative and positive cases were again categorized as subclinical and clinical mastitis Quinn et al. (1999). Lactation stage, parity, sequence of milking, type of floor, husbandry system and age of study animals were obtained by interviewing the owner of the herd. Lactation stage was categorized as early (1 up to 3 month), mid (4 up to 6 month) and late (>7 months); age as young adult (3 up to 5 year), adult (6 up to 9 year) and old (9 year and above); parity as few (1 up to 3 calves), moderate (4 up to 6 calves), and many (above 6 calves).

2.5.1. Data collection: A questionnaires were used to collect data on the possible risk factors to the mastitis prevalence such as; breed, presence of lesion on skin of udder, and /or teats; parity number and hygiene of milking process as good and poor (that is based on cardinal signs of inflammation, symmetry, size and consistency of udder quarters (Radostits et al., 2007).

2.5.2. Physical examination of udder and milk: The udder of the study cows was examined visually and by Palpation for presence of clinical mastitis. During examination attention was paid to cardinal signs of inflammation, symmetry, size and consistency of udder quarters (Radostits et al., 2007).

2.5.3. Californian mastitis test: The California mastitis test (CMT) was conducted to diagnose the presence of subclinical mastitis and it was carried out according to the procedures given by Quinn et al. (1999). The udder of the cow to be tested was cleaned with warm water and antiseptics and was dried with clean towel. Then the first few drops were discarded from each quarter. Following that squirt of milk from each quarter of udder, milk samples were poured in to four shallow cups in the CMT paddle and equal amount of CMT reagent was added to each cup and gentle circular motion was applied to the mixture on the horizontal plane. Based on the thickness of the gel formed by CMT reagent and milk mixture, test results were scored as 0 (negative), 1 (weak positive), 2 (distinct positive) and 3 (strong positive). Milk samples with test result of CMT 1 to 3, were classified as evidence of subclinical mastitis (Radostits et al., 2007; Quinn et al., 1999).

2.5.4. Sample collection and bacteriological examination: Milk samples were collected after the skin of the udder was washed with tap water and dried with clean towel and the teat were swabbed with cotton soaked in 70% ethyl alcohol. The samples were collected in sterile universal bottles held nearly horizontal, after discarding the first few squirts of milk (Quinn et al., 1999). All sample bottles were labeled for cow identification and for the respective quarters. Milk samples were immediately transported to the microbiology laboratory of the National Animal Health and Investigation Center (NAHDIC) in ice packed cool box for bacteriological analysis. All milk samples were subjected to bacteriological examination according to Quinn et al. (1999) commencing on the date of collection.

2.5.5. Culture and isolation of bacteria: In the laboratory a loop full of the milk sample was streaked on blood agar base enriched with 7% sterile sheep blood and MacConkey agar and incubation was made at 24 to 48 hours. Then plates were examined for growth, colony morphology and hemolytic characteristics on blood agar. Subcultures were done to obtain pure isolates for further identification. Culture positive plates were identified according to gram stain reaction, colony morphology and catalase test. Identification of bacterial species was done according to Quinn et al. (1999) using standard methods. Oxidase test was done to distinguish Enterobacteriaceae from other gram negative bacteria. Enterobacteriaceae were subjected IMVIC test for species level identification.

2.6. Data Analysis and Management: Data collected during the study period were entered into Microsoft Excel 2007 spread. STATA 11 statistical software was used for the analysis. Logistic regression was used to see the association between different risk factors like: breed, age hygiene of milking, parity, lactation stage p <0.05 were considered as significant.

3. Results
3.1. Prevalence of Bovine Mastitis: From the total of 358 lactating cows examined, 38% (136/358) were positive for mastitis. Of these, 7.3% (26/358) sand 30.7% (110/358) were found to be positive for clinical mastitis and subclinical mastitis based on the clinical diagnosis and CMT, respectively. The study also considered mastitis at quarter level. An overall prevalence of quarter level was 34.5% (490/1422); from this the clinical form was 6.6% (94/1422) and the subclinical was 27.9% (396/1422). Out of the 1432 quarters examined, 10 had blind teats (Table1).

### Table 1: Prevalence of clinical and sub clinical mastitis at cow and quarter levels

<table>
<thead>
<tr>
<th>Cow level (n=358)</th>
<th>Quarter level (n=1422)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Types of mastitis</td>
<td>No. of positive</td>
</tr>
<tr>
<td>Clinical</td>
<td>26</td>
</tr>
<tr>
<td>Subclinical</td>
<td>110</td>
</tr>
<tr>
<td>Total</td>
<td>136</td>
</tr>
</tbody>
</table>

Total animal examined in Shashemene were 165, out of these 35.8% (59/165) animals were positive for mastitis, from these, 8.5% (14/165) clinical and 27.3% (45/165) subclinical mastitis while 193 animals were examined in Kofale, out of these 39.9% (77/193) animals were positive for mastitis, from these 6.2% (12/193) clinical and 33.7% (65/193) subclinical mastitis (Table 2).

### Table 2: Summary of prevalence of clinical and subclinical mastitis at cow level in study area

<table>
<thead>
<tr>
<th>Districts</th>
<th>No. of cow examined</th>
<th>Total No. of cows affected</th>
<th>Over all Prevalence (%)</th>
<th>Form of mastitis</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>Clinical</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>No. of positive</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>Subclinical</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>No. of positive</td>
</tr>
<tr>
<td>Shashemene</td>
<td>165</td>
<td>59</td>
<td>35.8</td>
<td>14</td>
</tr>
<tr>
<td>Kofale</td>
<td>193</td>
<td>77</td>
<td>39.9</td>
<td>12</td>
</tr>
<tr>
<td>Total</td>
<td>358</td>
<td>136</td>
<td>38</td>
<td>26</td>
</tr>
</tbody>
</table>

3.2. Result of bacterial isolation: Analysis of bacteriological examination of milk samples was made to identify the main etiological agents involved in the disease. The organisms were identified on the basis of their cultural, staining characteristics and biochemical reactions. Milk sample of 101 quarters, which were positive for CMT, cultured for microbiological examination in the study period, about 7 bacterial species and 83(82.2%) bacterial isolates. The bacterial isolation rate and their prevalence are shown on (Table3). The predominant isolated bacteria were *Staphylococcus aureus* with isolation rate of 44.6% followed by *Streptococcus agalactiae* with isolation rate of 18.1%, CNS (coaglase negatives *Staphylococcus* species) (that was the third predominant isolated with isolation rate of 16.9%). *Pseudomonas aeroginosa* and *Klebsiella pneumonia* were the least isolate which accounts for 3.6% each.

### Table 3: Bacterial species isolated from the study area

<table>
<thead>
<tr>
<th>Bacterial species</th>
<th>Total number of isolates</th>
<th>Prevalence (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td><em>Staphylococcus aureus</em></td>
<td>37</td>
<td>44.6</td>
</tr>
<tr>
<td>CNS</td>
<td>14</td>
<td>16.9</td>
</tr>
<tr>
<td><em>Streptococcus agalactiae</em></td>
<td>15</td>
<td>18.1</td>
</tr>
<tr>
<td>Other <em>Streptococcus</em> species</td>
<td>7</td>
<td>8.4</td>
</tr>
<tr>
<td><em>E.coli</em></td>
<td>4</td>
<td>4.8</td>
</tr>
<tr>
<td><em>Pseudomonas aeroginosa</em></td>
<td>3</td>
<td>3.6</td>
</tr>
<tr>
<td><em>Klebsiellapneumonia</em></td>
<td>3</td>
<td>3.6</td>
</tr>
<tr>
<td>Total</td>
<td>83</td>
<td>100</td>
</tr>
</tbody>
</table>

CNS = Coagulate Negative *Staphylococcus*

3.3. Risk factors: Among the different potential risk factors considered for a univariate logistic regression, all the risk factors like: breed, age, parity, tick infestation of udder, sequence of milking, floor, husbandry system, lactation stage and hygiene of udder were found statistically significant (P<0.05). However, all became insignificant, except milking hygiene and stage of lactation when tested with multivariate logistic regression (Table 4). Both stage of lactation and milking hygiene were found to be statistically...
significant (P<0.05) associated with the occurrence of mastitis. Mastitis prevalence was found to be higher in early lactation and lower in mid lactation stages (Table 4). Statistical analysis showed the existence of significant difference (P<0.05) between the occurrence of mastitis and lactation stage. The occurrence of mastitis was higher in poor milking hygiene and lower at good milking hygiene. Furthermore, Animals over 8 years old were more frequently affected with the disease and those younger than 5 years were rarely affected. Also the highest prevalence of mastitis was observed in animals with parity of more than 6, followed by 3-6 and 1-2 parity (as indicated Table 4).

Table 4: Analysis result of risk factors for the occurrence of bovine mastitis in the study area

<table>
<thead>
<tr>
<th>Risk factors</th>
<th>Category</th>
<th>N&lt;sub&gt;o&lt;/sub&gt; of cows examined</th>
<th>N&lt;sub&gt;o&lt;/sub&gt; of cows affected</th>
<th>Prevalence (%)</th>
<th>Crude Odds ratio (95% CI)</th>
<th>Adjusted Odds ratio (95% CI)</th>
<th>P- value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Breed</td>
<td>Local</td>
<td>169</td>
<td>41</td>
<td>24.3</td>
<td>0.39(0.1583-0.9442)</td>
<td>0.75(0.2475-2.2640)</td>
<td>0.608</td>
</tr>
<tr>
<td></td>
<td>Cross</td>
<td>189</td>
<td>95</td>
<td>50.3</td>
<td>0.20(0.0506-0.7960)</td>
<td>0.98(0.1842-5.2174)</td>
<td>0.982</td>
</tr>
<tr>
<td>Age</td>
<td>3-5 years</td>
<td>176</td>
<td>8</td>
<td>4.5</td>
<td>0.09(0.0239-0.2986)</td>
<td>0.98(0.1842-5.2174)</td>
<td>0.982</td>
</tr>
<tr>
<td></td>
<td>5-7 years</td>
<td>88</td>
<td>43</td>
<td>48.9</td>
<td>0.04(0.0053-0.3077)</td>
<td>0.18(0.0193-1.7363)</td>
<td>0.139</td>
</tr>
<tr>
<td></td>
<td>&gt;8 years</td>
<td>94</td>
<td>85</td>
<td>90.4</td>
<td>0.04(0.0193-1.3706)</td>
<td>0.32(0.3091-3.2693)</td>
<td>0.335</td>
</tr>
<tr>
<td>Parity</td>
<td>1-3 calves</td>
<td>122</td>
<td>15</td>
<td>12.3</td>
<td>-</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td></td>
<td>4-6 calves</td>
<td>124</td>
<td>36</td>
<td>29</td>
<td>0.16(0.0193-1.3706)</td>
<td>0.32(0.3091-3.2693)</td>
<td>0.335</td>
</tr>
<tr>
<td></td>
<td>&gt;6 calves</td>
<td>112</td>
<td>85</td>
<td>75.9</td>
<td>0.04(0.0053-0.3077)</td>
<td>0.18(0.0193-1.7363)</td>
<td>0.139</td>
</tr>
<tr>
<td>Lactation stage</td>
<td>1-3 month</td>
<td>174</td>
<td>65</td>
<td>37.4</td>
<td>-</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td></td>
<td>4-6 month</td>
<td>144</td>
<td>45</td>
<td>31.3</td>
<td>0.82(0.3159-2.1188)</td>
<td>1.23(0.3271-4.626)</td>
<td>0.759</td>
</tr>
<tr>
<td></td>
<td>&gt;7 month</td>
<td>40</td>
<td>26</td>
<td>65</td>
<td>0.22(0.0783-0.6080)</td>
<td>0.08(0.0074-0.8448)</td>
<td>0.036</td>
</tr>
<tr>
<td>Floor</td>
<td>Good concrete</td>
<td>166</td>
<td>12</td>
<td>7.2</td>
<td>-</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td></td>
<td>Muddy soily</td>
<td>148</td>
<td>100</td>
<td>67.6</td>
<td>0.08(0.0189-0.3619)</td>
<td>0.54(0.9674-2.9667)</td>
<td>0.475</td>
</tr>
<tr>
<td></td>
<td>Bad concrete</td>
<td>44</td>
<td>24</td>
<td>54.5</td>
<td>0.09(0.0178-0.5086)</td>
<td>0.39(0.0542-2.7958)</td>
<td>0.348</td>
</tr>
<tr>
<td>Husbandry system</td>
<td>Extensive</td>
<td>146</td>
<td>35</td>
<td>24.0</td>
<td>-</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td></td>
<td>Intensive</td>
<td>156</td>
<td>62</td>
<td>39.7</td>
<td>0.31(0.1107-0.8700)</td>
<td>0.46(0.0956-2.261)</td>
<td>0.343</td>
</tr>
<tr>
<td></td>
<td>Semi-intensive</td>
<td>56</td>
<td>39</td>
<td>69.6</td>
<td>0.36(0.1005-1.3013)</td>
<td>12.45(0.8666-178.9095)</td>
<td>0.064</td>
</tr>
<tr>
<td>Sequence of Milking of cows</td>
<td>1&lt;sup&gt;st&lt;/sup&gt;Health,last mastitis</td>
<td>169</td>
<td>6</td>
<td>3.6</td>
<td>-</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td></td>
<td>Arbitrary</td>
<td>189</td>
<td>140</td>
<td>74.1</td>
<td>0.13(0.0384-0.4428)</td>
<td>0.56(0.1095-2.8609)</td>
<td>0.486</td>
</tr>
<tr>
<td>Milking hygiene</td>
<td>Neither washed nor Disinfected</td>
<td>124</td>
<td>105</td>
<td>84.7</td>
<td>-</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td></td>
<td>only washed</td>
<td>60</td>
<td>20</td>
<td>33.3</td>
<td>2.85(0.9335-8.7279)</td>
<td>1.47(0.3861-5.6199)</td>
<td>0.571</td>
</tr>
<tr>
<td></td>
<td>Washed and dried with towel</td>
<td>174</td>
<td>11</td>
<td>6.3</td>
<td>35.27(4.6749-266.1197)</td>
<td>10.38(1.1406-94.5100)</td>
<td>0.038</td>
</tr>
<tr>
<td>Tick infestation of udder</td>
<td>Absent</td>
<td>189</td>
<td>13</td>
<td>6.9</td>
<td>-</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td></td>
<td>Present</td>
<td>169</td>
<td>123</td>
<td>72.8</td>
<td>0.15(0.0487-0.4285)</td>
<td>0.47(0.1073-2.0375)</td>
<td>0.311</td>
</tr>
</tbody>
</table>

4. Discussion
The study was carried out to determine the prevalence of bovine mastitis and to identify the major bacterial causes of mastitis in Shashemene and Kofale districts on 358 animals and the overall prevalence was 38%. This result agree with the findings of Abera et al. (2012), Nessru(1999), Workineh et al. (2002), Fekadu(1995),who reported 37.1% in Shashemene,
37.2% in urban and peri-urban dairy farms at Addis Ababa, 38.2% in Adami Tulu, 38.65% in Chaffa valley respectively. However, it is relatively lower than the reports of Mekibib et al. (2010), Lakew et al. (2009), Sori et al. (2005) and Mungube et al. (2004) who recorded 71.1% from Holeta, 64.6% from Assela, 52.8% from Sebeta and 46.6% from central highlands of Ethiopia, respectively. Mastitis is a complex disease and the difference in results might be due to differences in management system of the farm, the breeds of cattle considered, technical know-how of the investigators and the geographical locations of the studies.

The present study revealed with prevalence of subclinical mastitis (30.7%) and clinical mastitis (7.3%) which is comparable with the findings of Nessru (1999) and Bishi (1998), who reported 32.2% subclinical and 5% clinical in urban and peri-urban dairy farms at Addis Ababa, 30.2% subclinical and 5.5% clinical in urban and peri-urban in and around Addis Ababa, respectively. Prevalence of subclinical mastitis is higher than that of clinical mastitis in the present study which is in the agreement with several earlier reports from different parts of Ethiopia (Abera et al., 2010; Mekibib et al., 2010; Lakew et al., 2009; Almaw et al., 2008; Getahun et al., 2008; Biffa et al., 2005; Mungube et al., 2004; Kerro and Tareke, 2003; Workineh et al., 2002) and elsewhere in Africa (Kivaria et al., 2004). Since, environmental factors play significant role, the prevalence of subclinical mastitis varies in dairy animals (Radostits et al., 2007). Subclinical mastitis has been reported to be higher than clinical mastitis owing to the defense mechanism of the udder, which reduces the severity of the disease (Erskine, 2001). In most reports including the present study, clinical mastitis is far lower than subclinical mastitis (Sori et al., 2005; Biffa et al., 2005; Almaw et al., 2008; Lakew et al., 2009; Haftu et al., 2012). This could be attributed to little attention given to subclinical mastitis, as the infected animal shows no obvious symptoms and secrets apparently normal milk and farmers, especially small holders, are not well informed about invisible loss from subclinical mastitis. In Ethiopia, the subclinical forms of mastitis received little attention and efforts have been concentrated on the treatment of clinical cases (Almaw et al., 2008).

Quarter prevalence of mastitis 34.5% found in this study was comparable with the finding of Nessru et al. (1997) and Bachaya et al. (2011) in Pakistan, who reported the 37%, and 35.25%, but higher than the report made by Biffa et al. (2005) who reported quarter prevalence of 28.2%.

From the 101 CMT positive quarter milk samples, 83 (82.2%) were bacteriologically positive up on culturing, while 18 (17.8%) were bacteriologically negative, which is in line with the results of Aregaw (1992), who reported 18% bacteriologically negative samples, however; higher than the reports of Sori et al. (2005) who reported a prevalence of 9.82%. The failure to isolate bacteria from the CMT positive milk samples could be partly associated with spontaneous elimination of infection, low concentration of pathogen in milk, intermittent shedding of pathogen, intracellular location of pathogens and presence of inhibitory substance in milk (Radostits et al., 2000). It might also be due to some cases of delayed healing of infection from which organisms may have disappeared or reduced, while infiltration of leukocytes continued until complete healing (Sori et al., 2005).

In this study, most of the bacterial pathogens isolated from milk samples were Staphylococcus species, S. aureus (44.6%) comparable with the finding of Abera et al. (2010) who report of (42.1%). The relative high prevalence of S. aureus in the current study shows the absence of dry cow therapy and low culling rate of chronically infected animals practice in the study area. The bacteria usually establish chronic, subclinical infections and are shed in the milk, which serves as a source of infection for other healthy cows during the milking process. Streptococcus species were also found prevalent with 26.5% share of the total isolates: Streptococcus agalactiae 18.1% and other Streptococcus species 8.4%. This finding coincides with that of Hawari and Al-dabbas (2008), and Zerihun (1996), who reported 26.2% in Jordan, 27% in Stela Dairy Farm, Ethiopia, respectively. However, the finding is higher than the reports of Bitew et al. (2010) at Bahir Dar and its environ (13.9%) and Sori et al. (2005) in and around Sebeta (3.73%) and lower than the report of Atyabi et al. (2006) at farms around Tehran (33.54%). The predominance of these two bacterial species (Staphylococcus and Streptococcus) is due to frequent colonization of teats as they are commensals of the skin. Then they can easily get access to the teat canal during milking or suckling and can be transmitted from quarter to quarter and from cows to cows during milking practices. Their ability to exist intracellularly and localize within micro-abscessation in the udder and hence, resistant to antibiotic treatment (MacDonald, 1997) could also be important factor contributing to the predominance of these organisms.

E. coli were the predominant bacteria among the coliforms with an isolation rate of 4.82% in this study which is in consent with the observations of Mekuria (1986) and Biffa (1994) who reported 4.60%, 3.64% and 3.14%, respectively from different parts of Ethiopia. In this study, Klebsiella species accounted for 3.6% among coliforms.

There is also other coagulase negative Staphylococcus which contributes about 16.9% of the...
isolates which is in line with the report of Sori et al. (2005) in and around Sebeta (14.93%).

The occurrence of bovine mastitis and lactation stage was significantly (p<0.05) associated. That is, higher infection in cows in early lactation stage followed by late and medium lactation stages, that concurs with previous reports (Biffa et al., 2005). The early lactation stage infection might be due to the carryover of infection from dry period. In cows most new infections occur during the early part of the dry period and in the first two months of lactation (Radostits et al., 2007). Absence of dry cow therapy regime could possibly be the major factor contributing to high prevalence at early lactation and early infection associated with delayed diapedesis of neutrophils in to the mammary gland (Schalm et al., 1971).

Prevalence of mastitis was significantly (p<0.05) associated with milking hygienic practice. Cows at farms with poor milking hygiene standard are severely affected than those with good milking hygiene practices (Lakew et al., 2009; Sori et al., 2005). This might be due to absence of udder washing, milking of cows with common milkers’ and using of common udder cloths, which could be vectors of spread especially for contagious mastitis.

5. Conclusion and Recommendations

The present study showed that an overall prevalence of 38% bovine mastitis was recorded in the study area. Milk sample of 101 quarters, which were positive for CMT were cultured for microbiological examination in the study period, about 7 bacterial species and 83(82.2%) bacterial isolates were found. The predominant isolated bacteria were Staphylococcus aureus with isolation rate of 44.6% followed by Streptococcus agalactiae with isolation rate of 18.1%. Among the different potential risk factors considered for univariate logistic regression all the risk factors like: breed, age, parity, tick infestation of udder, sequence of milking, floor, husbandry system, lactation stage and hygiene of udder were found statistically significant (P<0.05). However, all became insignificant, except milking hygiene and stage of lactation when tested with multivariate logistic regression. Mastitis is still the most important disease of dairy animals in the study area.

In line with above facts the following recommendations are forwarded:

➢ Awareness creation should be given to the dairy herds on the impacts of bovine mastitis
➢ All quarters of the udder of each cow should be periodically checked for the timely treatment and prevention.
➢ To confirm the presence of sub clinical mastitis in the herd bacterial culture, SCC and CMT should be used.
➢ As there is no sufficient information about bovine mastitis in the study area, detailed and organized studies should be under taken to come up with relevant and appropriate control and preventive measurement.

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Corresponding author:
Dr. Tilahun Zenebe
School of Veterinary Medicine, Wollega University
Nekemte, Ethiopia, P.O. Box 395
Email- tilahun.zenebe@yahoo.com

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