**Effect of total aflatoxin on the growth characteristics and chlorophyll level of sesame (*Sesamum indicum L*.)**

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**Abstract:** This study investigated the effect of total aflatoxin (1000µg/l, 500µg/l, 250µg/l, 125µg/l, and 0µg/l) on the chlorophyll level and some growth characteristics of sesame (*Sesame indicum)* using bogoro, E8, Ex-sudan varieties as the plant bioindicators. The experiment was conducted on the Department of Biological Sciences experimental field, Faculty of Science, University of Abuja. Abuja. The experimental design adopted was Randomized Complete Block Design (RCBD) involving three replicates. The seedling emergence, shoot length, root length, seedling vigour and chlorophyll level of the sesame cultivars at 10 Days After Sowing (DAS) under the control plot was significantly (*p*<0.05) higher than other treatments. All the parameters measured under the control plots of E8 and Ex-sudan cultivar were significantly (p<0.05) higher than other treatments. However, the number and area of leaves and number of flowers of *bogoro* cultivar at 56 DAS were significantly (p<0.05) higher at 125µg/l treatment than at other levels. The study showed an inhibitory effect on all the growth characteristics of sesame by total aflatoxin especially above 125µg/l. The biotest of mycotoxin on various crop seeds is imperative. Also the fate of total aflatoxin on the sown seeds in the rhizosphere requires investigation.

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**1. Introduction**

Sesame (*Sesamum indicum* L*.*) is a flowering plant in the genus *Sesamum* and the family Pedaliaceae. It is widely naturalized in tropical regions around the world and it is cultivated for its edible seeds, which grow in pods (Bedigian, 2010). Sesame is a common staple food and very drought-tolerant, in part due to its extensive root system, it requires adequate moisture for germination and early growth (FAO, 2012). For thousands of years, sesame seeds have been a source of food and oil. Sesame has one of the highest oil content of any seed, some varietals exceeding 50% oil content compared to soybean’s 20% (Langham, 2008). Sesame oil is one of the most stable vegetable oils, with long shelf life, because of the high level of natural antioxidants (sesamin, sesamolin, and sesamol). Oil from the seed is used in cooking, as salad oils and margarine, and contains about 47% oleic and 39% linoleic acid (Ivon *et al.,* 2005). Sesame seed is also rich in protein, at 25% by weight. The flour that remains after oil extraction is between 35 to 50 % protein, has good effective carbohydrates, and contains water-soluble antioxidants (sesaminolglucosides) that provide added shelf-life to many products. This flour, also called sesame meal, is an excellent high-protein feed for poultry and livestock. Sesame seeds may be baked into crackers, often in the form of sticks (Oplinger *et al.*, 2000). In Greece the seeds are also used in cakes. Fast-food restaurants use buns with tops sprinkled with sesame seeds. Aflatoxin-producing Aspergillus could inhabit the soil, decaying vegetation, hay, and grains undergoing microbiological deterioration and it invades all types of organic substrate whenever conditions are favourable for its growth, favourable conditions include high moisture content (at least 70%) and high temperature (Machida, 2010; Leong *et al.,* 2011, McDaniel, 2011). Chlorophyll is an extremely important biomolecule, chlorin [pigment](http://en.wikipedia.org/wiki/Pigment) found in the [chloroplasts](http://en.wikipedia.org/wiki/Chloroplast) of [plants](http://en.wikipedia.org/wiki/Plant). It is critical in [photosynthesis](http://en.wikipedia.org/wiki/Photosynthesis) as it allows plants to absorb [energy](http://en.wikipedia.org/wiki/Energy) from light (Anthony *et al.*, 2003). Chlorophyll molecules are specifically arranged in and around [photosystems](http://en.wikipedia.org/wiki/Photosystem) that are embedded in the [thylakoid](http://en.wikipedia.org/wiki/Thylakoid) membranes of [chloroplasts](http://en.wikipedia.org/wiki/Chloroplast) (Chen *et al.,* 2010). Chlorophyll content can be an indicator of the plant’s condition.

Biotests of mycotoxin can facilitate an objective evaluation of the effect of a toxin level, due to the fact that all higher plants have certain sensitivity to different xenobiotics found in the soil environment. El-Taher et al., 2012 recommended that the characteristics of plant bioindicators to be used in phytotests should have small and even seeds, uniform germination power and energy of seeds, a short emergence period (1-2 days), a short vegetation period, high biomass of stems, leaves or roots and have high sensitivity in relation to one chemical group such as aflatoxins, fumonisins. There is dearth of scientific reports on biotests of mycotoxin using crop seeds as the bioindicator in Nigeria, thus this research investigated the effect of total aflatoxin on the seedling emergence, growth characteristics and chlorophyll level of three sesame varieties.

**2 Materials and methods**

**2.1Collection of seed samples**

The seeds of sesame such as Ex-sudan, Bogoro and E8 were collected from the National Cereal Research Institute (NCRI) Badeggi, Bida, Niger State in Nigeria. The E8 variety are characterized by early maturing, high yielding and yellowing of leaves, the Ex-sudan are characterized by high yielding and late maturing(120 days), while the Bogoro local are dwarf and short. The seeds were carefully cleaned and freed from foreign materials.

**2.2 Experimental site and layout**

The study was conducted in the field which were carried out in black polythene bags filled with rich sandy loam soil in texture, sand, silt, and clay content was 74, 16, and 10% respectively, slightly acidic in reaction (*p*H = 5.30), low to medium in electrical conductivity (0.150dsm-1), with low levels of organic carbon (0.0.87%) and nitrogen (0.175kg/ha). The bags were randomly arranged on the experimental field of Department of Biological Sciences, Faculty of Science, University of Abuja, Abuja. The experimental plots were arranged in Randomized Complete Block Design (RCBD) involving five concentration levels of total Aflatoxin (1000µg /l, 500µg /l, 250µg /l, 125µg /l and 0µg /L of distilled water) to each of the three sesame varieties (Ex- sudan, Bogoro, E8) seeds before sowing.

**2.3 Application of total aflatoxin to seed samples**

The chemical, Total Aflatoxin; Aflastandard product code: p22/ p22A were procured from R. Biopharm Rhone Ltd, Indonesia through Chromogene Ltd and the bottle stored at 2oC - 8oC.The stock solutions were apropriately diluted at the time of sowing to obtain the required concentration.1000 µg per litre of total Aflatoxin was serial diluted to obtain 500 µg, 250 µg, 125 µg, 0µg per distilled water. All the sesame seeds were initially soaked in distilled water for an hour, before 10 gram of the seeds were subsequently transferred to the beaker (500ml) filled with each level of concentration. The setup was allowed to stay for three hours before sowing.

**2.4 Sowing of treated Seeds**

On the 28th August 2012, 10 seeds were sown per bags and covered lightly with top soils. Regular watering was carried out as required.

**2.5 Data Collection**

Data were collected on the germination %age, shoot length, root length, seedling vigour, chlorophyll level, number of leaves, area of leaves and number of flowers.

**2.6 Seed emergence, shoot and root length** The percentage of seedling emergence of the various cultivars were taken by counting the plants that emerged after ten days, and was multiplied by hundred (100). The shoot and root length were measured in millimetre, which was taken by selective uprooting of 3 plants at a time.

**2.7 Seedling vigour**

The seedling vigour of various cultivars was taken by using the formula adopted by (Okelola, 2005):

(sl + rl) x g%

where sl= shoot length; rl = root length; g% = germination percentage

**2.8 Chlorophyll level**

The chlorophyll level in sesame was recorded by aid of a chlorophyll-metre (Atleaf®) assembled, tested and calibrated in the United States. It was used to determine the percentage of chlorophyll on 3 selected leaves per pot of sesame seedlings. Atleaf® is a powerful, handheld, easy to use device for noninvasively measuring the relative chlorophyll content of green leaf plants. Chlorophyll content can be an indicator of the plant’s condition. The device is. The Plant relative chlorophyll concentration was measured by inserting a leaf into the device aperture, Green leaves of up to 3mm thickness can be measured. It only takes one press of the key to perform a basic measurement (Gitelson, 1999). The device has simple, one-key operation for measuring the optical density difference at 2 wavelengths (660nm and 940nm), displays 2 lines x 16 characters and it powered by 2 AA (1.5V) batteries.

**2.8 Number of Leaves and flowers**

The data on the number of leaves and flowers were taken by counting of the available leave on each cultivar under each treatment at 56DAS.

**2.9 Area of leaves**

The area of leaves of the sesame cultivars were measured on 56DAS using graph sheet with cm2 grids.

**2.10 Statistical Analysis of Data** Data collected was subjected to ANOVA and the means were separated with Duncan Multiple Range (DMRT). Varietal x concentration effects were determined by using Statistical Analysis Package SPSS Statistical Version 17.0.

**3. Results**

Table 1 shows that the seedling emergence of bogoro at 10DAS under the control plot was significantly (p<0.05) higher than other treatments. There was no significant (p>0.05) difference in the emergence of bogoro cultivar applied with 500µg/l and 100µg /l. Also, the shoot and root length, seedlings vigour and chlorophyll level of the bogoro cultivar under the control plot was significantly (p<0.05) higher than those treated with aflatoxin. Application of total aflatoxin at 500 or 1000µ/l significantly (p<0.05) reduced all the parameters measured than under other treatments.

The seedling emergence, shoot length, root length, seedling vigour and chlorophyll level of E8 sesame cultivar in the control plots were significantly (p<0.05) higher than other treatments. There was no significant (p>0.05) difference between the seedling emergence of the E8 seeds applied with 125ug/l and the control.

The germination emergence, shoot and root length, seedling vigour and chlorophyll level of Ex-sudan cultivar in the control plot was significantly (p<0.05) higher than other treatments. Total aflaotoxin application at 1000µg/l resulted in the lowest significant reduction in the germination percentage, root length and the chlorophyll level. The shoot length of 125µg/l treatment of the Ex-sudan cultivar was the lowest. The interaction between the varieties and treatments led to highly significance (p<0.05) difference on the germination percentage, shoot length, root length, seedling vigour and chlorophyll level as shown in the Table 1.

Table 1. Effect of total aflatoxin on the germination percentage, shoot length, root length, seedling vigour and chlorophyll level of three sesame cultivars at 10DAS

|  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- |
| **Variety** | **Concentration** (µg /L) | **Seedling emergence** | **Shoot length (mm)** | **Root length (mm)** | **Seedling vigour (x100)** | **Chlorophyll level (index)** |
| Bogoro | 0 | 96.00a | 19.75a | 16.50a | 1895.96a | 35.85a |
|  | 125 | 90.00b | 17.50b | 15.25b | 1575.06b | 31.77b |
|  | 250 | 70.00c | 17.00b | 12.25c | 1190.66c | 21.50c |
|  | 500 | 40.00d | 12.75c | 10.75d | 510.00d | 20.10d |
|  | 1000 | 40.00d | 13.00c | 10.80d | 520.66d | 20.20d |
| E8 | 0 | 99.00a | 31.00a | 36.75a | 3068.66a | 32.30a |
|  | 125 | 99.00a | 16.50c | 20.25d | 1633.53c | 30.22c |
|  | 250 | 75.00b | 19.50b | 32.00b | 1852.56b | 31.05b |
|  | 500 | 40.00c | 12.75d | 10.75e | 510.00 e | 20.10e |
|  | 1000 | 50.00c | 16.00c | 24.70c | 1440.66d | 28.60d |
| Ex-sudan | 0 | 70.00a | 28.25a | 22.50a | 1977.55a | 34.24a |
|  | 125 | 55.00c | 20.00d | 20.00b | 1100.66d | 30.00b |
|  | 250 | 65.00b | 23.40c | 22.50a | 1521.06b | 29.93b |
|  | 500 | 55.00c | 24.50c | 19.80b | 1343.83c | 29.23b |
|  | 1000 | 45.00d | 25.00b | 18.75c | 1125.66d | 27.60c |
| Interaction (variety x level) |  | \*\* | \*\* | \*\* | \*\* | \*\* |

Figure 1. Effect of total aflatoxin on the seedling vigour on sesame cultivars

Figure 2. Effect of total aflatoxin on the chlorophyll level on sesame cultivars

The effects of total aflatoxin application on the number of leaves, area of leaves and number of flowers of three cultivars of sesame at 56DAS on the field are as shown in Table 2. The number of leaves and number of flowers of the 3 sesame cultivars applied with total aflatoxin were adversely affected.

It was indicated that the number of leaves (56DAS) in bogoro under the 1000µg/l treatment was significantly (p<0.05) higher than other levels of treatments. There was no significant (p>0.05) difference in the number of leaves of bogoro applied with 500ug/l and the control. The area of leaves of bogoro treatment in the 1000µg/l treatment was significant (p<0.05) higher than other treatments. There was no significant (p>0.05) difference between the area of leaves of the bogoro seeds applied with 500µg/l, 125µg/l and the control. The number of flowers under the 1000µg/l treatment in the bogoro cultivar was significantly (p<0.05) higher than other treatments. Total aflatoxin application at 250µg/l resulted in the lowest significant reduction in the number of flowers of the bogoro cultivar.

Table 2. Effect of total aflatoxin on the no of leaves, area of leaves and no f flowers on three sesame cultivars at 56 DAS

|  |  |  |  |  |
| --- | --- | --- | --- | --- |
| **Variety** | **Concentration**  (µg /L) | **No of leaves** | **Area of leaves (mm2)** | **Number of flowers** |
| Bogoro | 0 | 11.16b | 11.22c | 3.16b |
|  | 125 | 19.00a | 18.64a | 6.66a |
|  | 250 | 10.33c | 16.56b | 2.50d |
|  | 500 | 9.66d | 11.05c | 1.80e |
|  | 1000 | 8.85d | 15.49b | 2.66c |
| E8 | 0 | 13.83a | 29.87a | 7.66a |
|  | 125 | 12.83b | 16.86c | 7.00b |
|  | 250 | 12.33c | 22.15b | 6.00b |
|  | 500 | 9.66d | 11.05d | 1.80c |
|  | 1000 | 11.66e | 13.00d | 2.16c |
| Ex-sudan | 0 | 63.00a | 28.35a | 8.00a |
|  | 125 | 51.50b | 22.40b | 8.00a |
|  | 250 | 49.00c | 20.25c | 5.00b |
|  | 500 | 46.00d | 16.10d | 3.50b |
|  | 1000 | 26.50e | 10.85 e | 3.00c |
|  | Interaction (varietal x level) | \*\* | \*\* | \*\* |

The number of leaves of the E8 cultivar in the control plot was significantly (p<0.05) higher than other treatments. Application of aflatoxin at 500µg/l treatment significantly (p<0.05) reduced all the parameters measured than under other treatments. Area of leaves of the E8 cultivar in the control plot was significantly (p<0.05) higher than other treatment. There was no significant (p>0.05) difference between the area of leaves of the E8 seeds applied with 500µg/l and 1000µg /l. The number of flowers of E8 cultivars under the control plot was significantly (p<0.05) higher than other treatment. There was no significant (p>0.05) difference between the number of flower of E8 cultivar of treatments applied with 500µg/l and 1000µg/l.

The number of leaves, area of leaves and number of flowers of Ex-sudan cultivar in the control plot was significantly (p<0.05) higher than other treatments. Total aflatoxin application at 1000µg/l resulted in the lowest significant reduction in the number and area of leaves and number of flowers. The interaction between the varieties and treatment produced a highly significance difference on the number and area of leaves and number of flowers (Table 2).

**4. Discussion**

The phytotoxic effect of toxins may be observed on the basis of the reduction of dry or fresh weight of roots or above ground parts (stems, leaves) of test plants (Demczuk et al., 2004); On the basis of selected parameters, such as the reduction of root length, the toxic effect of a toxin may be determined already after approx. 24h, while the dynamics of root growth - after 3-5 days from the onset of the test. In turn, the reduction in fresh or dry weight of aboveground parts of plants may be established after approximately 10-14 days in a conventional biotest (Sekutowski and Sadowski, 2006).

In this study, there was an inhibitory response on the chlorophyll and growth characteristics of the biotest sesame cultivars, at high concentration of aflatoxin application. Sinha and Kumari (1990) and Adekunle and Basir (1997) reported that cowpea *(Vigna unguiculata L.* Walp) seed germination and chlorophyll formation were inhibited with increased aflatoxin concentration. Fratamico, et al., (2008) observed that preharvest aflatoxin contamination of peanuts and corn is favoured by high temperatures, prolonged drought conditions, and high insect activity; while postharvest production of aflatoxins on corn and peanuts is favoured by warm temperatures and high humidity. Crison (1997) observed that seedling growth was not inhibited at lower concentration of aflatoxin per ml of agar substrate but there was an inhibitory effect on elongation of the hypocotyls and the root in the species studied at higher tage age of aflatoxin.

Application of 125µg/l aflatoxin showed a remarkably different effect on bogoro cultivar by stimulating increased number and area of leaves. Many toxic substances like pesticides have been reported to have shown growth stimulating or hormone-like properties at sublethal concentrations (Wu *et al*., 1972). This is by the stimulation of DNA-directed RNA and protein synthesis

To periodically quantify the concentration of a tested toxin in the rhizosphere, a set of PhytotoxkitTMbiotests allowing for the evaluation of phytotoxicity of tested samples within a short time (1-3 days) could be used Sekutowski and Sadowski 2009, Mahoney, 2010). The determination of the level of degradation rate and translocation of toxic active ingredients in the soil is significant in crop environment. Instrumental methods, such as liquid high performance chromatography (HPLC), make it possible to determine the total content of toxins in the soil days or weeks after the application of the toxin (Ahmad and Crawford 1990, Sekutowski and Sadowski, 2006).

When selecting a bioindicator for a test it is necessary to take into consideration the age and the location of thev plant and the sensitivity of individual tissues to the tested toxin. Sadowski and Crawford (1990) reported that the youngest roots and leaves of test plants turned out to be most sensitive. They further observed that an increase in the initial soil moisture content by 2% caused a marked shift of toxin deeper within the soil profile practically for all the tested active ingredients.

It is evident from this research that the most on seed emergence of sesame cultivars was achieved at It was evident that high concentration of aflatoxins at 500 and 1000 µg /l. had significant inhibitory effect on the shoot length, root length, seedling vigour, and chlorophyll level in sesame. At 125µg/l concentration aflatoxin, a significant stimulatory effect on the number of leaves, area of leaves and number of flowers of bogoro cultivar was observed. In all situations, the chlorophyll level of all sesame cultivars was higher at lower concentration and was more obvious on E8 and Ex-sudan cultivars.

Contaminated seeds with aflatoxigenic fungi should not be used for propagation except they are pretreated with fungitoxicant. There is a need for further study on the effects of other mycotoxins on crop seeds and the rate of degradation and translocation of toxin in the crop rhizosphere.

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**Reference****s**

1. Bedigian D, Sesame: The genus Sesamum. CRC Press, London. 2010, pp 56
2. FAO - Food and Agriculture Organization of the United Nations. “Production crops sesame seeds. [*http://faostat.fao.org/site/567/DesktopDefault.aspx?PageID=567#ancor*](http://faostat.fao.org/site/567/DesktopDefault.aspx?PageID=567#ancor)*.* 2012; Retrieved 2012-06-17
3. Langham RD, Phenology of Sesame. American Sesame Growers Association. [*http://www.sesamegrowers.org/langham144-182.pdf.* 2008](http://www.sesamegrowers.org/langham144-182.pdf.%202008); Retrieved2012-09-17
4. Ivon EJ, Milder ICW, Arts BV, Dini P, Venema P, Hollman C H, Lignan contents of Dutch plant foods: a database including lariciresinol, pinoresinol, secoisolariciresinol and matairesinol. British Journal of Nutrition (2005), 93: 393-402
5. El-Taher EM, Abd El-Ghany TM, Alawlaqi MM., Mona SA, Biosecurity for reducing ochratoxin a productivity and their impact on germination and ultrastructures of germinated wheat grains Journal of Microbiology, Biotechnology and Food Sciences, 2012 : 2 (1) 135-151
6. Oplinger ES, Putnam DH, Sesame. Purdue University. 2000; Accessed [*http://www.hort.purdue.edu/newcrop/afcm/sesame.html*](http://www.hort.purdue.edu/newcrop/afcm/sesame.html)*.*
7. Machida M, Gomi K. Aspergillus*: Molecular Biology and Genomics*. Caister Academic Press, 2010.
8. Leong Y, Ismail N, Latiff A, Manaf N, Rosma, A, "Determination of aflatoxins in commercial nuts and nut products using liquid chromatography tandem mass spectrometry". World Mycotoxin Journal. 2011; 4 (2): 119–127.
9. McDaniel A, "Effect of Matrix Clean-Up for Aflatoxin Analysis in Corn and Dried Distillers Grains". Natural Resources. 2011; 02 (04): 250–257.
10. Anthony WD, Larkum, SE Douglas K, John, A.R, Photosynthesis in algae. Kluwer, London. 2003; pp 45
11. Chen M, Schliep M, Willows RD, A Red-Shifted Chlorophyll,Science 2010; 329(7):1318–1319.
12. Okelola, F. S. Variation and relationship between seed vigour and yield in rice (*Oryza sativa L.).* M. Agric. Dissertation*.* University of Agriculture Abeokuta, Nigeria 2005.
13. Gitelson AA, Buschmann C, HK, Lichtenthaler, The Chlorophyll Fluorescence Ratio F735/F700 as an Accurate Measure of the Chlorophyll Content in Plants. *Remote* Sensing of Environment. 1999; 69 (3): 296.
14. Demczuk AE, Sacała E, Grzyś Zmiany aktywności syntazy acetylomleczanowej (ALS) pod wpływem herbicydu Titus 25 DF u różnych odmian ogórka. Progr. Plant Post. Ochr. Roślin. 2004; 44(2): 645-647.
15. Sekutowski T, Sadowski J. Use of bioassays for assessment of residues level of herbicides active ingredients in soil. Pesticides/Pestycydy. 2006; (1-2): 59-64.
16. Sinha KK, P. Kumari, Some physiological abnormalities induced by aflatoxinB1 in mung seeds. Mycopathologia. 1990; 110 (9):77-79
17. Adekunle AA, Bassir O, The effects of afatoxinB1 and G1 on the germination and leaf colour of cowpea (*Vigna sinensis*)*.* Mycopathologia et mycologia applicata*.* Klower academic publisher, Ibadan, Nigeria 1997, 1-12.
18. Fratamico PM, Foodborne Pathogens: Microbiology and Molecular Biology. Horizon Scientific Press 2008; 1- 67
19. Crison EV, Effect of Aflatoxin on seedling growth and Ultrastructure in plants. Applied Microbiology. 1997; 26(6):991-1000
20. [Wu Q,](http://informahealthcare.com/action/doSearch?action=runSearch&type=advanced&result=true&prevSearch=%2Bauthorsfield%3A%28Wu%2C+Q%29)  [JezkovaA,](http://informahealthcare.com/action/doSearch?action=runSearch&type=advanced&result=true&prevSearch=%2Bauthorsfield%3A%28Jezkova%2C+A%29)  [Yuan Z ,](http://informahealthcare.com/action/doSearch?action=runSearch&type=advanced&result=true&prevSearch=%2Bauthorsfield%3A%28Yuan%2C+Z%29)  [Pavlikova L,](http://informahealthcare.com/action/doSearch?action=runSearch&type=advanced&result=true&prevSearch=%2Bauthorsfield%3A%28Pavlikova%2C+L%29)  [Dohnal V,](http://informahealthcare.com/action/doSearch?action=runSearch&type=advanced&result=true&prevSearch=%2Bauthorsfield%3A%28Dohnal%2C+V%29) Kuca K, Biological degradation of aflatoxins. Drug reviews. 2009, 41 (1) pp1-7
21. Sekutowski T, Sadowski, J, PHYTOTOXKITTM microbiotest used in detecting herbicide residue in soil. *Environ.* Prot. Eng.2009; 35(1): 105-110.
22. Mahoney N, Russell JM, “A rapid analytical methods for determination of aflatoxins in aflatoxins in plant-derived dietary supplement and cosmetic oils” *J. Agric* Food Chem*.* 2010; 58 (7): 4065 - 4070.
23. Ahmad I, Crawford G, Trace residual analysis of the sulfonylurea herbicide chlorsulfuron in soil by gas chromatography-electron capture detection. Journal Agric. Food Chem. 1990; 38: 138-141.
24. Ali AO, Ahmed MI, Elkhalifa EA, The utilization of seinat seeds flour as sesame seeds substitute in the production of Sudanese traditional food Khemiss-tweria. Proceedings of the 4th International Conference on Innovation in Food Science and Nutrition: Future Challenges, September 27-29, 2010, NRC, Cairo, Egypt 2010; 1-25

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