

Role of CD14 Gene polymorphism and IgE in pathogenesis of Acute Bronchial Asthma

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Abstract: Acute bronchial asthma is characterized by acute episodes of obstruction related to airway inflammation mostly in response to respiratory tract infection. CD14 play a key role in inflammatory pathways. The aim of this study is to determine the influence of CD14 gene polymorphism and its plasma levels on the predisposition to develop asthma and its severity. This study was conducted on 20 patients during acute attack of asthma (group I) and in convalescence (after 6 weeks of treatment of the same patients, group II). Asthma severity scoring system was performed to all patients according to (Martin et al; 2006). Arterial blood gases, plasma levels of sCD14 and serum total IgE were measured by ELISA. CD14 gene polymorphism was detected by restriction fragment polymorphism (PCR). During acute asthma, plasma levels of sCD14 were higher than during convalescence: sCD14, 3.23 ± 0.48 versus 2.64 ± 0.24 $\mu\text{g/ml}$, ($p = 0.001$). Higher plasma levels of sCD14 were present during the acute attack in those with 159TT and 159TC ($p = 0.002$ & $p = 0.001$, respectively), whereas in 159CC homozygous, sCD14 levels were insignificantly increase during the acute attacks ($p = 0.119$). A significant negative correlation was found between sCD14 and asthma score ($r = 0.58$, $p = 0.001$). There was a significant positive correlation between IgE and plasma level of sCD14. These results suggest that in acute asthma production of sCD14 is increased in an attempt to control airway inflammation, and for subjects whose genotype limits or prevents these increases, the ability to control airway inflammation is impaired resulting in more severe asthma.

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Introduction:

Acute asthma is characterized by acute episodes of obstruction related to loss of control of airway inflammation mostly in response to a viral respiratory-tract infection (Martin et al., 2006). Many genetic variations associated with asthma and asthma phenotypes have been reported (He et al., 2001). Polymorphisms in several candidate genes have been identified and associated with asthma (Pinto et al., 2009). CD14 is important in clearly defined immunologic and inflammatory pathways (Antal-Szalmas 2000) Single nucleotide polymorphisms in the promoter regions of their genes alter the amount of expressed protein (Ohchi et al., 2004) and have been extensively studied in adults and children (Koppelman et al., 2001). Differences in gene expression have been identified among individuals with stable asthma, yet dysregulation of pro - or anti-inflammatory processes is likely to have the most critical influence during the early stages of an acute asthma attack, when the loss of control of inflammation could be expected to be maximal (Smit et al., 2008).

CD14 is a key component of the innate immune system and its gene is located on chromosome 5q31.1 (Isabel et al., 2008). It is

expressed on monocytes and macrophages, functions as a receptor for lipopolysaccharide receptor (Ulevitch and Tobias 1995), and exists in a membrane-bound form and a soluble form (sCD14). CD14 plays a critical role in determining the balance of Th1:Th2 cytokines, with activation promoting the release of interleukin 12 and deviation of immune responses towards an antiviral T-helper type 1 (Th1) response (Macatonia et al., 1995). A C-T polymorphism at position 159 in the promoter of CD14 (C-159T) modulates the cellular response to endotoxin and significantly influences total IgE levels (Kowal et al., 2008). The effect of this genetic variant on the cytokine response of the inflammatory cells is incompletely understood (Keskin et al., 2006). This polymorphism has been shown to be associated with increased levels of sCD14 and decreased serum immunoglobulin E (IgE) (Buckova et al., 2006) and the expression of a more severe atopic phenotype in previous studies (Smit et al., 2007).

CD14 gene may have had age-dependent effects on asthma-related phenotypes, including atopy and airway hyper responsiveness (Kiley et al., 2007). The CD14 protein is part of the receptor complex for endotoxin (O'Donnell et al., 2004) which is a potent

inflammatory agent and may contribute to the high prevalence of respiratory disorders (*Vercelli 2004*). Genetic variation in the CD14 gene may modify the interaction between endotoxin exposure, allergic diseases, and asthma severity (*Hasday et al., 1999*). The aim of this study is to determine the influence of CD14 gene polymorphism and its plasma levels on the predisposition to develop asthma and its severity.

Subjects and methods:

This study included 20 patients who attending Chest Disease Unite, Internal Medicine Departments, Mansoura University Hospital during acute attack of asthma (group I) and in convalescence (after 6 weeks of treatment of the same patients, group II). The patients were subjected to the following: Full history taking and complete clinical examination with stress on duration and severity of the disease; Arterial Blood Gases; Asthma severity scoring system was preformed to all patients according to (*Martin et al., 2006*); Plasma levels of sCD14 was measured by ELISA using commercially available ELISA kits (*R&D systems, Minneapolis, MN*); Serum total IgE was measured by ELISA (*Biomerieux Vitek Inc., Marcy, France*) (*Panhuysen et al., 1995*); CD14 gene polymorphism by restriction fragment polymorphism (PCR).

Extraction of genomic DNA:

Blood was collected in EDTA-tubes and stored at room temperature until the genomic DNA was extracted using QIA amp DNA Mini Kit from QIAGEN according to the manufacturer`s instructions .

Determination of CD14 polymorphism (*Ouburg et al., 2005*):

The C-T substitution in the proximal **CD14** promoter GC box at position -260 from the translation start site results in a **Hae III** restriction site. The primers, 5' TCA CCT CCC CAC CTC TCT T 3' (sense) and 5' CCT GCA GAA TCC TTC CTG TT 3' (antisense) (Invitrogen Life Technologies, Breda, The Netherlands), flanking this restriction site. Amplification was performed using a thermal cycler (Biometra). The parameters were an initial denaturation at 95°C for 5 min, followed by 35 cycles: denaturation at 95°C for 30 s, annealing at 59°C for 30 s, and elongation at 72°C for 1 min. The final elongation was at 72°C for 7 min followed for a cooling to 4°C. Then digested overnight at 37°C with **Hae III** (Invitrogen, The Netherlands). These fragments were analyzed by electrophoresis on 4% low melting agarose gels stained with ethidium bromide. The assay thus yields one 497-bp band for the CC genotype , three bands of 144, 353, and 497 bp for CT heterozygotes, and two bands of 144 and 353 bp for TT homozygotes.

Table (1) Scoring system of asthma severity

	1 Point	2 points	3 points
oxygen saturation	>90 %	90-95 %	<90%
Auscultation	Nil to mild end expiratory wheeze	expiratory wheezing	Inspiratory and expiratory wheezing or diminished breath sounds
Retractions	None or intercostal	Intercostal and substernal	Intercostal and substernal and supraclavicular
Dyspnoea	Speaks in sentences or coos and babbles	Speaks in partial sentences or utters short cries	Speaks in single words or short phrases or grunts
Respiratory rate: (/min)	2 – 6 years <36 7 – 9 years <31 10 – 13 years <27 >13 years <24	36 – 39 32 – 35 28 – 30 25 – 27	>39 >35 >30 >27

Statistical analysis:

The data was collected, presented and statistically analyzed with the computer program SPSS version 9 using the following tests, ANOVA test, Fisher Exact test, Chi-square test, unpaired test

and linear regression test. Probability (p) values of less than 0.05 were considered significant.

Results:

During acute asthma, plasma level of sCD14 was higher than during convalescence: CD14, 3.23±0.48 µg/ml versus 2.64±0.24 µg/ml (p=0.001);. Genotype percentages in acute asthma, the CC, CT& TT genotypes percentages for CD14 C-159T were (25%,42%,32.5% respectively) [Table 2]. During the acute episodes of asthma, a plasma level of sCD14 was significantly related to genotype for CD14 C-159T. Mean plasma sCD14 levels were highest in subjects with CD14-159TT (3.64±0.21 µg/ml), followed by heterozygous (3.2±0.36 µg/ml), and lowest for those with CD14-159CC (2.61±0.1 µg /m) p< 0.05 [Table 3]. While, IgE concentration was higher in CD14-159 CC genotype than both CT and TT genotypes respectively (p<0.05) [Table 5]. Higher

plasma levels of CD14 were present during the acute attack than convalescence in those with 159TT and 159TC (p=0.002 and p=0.001, respectively), whereas in 159CC homozygous, CD14 levels there was insignificant increase during the acute attacks than convalescence (p=0.119). [Table 4] CD14-159CC genotypes had significant higher mean asthma severity scores compared with the other genotypes (CD14 C-159T, CC 11.4 vs. CT 9.1 vs. TT 9.6, p <0.05, [Tables 6]. A significant negative correlation was found between sCD14 and asthma score (r=0.58, p=0.001). As regard IgE level, there was a significant positive correlation between IgE and plasma level of sCD14, and asthma score [Table 7].

Table (2) plasma sCD14 and CD14 genotype frequencies

	Group I (acute asthma)	Group II (Convalescence)	t	P
sCD14	2.5-4.1	2.1-3.2		
Range (µg /ml)	3.23±0.48	2.64±0.24	9.46	0.001*
Mean ±SD				
Genotypes				
	CC	CT	TT	
CD14 C-159T n (%)	10 (25)	17 (42.5)	13 (32.5)	

* Significant (p < 0.05).

Table (3) plasma concentration of sCD14 in relation to its corresponding genotypes:

CD14 C-159T	CC	CT	TT
Range(µg/ml)	2.5-2.8	2.9-3.5	3.3-4
Mean	2.61	3.27	3.64
SD ±	0.1	0.36	0.21
F (P)	40.24 (0.000*)		
Tukey's test			
CC &	CT	0.003*	CT & TT
	TT	0.001*	0.002*

Table (4): plasma sCD14 concentrations in relation to corresponding genotypes during acute asthma attack and convalescence

Genotypes	Acute asthma	Convalescence	t	p
CD14 C-159T				
TT	3.3-4	2.4-3.1		
Range	3.64±0.21	2.74±0.21	14.02	(0.002)*
Mean±SD				
TC	2.9-3.5	2.2-2.9	9.04	(0.001)*
Range	3.27±0.36	2.55±0.21		
Mean±SD				
CC	2.5-2.8	2.3-2.7	1.72	(0.119)
Range	2.61±0.1	2.5±0.12		
Mean±SD				

Table (5): Serum IgE concentrations among CD14 genotypes in all studied groups

Serum IgE	CD14 genotypes C-159T		
	CC	CT	TT
Range(µg/ml)	100-260	130-250	55.3-125.6
Mean±SD	198.5±49	191±41.1	87.9±24.2
F (P)	32.7 (0.000*)		
Tukey's test			
CC &	CT	0.82 NS	CT & TT 0.000*
	TT	0.000*	

NS: non significant

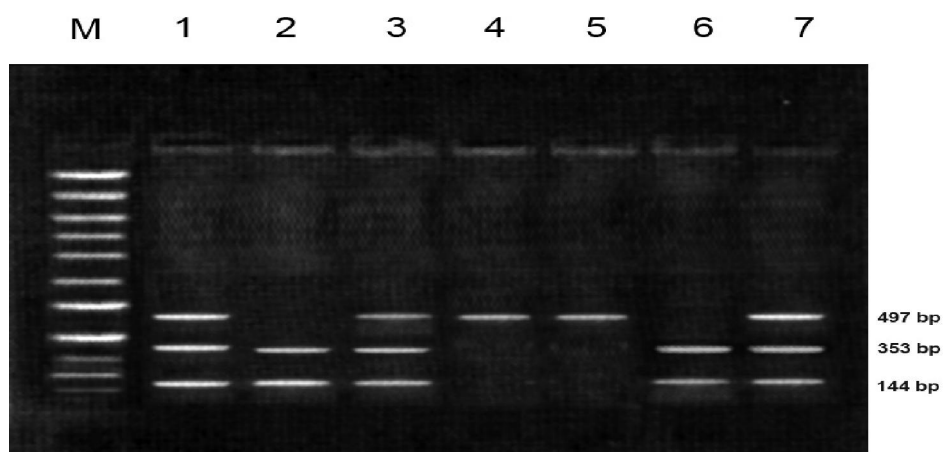


Fig (1): Agarose gel electrophoresis for PCR products stained with ethidium bromide detecting Polymorphism at 159 bp of flanking region of the CD14 gene by restriction fragment assay. CC homozygotes are in lanes(4 and 5) with single band of 497 bp, TT homozygotes are in lanes(2 and 6) with two bands of 144 and 353 bp: and CT

Table (6): Asthma severity score and its relationship to CD14 genotypes:

Asthma severity score	CD14 genotypes C-159T		
	CC (n=10)	CT (n= 17)	TT (n= 13)
Range	8-15	5-11	6-10
Mean ±SD	11.4±2.0	9.1±1.8	9.6±2.0
F (P)	5.98 (0.005*)		
Tukey's test			
CC &	CT	0.004*	CT & TT 0.53 NS
	TT	0.003*	

Table (7): Correlation between asthma score and CD14 and IgE:

	Asthma score		IgE	
	r	p	r	p
sCD14	-0.58	0.001	0.517	0.002
IgE	0.436	0.005		

Discussion:

Bronchial asthma is a complex disease caused by a combination of genetic and environmental factors (*Hammad and Lambrecht 2008*) and exposure to various environmental stimuli can effect on the pathogenesis of the disease (*Tamari et al., 2009*). Many genetic variations associated with asthma and asthma phenotypes have been reported (*Cookson 2004*). Although a large number of studies have identified possible genetic loci and chromosomal mutations that may be involved in the development of asthma or these related phenotypes with stable disease (*Schwartz 2009*), additional researches are needed to clarify the influence of genetic differences in acute asthma and asthma exacerbation, as transcription of genes involved in asthmatic inflammation is likely to be maximal during acute exacerbation. CD14 is a multifunctional receptor expressed in the surface of monocytes, macrophages and neutrophils or serum soluble (*Ulevitch and Tobias 1995*). It is the main receptor of lipopolysaccharides (LPS) or inhaled endotoxins, which are potent inducers of pulmonary inflammation and may activate the immune system and cause Th1 differentiation and/or Th2 suppression (*Isabel et al., 2008*). It has been proposed that altered CD14 expressions, more increased in asthmatics after LPS inhalation, can change the balance of Th1-Th2 cells, influencing IgE levels and inflammations in allergic diseases such as asthma (*Alexis et al., 2001*). Thus changes in CD14 expression seem to be important, especially in allergic asthma, and functional polymorphic variants of the CD14 gene modulate the response to LPS (*Simpson et al., 2006*). The most prevalent single nucleotide polymorphism in CD14 is the C-159T polymorphism, which is associated with changes in serum IgE level (*William et al., 2006*). The genetic variants of the CD14 promotor region affect both monocyte expression of CD14 and serum concentration of soluble CD14 (*Kowal et al., 2008*). In order to investigate the influence of genetic difference in acute asthma to elucidate the underlying mechanisms of acute exacerbation, we study genetic polymorphism for CD14 and its t plasma level during acute asthma and during convalescence. However the association between the C-159T CD14 polymorphism and asthma has been demonstrated in some but not all studies (*Sharma et al., 2004; Kedda et al., 2005; Zambelli-Weiner et al., 2005 and Tan et al., 2006*). In the present study, we found that acute asthma, plasma level of sCD14 was higher than during convalescence ($p < 0.001$) and this higher level were present in one genotype than others. During the acute episodes of asthma, plasma levels of sCD14 were significantly related to genotype for CD14 C-159T. Mean plasma sCD14 levels were highest in subjects with CD14-159TT, followed by heterozygous and lowest for those with CD14-159CC. These results are in

agreements with studies of *Martin et al., 2006* and *Kowal et al., 2008*, they found that sCD14 were higher only in subjects with CD14-159 TT and 159CT but not in those with 159CC. This study demonstrated that genotype-specific differences in plasma levels of sCD14 during acute exacerbation was even more pronounced than during convalescence, a higher plasma levels of CD14 was present during the acute attack in those with 159TT and 159TC ($p = 0.002$ and $p = 0.001$, respectively), whereas in 159CC homozygous, CD14 levels there was insignificant increase during the acute attacks ($p = 0.119$). Although *Kedda et al., 2005* found that C-159T polymorphism has been associated with increased sCD14 in children and there were no difference in CD14 serum levels in adults with different genotypes for CD14. Enhanced expression of CD14 and increased sCD14 levels that seen in subjects carrying the TT genotype leads to increased activation of cells involved in the immune response and increased secretion of proinflammatory mediators including TNF- α , IL-1B, IL-12 and IFN- γ skewing the immune response towards Th1 (*Keskin et al., 2006; Renckens et al., 2006 and Hong et al., 2007*). Serum level of IgE concentration was higher in CD14 -159 CC genotype than both CT and TT genotypes respectively ($p_1 = 0.002$, $p_2 = 0.003$). These results are in accordance with *Baldini et al., 1999* who found that CD14/-159 TT homozygous showed significantly lower levels of total serum IgE than did both CC and CT subjects and they explain this by their finding that an association between sCD14 levels and low IL14 and high IFN- γ , which intern may regulate IgE responses and consequently IgE levels (*He et al., 2001; Pinto et al., 2009; Buckova et al., 2006 and Williams et al., 2006*). In this study, CD14-159CC genotypes had significant higher mean asthma severity scores compared with the other genotypes (CD14 C-159T, CC 11.4 vs. CT 9.1 vs. TT 9.6, $p < 0.05$). Those subjects who was CD14 -159CC was over three times more likely to have moderate or severe attacks of acute asthma compared with the other genotypes (*Zambelli-Weiner et al., 2005*). The preferential transmission of the C allele to children with severe atopic asthma was demonstrated in an Indian population (*Sharma et al., 2004*) and an association between the T allele and lower asthma severity scores was found in population from Barbados (*Zambelli-Weiner et al., 2005*). Thus analysis of the frequency of the C allele in our patients provides further support for the association between that polymorphism and asthma severity as the increased frequency of the C allele was found in patients with moderate to severe asthma but not in mild asthmatics, linking the C allele of CD14 with increased risk of asthma severity and exacerbation. A potential mechanism in determining severity of an acute attack is impaired ability to increase sCD14 (*Kowal et al., 2008*). The effect of CD14

C-159T on asthma severity was partly dependent on plasma sCD14 levels, with higher levels associated with a milder asthma attack, possibly because of increased Th1 cytokine release (*Keskin et al., 2006 and Kedda et al., 2005*). A significant negative correlation was found between sCD14 and asthma score ($r=0.058$, $p=0.001$). Consistent with this reports, *Martin et al., 2006*, found that during acute asthma attacks in children, plasma sCD14 levels were inversely correlated with severity, suggesting a protective role for sCD14. Plasma sCD14 levels have been directly correlated with IFN- γ and inversely correlated with IL-4, suggesting that in children with acute asthma, the beneficial effect of greater CD14 activity may result from increased Th1 and decreased Th2 responses (*He et al., 2001 and Isabel et al., 2008*). A relative predominance of Th1 over Th2 cytokines assists in the elimination of viral infections (*Ramshaw et al., 1997 and Hoebee et al., 2003*), thus lower levels of sCD14 may allow ongoing viral replication and inflammation. These results suggest that in acute asthma production of CD14 is increased in an attempt to control airway inflammation, and for subjects whose genotype limits or prevents these increases, the ability to control airway inflammation is impaired, resulting in more severe asthma. Therefore, this study supports the concept that sequence variations in the CD14 gene are likely to play a role in the development of asthma.

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References:

1. Martin A.C, Laing I.A, KimKhoo S. et al.: Acute asthma in children: relationships among CD14 and CC16 genotypes, plasma levels, and severity. *Am J Respir and Crit Care Med.*2006;173: 617-22.
2. He JQ, Joos L, Sandford AJ:Recent developments in the genetics of asthma. *Pharmacogenomics* 2001; 2: 329-39.
3. Pinto LA, Stein RT, Ribeiro JD: Genetic association with asthma and virus – induced wheezing: a systematic review. *J Bras Pneumol.*2009;35(12): 1220-26.
4. Antal-Szalmas P: Evaluation of CD14 in host defense. *Eur J Clin Invest.*2000;30: 167-79.
5. Ohchi T, Shijubo N, Kawabata I, et al: Polymorphism of Clara cell 10-kD protein gene of sarcoidosis. *Am J Respir and Crit Care Med.*2004;169: 180-6.
6. Koppelman GH, Reijmerink NE, Colin Stine O, et al: Association of a promoter polymorphism of the CD14 gene and atopy. *Am J Respir Crit Care Med.*2001;163: 965-9.
7. Smit LA, Siroux V, Bouzigon E. et al: CD14 and Toll- like receptor gene polymorphisms, Country living , and asthma in adults. *Am J Respir Crit Care Med.*2008;179: 363-68.
8. Isabel CJ, Elisangela J, Adyleia AD. Et al: Association of TGF-B, CD14, IL-4, IL-4R and ADAM33 gene polymorphisms with asthma severity in children and adolescents. *J de Pediatria.*2008;84(3): 203-10.
9. Ulevitch RJ, and Tobias PS: Receptor-dependent mechanisms of cell stimulation by bacterial endotoxin. *Annu Rev Immunol.*1995;13: 437-57.
10. Macatonia SE, Hosken NA, Litton M, et al: Dendritic cells produce IL-12 and direct the development of Th1 cells from naive CD4+ T cells. *J Immunol.*1995;154: 5071-9.
11. Kowal K, Bodzenta-Lukaszyk A, Pampuch A., et al: Analysis of -675 4G/5G SERPINE1 and C-159T CD14 polymorphisms in house dust mite-allergic asthma patients. *J Investig Allergol Clin Immunol.*2008;18(4): 284-92.
12. Keskin O, Birben E, Saçkesen C, et al: The effect of CD14-c159T genotype on the cytokine response to endotoxin by peripheral blood mononuclear cells from asthmatic children *Ann Allergy Asthma Immunol.* Sep 2006; 97(3): 321-8.
13. Buckova D, Lzakovkova Holla L, Znojil V, Vasku A: Polymorphisms of the CD14 gene and atopic phenotypes in Czech patients with IgE-mediated allergy. *J Hum Genet.*2006;51: 977-83.
14. Smit A. M., Bongers S. I. M., H Ruven. J. T., G. et al: Atopy and new-onset asthma in young Danish farmers and CD14, TLR2, and TLR4 genetic polymorphisms: a nested case-control study. *Clinical & Experimental Allergy.*2007;37: 11, 1602-8.
15. Kiley J, Smith R and Noel P: Asthma phenotypes. *Curr Opin Pulm Med.*2007;13: 19-23.
16. O'Donnell AR, Toelle BG, Marks GB, et al: Age-specific relationship between CD14 and atopy in a cohort assessed from age 8 to 25 years. *Am J Respir and Crit Care Med.*2004;169: 615-22.
17. Vercelli D: Genetics, epigenetics, and the environment: switching, buffering, releasing. *J. Allergy. Clin. Immunol.*2004;113: 381-6.
18. Hasday JD, Bascom R, Costa JJ, et al: Bacterial endotoxin is an active component of cigarette smoke. *Chest.*1999;115: 829–35.
19. Panhuysen CI, Bleecker ER, Koeter GH et al: Dutch approach to the study of the genetics of

- asthma. *Clin. Exp. Allergy*.1995;25 (Suppl 2): 35-8.
20. Ouburg S., Spaargaren J., Janneke E den Hartog J.E., et al: The CD14 functional gene polymorphism -260 C.T is not involved in either the susceptibility to *Chlamydia trachomatis* infection or the development of tubal pathology. *BMC Infect Dis*.2005;5: 114.
 21. Hammad H, and Lambrecht BN: Dendritic cells and epithelial cells: linking innate and adaptive immunity in asthma. *Nat Rev Immunol*.2008;8: 193-204.
 22. Tamari M, Harada M, Hirota T and Nakamura Y: Host molecular defense mechanisms against *chlamydia pneumonia* and genetic studies of immune-response-related genes in asthma. *Recent Patents on Inflammation & Allergy Drug Discovery*.2009;3(1): 17-25.
 23. Cookson W: The immunogenetics of asthma and eczema: A new focus on the epithelium. *Nat Rev Immunol*.2004;4: 978-88.
 24. Schwartz DA: Gene-Environment interaction and airway disease in children. *Pediatrics*.2009;123: S151-S159.
 25. Alexis N, Eldridge M, Reed W., et al: CD14-dependent airway neutrophil response to inhaled LPS: role of atopy. *J Allergy Clin Immunol*.2001;107: 31-35.
 26. Simpson A, John SL, Jury F. et al: Endotoxin exposure, CD14 and allergic disease. An interaction between genes and the environment. *Am J Respir Crit Care Med*.2006;174: 386-92.
 27. Williams LK, Mcphu RA, Ownky DR. et al: Gene-environment interaction with CD14 C-260T and their relationship to total serum IgE levels in adults. *J Allergy Clin Immunol*.2006;118: 851-57.
 28. Sharma M, Batra J, Mabalirjan U et al: Suggestive evidence of association of C-159T functional polymorphism of the CD14 gene with atopic asthma in northern and northwestern Indian populations. *Immuno genetics*.2004;56: 544-47.
 29. Kedda MA, Lose F, Duffy D. et al: The CD14 C-159T polymorphism is not associated with asthma or asthma severity in an Australian adult population. *Thorax*.2005;60: 211-14.
 30. Zambelli-Weiner A, Ehrlich E, Stocktan ML. et al: Evaluation on CD14/-260 polymorphism and house dust endotoxin exposure in the Barbados asthma. Genetics study. *J Allergy Clin Immunol*.2005;115: 1203-209.
 31. Tan CY, Chen YL. Wu LSH . et al: Association of CD14 promotor polymorphisms and soluble CD14 levels in mite allergen sensitization of children in Taiwan. *J Hum Genet*.2006;51: 59-67.
 32. Renckens R, Pater JM, Van der Poll T: Plasminogen activator inhibitor type-1 deficient mice have enhanced IFN-g response to lipopolysaccharide and staphylococcal enterotoxin B. *J Immunol*.2006;177: 8171-76.
 33. Hong SJ, Kim HB, Kang MJ. et al: TNF- (-308G/A) and CD14 (-159T/C) polymorphisms in the bronchial responsiveness of Korean children with asthma. *J Allergy Clin Immunol*.2007;119(2): 398-404.
 34. Baldini M, Lohman IC, Halonen M et al: Polymorphism in the 5' flanking region of CD14 gene is associated with circulating soluble CD14 levels and with total serum immunoglobulin E. *Am J Respir Cell Mol Biol*.1999; 20: 976-83
 35. Ramshaw IA, Ramsay AJ, Karupiah G, et al. (1997): Cytokines and immunity to viral infections. *Immunol*. 1997;Rev;159:119-35.
 36. Hoebee B, Rietveld E, Bont L. et al: Association of severe respiratory syncytial virus bronchiolitis is with interleukin-4 and interleukin-4 receptor alpha polymorphisms. *J Infect Dis*.2003;187: 2-11.

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