### Toxicological Evaluation of Graded Levels of Areca catechu seed flour on Performance of Albino Rats

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Abstract: The toxicological evaluation of graded levels of *Areca catechu* seed flour (ACSF) on albino rats was investigated in order to determine its suitability as an additive in feed supplement. The proximate analysis of ACSF showed that it had high carbohydrate and crude fibre values of  $79.18\pm0.64\%$  and  $34.25\pm0.01\%$  but low protein, moisture, and ash contents of  $4.20\pm0.08\%$ ,  $6.79\pm0.02\%$ , and  $1.76\pm0.03\%$  respectively. The ACSF also contains iron, magnesium, copper and zinc at high concentrations. The result of the phytochemical analysis revealed that ACSF contains tannins, flavonoids and alkaloids. Twenty eight rats with an average weight of 40-60g were randomly allotted to four dietary treatment containing 0, 10, 20 and 30% of ACSF incorporated into the feed. During the rat experiment which lasted for eight weeks, rats in both the control and experimental groups appeared to suffer no major toxicological effect and weekly monitoring showed good physical appearance. Histopathological analysis of the organs of the rats showed no visible lesion in both the kidney and liver in the 10% incorporation. ACSF might be a good additive in feed supplemented at 10% level of inclusion.

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Keywords: Areca catechu seeds; mineral element; proximate; phytochemical; rats feed and toxicological evaluation

### Introduction

Various parts of African savanna and forest tree species are used for food, wood or traditional and medicinal purpose. Still the lack of chemical knowledge for many species impedes a rational use of this biomass to contribute to sustainable development (Ambe, 2001; Cook *et al.*, 2000).

*Areca* nut (*Areca catechu*), a family of Arecaceae, popularly known as betel nut, is one of the oldest known masticatories amongst Asia (Trease & Evan, 2009). It is the seed of the Areca palm which grows in much of the tropical Pacific, Asia, and parts of east Africa and even in Nigeria. It is a medium sized tree growing to 30m tall and with a trunk having 20-30cm in diameter. The leaves are 1.5-2cm long, pinnates, with a rigid and segments closely packed. Fruits are hard, ovoid, red-orange colored. They possess a fibrous mesocarp and a thin woody endocarp enveloping a seed (William et al., 2002).

The chemical constituents of Areca nut are mainly polyphenols including tannins, flavonoids and nine closely related alkaloids (George et al., 2006) which include arecoline, arecaidine, arecaine, arecolidine. guvacine. isoguvacine, guvacoline. conine, norarecoline to pyridine group 6,7 besides carbohydrates, fats and minerals. Among the alkaloids, arecoline (0.12-0.24% in ripe nut) has been reported to be the main alkaloid having cholinergic muscarinic agonistic activity and mostly present in the seed as salt of tannic acid (Jayalakshmi et al., 1982). Areca catechu extract has being reported to have central nervous stimulating effect which is mostly due to presence of these alkaloids (Dhinegra & Sharma, 2006; Azeez *et al.*, 2007). It was cited for its various medicinal properties, specially antibacterial and antiviral activities (Reena *et al.*, 2009).

Studies have been conducted on the use of a "pure" paan preparation: areca nut, betel leaf, and lime only. One animal study that was carried out in 1989 found that unprocessed areca nuts, even at high doses, displayed only a very weak carcinogenicity in mice, whereas use of processed areca nuts, as commonly used in paan preparations, caused cancer (Rao et al., 1989). Since 1971, many studies have showed areca nut extracts to cause cancer in rodents, and other study further determined that paan, when consumed with and without tobacco, increased oral cancer risk by 8.4 and 9.9 times, respectively compared to those who do not consume paan. Areca nut powder extract has been reported to be capable of reducing silver ions to silver nanoparticles, which may be useful as antimicrobial agents (Bath et al., 2012).

In rural communities and regions where there is food deficit, it is used as an appetite suppressant (Strickland *et al.*, 2003). Lopez-Vilchez *et al.* (2006) reported a case of neonatal withdrawal syndrome in an infant born to a woman who was a chronic *A. catechu* nut user. Arecoline, the principal neuroactive alkaloid in *A. catechu* seed, was found in the mother's placenta. The habit of chewing *A. catechu* nut independently contributes to the risk of both hyperglycaemia and type-2 diabetes in Taiwanese men. This association is dose-dependent with respect to the duration of *Areca* nut use and the quantity of *areca* nut chewed per day (Tung et al., 2004). The utilization of ACSF in livestock diets has not been investigated. This study therefore aims at carrying out toxicological evaluation of graded levels of Areca catechu seed flour on performance of albino rats.

#### **Materials and Methods** Materials

Mature Areca catechu seeds were picked from the trees within the University of Ibadan, Ibadan in Ovo State, Nigeria. The thin layers covering the seeds were manually decorticated to remove the kernels. Only healthy looking kernels, without infection or damage, were chosen for the analysis. The kernels of Areca catechu after being dried under the sun were grounded using a mechanical grinder to give the coarse seed flour. The flour obtained was further dried under the sun, reduced to a fine powder and then stored at room temperature until needed for analysis.

# **Proximate analysis**

Moisture, crude fat, crude protein, ash and crude fibre contents of ACSF and the compounded feeds were analyzed using the method of AOAC (1990). Carbohydrate content was determined by difference [100 - (moisture content + crude protein + crude fat + ash + crude fibre)] (Ajayi et al., 2013).

# Analysis of mineral elements

Mineral composition of ACSF was determined following the method used by Ajayi et al. (2006) and Idouraine et al. (1996). 1g of the seeds was ashed in a muffle furnace at 550°C for 5h until a white ash was obtained. The minerals were extracted from ash by adding 3ml of concentrated HNO<sub>3</sub> (63%). The digest was carefully filtered into 100ml standard bottle and made up to mark with deionized water. All the minerals were estimated with the use of an atomic absorption spectrophotometer (Perkin Elmer model 703, USA) with the exception of sodium which was determined by flame photometric method.

## **Phytochemical screening**

ACSF was subjected to phytochemical screening for the identification of carbohydrates (Molisch test), (Fehling's reducing sugars test). alkaloids (Dragendorff's, Mayer's, and Wagners's tests), glycosides (hydrolysis test), flavonoids (ammonia and aluminium chloride tests), saponins (using frothing and emulsion), tannins (ferric chloride and lead acetate tests), steroids and terpenoids following the methods of Ajavi et al. (2011) and Harbone (1984).

# Feed compounding

Four groups of animals were used for this study. The control rats were fed with diet containing 0 % ACSF while the other groups were fed with feed containing ACSF incorporated at different levels (10%, 20% and 30%) i.e. the diet of the test groups is

the control diet substituted with ACSF at 10%, 20% and 30% (Table 2). The diets were prepared according to the procedure described by Souza et al. (2007) with slight modification. Each of the diet was pelletized, dried for two days and packed in four separate transparent plastic buckets.

# Animal, diets and feeding

Twenty eight Wister rats of about eight weeks old weighing between 45g-60g were bought from the animal house, Veterinary Extension Unit, University of Ibadan, Nigeria. The rats were grouped according to their body weight into four groups of seven rats each, housed in perforated plastic cages for the study. They were weighed at the beginning of the experiment as zero (0) day. Their feed intake was monitored daily while their body weight was noted weekly. The rats had access to feed and water ad-libitum throughout the period of the study.

### Analysis of Haematological and biochemical parameter

3ml of rat blood was collected into Ethylene Diamine Tetra Acetic Acid bottles through ocular puncture and used for haematological analyses. The packed cell volume, haemoglobin concentration, red blood cell, white blood cell, differential counts, mean corpuscular volume corpuscular and mean haemoglobin were concentration determined according to the methods of Jain (1986). Albumin and globulin were determined by colorimetry. The albumin/globulin ratio was obtained by dividing the calculated albumin value by the calculated globulin value. Aspartate aminotransferase and alanine aminotransferase were also determined (MAFF, 1984). **Tissue pathology** 

Small portions of heart, liver, kidney, lungs and brain were harvested and stored in 10% formalin for histological analyses. These tissues were fixed and put through series of dehydration in graded concentration of xylene. They were embedded in wax, sectioned at 5µ and transferred to clean glass slides. The thin sections were stained with haemotoxylin and eosin (H and E) dyes for examination under the light microscope for histological changes following the method described by Jain (1986).

## Statistical analysis

Numerical data obtained from this study were expressed as mean values±standard error. Organ weights. biochemical and haematological determinations were analyzed using two ways ANOVA. A probability level less than 5 % (p < 0.05) was considered significant.

## Results

Tables 1-5 and figures1- are shown below:

Properties	Values
Weight of 20 seeds(g)	75.6
Weight of a seeds(g)	$3.78 \pm 0.33$
Seed length (cm)	$3.58 \pm 0.28$
Seed width (cm)	$2.91 \pm 0.10$
Colour of unripe seed	Green
Colour of ripe seed	Orange
State of the seed	Very hard
Parameters	Designation
Saponin	Present
Tannins	Absent
Flavonoids	Present
Steroids	Present
Cardiac glucoside	Present
Alkaloids	Present
Reducing sugar	Absent
Phenol	Absent
Anthraquinones	Absent
Glycosides	Absent
Phlobatannins	Absent
Parameters	Concentration(mg/Kg)
Potassium	3170.00
Calcium	2170.00
Magnesium	674.00
Sodium	220.00
Iron	74.20
Manganese	18.00
Copper	9.95
Zinc	5.90
Phosphorus	1.25

Table 1. Physical properties/Phytochemical screening/Mineral composition of ACSF

Table 2. Percentage composition of the experimental diets

Ingredients	%	Control (g)	0%	10%	20%	30%
Maize	40	3200	3200	2880	2560	2240
Soy bean	18.21	1,456.8	1456.8	1311.12	1165.44	1019.76
Bone	3.30	264	264	237.6	211.2	184.8
Salt	0.79	63.2	63.2	56.88	50.56	44.24
Groundnut cake	14.2	1,136	1,136	1022.4	908.8	795.2
Corn bran	7.08	566.4	566.4	509.76	453.12	396.48
Palm kernel cake	7.08	566.4	566.4	509.76	453.12	396.48
Oyster shell	2.26	180.8	180.8	162.72	144.64	123.56
Wheat	7.08	566.4	566.4	509.76	453.12	396.48
ACSF		0.00	0.00	800	1600	2400
Total (g)	100	8000	8000	8000	8000	8000

		Control	Experimental		
Parameter	ACSF	0%	10%	20%	30%
Moisture	$6.79 \pm 0.02^{\circ}$	10.14±0.01 <sup>a</sup>	$09.12 \pm 0.02^{b}$	$8.80 \pm 0.06^{b}$	8.39±0.05 <sup>b</sup>
Ash content	$1.76\pm0.03^{\circ}$	$14.10 \pm 0.02^{a}$	15.61±0.02 <sup>a</sup>	10.89±01 <sup>b</sup>	12.18±0.03 <sup>b</sup>
Crude protein	4.20±0.08	19.23±0.05 <sup>a</sup>	20.06±0.13 <sup>a</sup>	16.93±0.5 <sup>b</sup>	15.39±0.13 <sup>b</sup>
Crude fat	$8.40 \pm 0.08^{\circ}$	$15.46 \pm 0.6^{a}$	$13.60 \pm 0.03^{b}$	13.68±0.01 <sup>b</sup>	13.64±0.04 <sup>b</sup>
Crude fibre	34.25±0.01 <sup>a</sup>	$7.580 \pm 0.14^{d}$	$11.01\pm0.03^{\circ}$	15.91±0.04 <sup>b</sup>	15.77±0.03 <sup>b</sup>
Carbohydrate	79.18±0.64 <sup>a</sup>	41.07±0.06 <sup>c</sup>	41.61±0.14 <sup>c</sup>	$49.70 \pm 0.09^{b}$	$47.41 \pm 0.14^{b}$

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Table 3.	Proximate	composition	of ACSF	and ex	perimental	diets

Mean±SD for three replicate analyses

Values in the same row with the same superscripts are not significantly different at (P < 0.05)

Table 4 Serum biochemistr	v and haematological	analysis of rats fed	with of ACSF
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Parameter	0%	10%	20%	30%
Serum biochemistry				
Total protein	$7.86 \pm 0.3^{a}$	6.8±0.79 <sup>b</sup>	6.6±0.2 <sup>b</sup>	6.7±0.10 <sup>b</sup>
Albumin	5.07±0.22 <sup>a</sup>	4.03±0.97 <sup>b</sup>	3.73±0.15 <sup>c</sup>	3.78±0.05 <sup>c</sup>
Globulin	2.33±0.60 <sup>b</sup>	2.8±0.2 <sup>a</sup>	2.3±0.56 <sup>b</sup>	2.9±0.00 <sup>a</sup>
Al/glob	$2.17 \pm 0.57^{a}$	$1.44 \pm 0.46^{b}$	$1.67 \pm 0.35^{b}$	1.3±0.00 <sup>b</sup>
AST	$40.0 \pm 1.00^{b}$	39.33±1.36 <sup>b</sup>	43±1.22 <sup>a</sup>	44.33±0.58 <sup>a</sup>
ALT	28.33±0.94 <sup>c</sup>	28.67±0.58 <sup>c</sup>	30.3±0.58 <sup>b</sup>	32.00 ±1.00 <sup>a</sup>
ALP	80.33±6.67	$79.34 \pm 3.80$	85.00 ±16.52	91.33±9.02 <sup>a</sup>
Urea	14.67±0.71 <sup>a</sup>	14.33±0.58 <sup>a</sup>	14.33±0.58 <sup>a</sup>	14.33±0.57 <sup>a</sup>
Creatine	0.73±0.11 <sup>a</sup>	$0.7 \pm 0.10^{a}$	$0.5 \pm 0.00^{b}$	0.57±0.06 <sup>b</sup>
Haematology				
PVC (%)	48.86±2.54 <sup>a</sup>	46.57±3.10 <sup>b</sup>	41.85±2.26 <sup>c</sup>	44.86±3.63
MCH (%)	20.1±0.53 <sup>a</sup>	19.87±0.69 <sup>a</sup>	20 <sup>b</sup> .00±0.62 <sup>a</sup>	19.84±0.64 <sup>a</sup>
MCHC (%)	33.11±0.66 <sup>a</sup>	33.42±0.70 <sup>a</sup>	33.44±1.18 <sup>a</sup>	33.42±1.04 <sup>a</sup>
MCV (%)	60.72±1.48 <sup>a</sup>	59.37±1.57 <sup>a</sup>	59.81±1.30 <sup>a</sup>	60.18±1.21 <sup>a</sup>
Hb (mg/dl)	16.73±1.05 <sup>a</sup>	15.38±1.07 <sup>a</sup>	14.00±0.79 <sup>a</sup>	15.00±1.30 <sup>a</sup>
RBC (w <sup>6</sup> /ul)	8.14±0.60 <sup>a</sup>	7.75±0.73 <sup>a</sup>	$7.00\pm0.48^{\circ}$	7.56±0.67 <sup>b</sup>
WBC $(w^3/ul)$	8792.85±2794.55 <sup>a</sup>	9957.14±2,457.75 <sup>a</sup>	8464.28±1902.88 <sup>a</sup>	6678.57±3.05 <sup>b</sup>
Lymphocytes (%)	79±19.35 <sup>a</sup>	79.14±12.75 <sup>a</sup>	73.43±19.44 <sup>b</sup>	74.85±7.19 <sup>b</sup>
Neutrocyte (%)	20.57±8.83 <sup>b</sup>	27.54±12.96 <sup>a</sup>	22.00±948 <sup>b</sup>	17.17±7.40 <sup>c</sup>
Eosinocyte (%)	2.00±1.15 <sup>a</sup>	1.28±0.83 <sup>b</sup>	1.14±0.89 <sup>b</sup>	2.00±1.15 <sup>a</sup>
Monocyte (%)	2.43±1.27 <sup>a</sup>	2.00±1.41 <sup>b</sup>	2.00±1.5 <sup>b</sup>	1.86±2.87
Platelet(cell/cu.mm)	149714.28±70,554 <sup>b</sup>	166428.57±57,616 <sup>a</sup>	139857.1±39,180 <sup>c</sup>	111,514±55,820 <sup>b</sup>

Means  $\pm$  SD for seven rats per group AST- Aspartate aminotransferases ALT- Alanine aminotransferases, A/G Ratio- albumin globulin ratio. Values in the same row with the same superscripts are not significantly different at (P< 0.05)

Table 5. Histological result on the kidney and liver of the rats in the control and experimental groups

Control	rol group Experimental group		al group	
Tissues	0%	10%	20%	30%
Kidney	There is moderate cellular infiltration and hemorrhage into the interstitium.	No visible lesions seen.	There is severe portal and central venous congestion.	No visible lesions seen.
Liver	No visible lesions seen.	No visible lesions seen.	No visible lesions seen.	The hepatocytes appear shrunken. There is moderate periportal and diffuse cellular infiltration by macrophages and few lymphocytes.



Fig. 1. A chart showing the weekly feed consumption by rats fed with graded level of ACSF



Fig. 2. A chart showing the weekly body weight of the rats fed with graded level of ACSF



Fig. 3. A chart showing the weight of the brain, liver, kidney, heart, lung, spleen and intestine of the rats fed with graded levels of ACSF



Fig. 4. Photomicrograph of the kidney of a control rat showing moderate cellular infiltration and hemorrhage into the interstitial (x550)



Fig. 5. Photomicrograph of the kidney of 20% rat experimental rat showing severe portal and central venous congestion (x550)



Fig. 6. Photomicrograph of the liver of 30% experimental rat showing hepatocytes that appears shrunken. There is moderate periportal and diffuse cellular infiltration by macrophages and few lymphocytes (x550)

# Discussion

#### **Physical characteristics**

The physical characteristics of *Areca catechu* seeds is presented on Table 1. The seed has an average weight of  $3.78 \pm 0.33$ g; an average length of  $3.58 \pm 0.28$  cm and an average width of  $2.91 \pm 0.10$  cm. The seed is green when unripe but becomes orange when ripe.

#### Phytochemical analysis

ACSF were screened for phytochemical properties and the result on Table 1 showed that it contained saponin, phenols, steroids, flavonoids, cardiac glucoside and carbohydrate. These phytochemicals exhibit diverse pharmacological and biochemical actions when ingested by animals. Alkaloids are known to show medicinal as well as physiological activities (Sofowora, 1993).

#### Mineral elements

ACSF is very rich in potassium, calcium and magnesium (Table 1). Potassium (3170.00 mg/Kg) had the highest value followed by calcium (2170.00 mg/Kg), magnesium (674.00 mg/Kg) and slightly sodium (220.00 mg/Kg). These minerals in the diet are generally required for metabolic reactions, transmission of nerve impulse and rigid bone formation among others (Egwin *et al.*, 2010). Other elements present in ACSF such as iron (74.20 mg/Kg), phosphorus (1.25 mg/Kg), manganese (18.00 mg/Kg), copper (9.95 mg/Kg) and zinc (5.90 mg/Kg) are low.

# Proximate composition

The proximate analysis of ACSF is presented in Table 3. The ash content is  $1.76\pm0.03\%$  and it is higher than the value reported for *Propolis africana* seeds (Aremu *et al.*, 2007). The crude fibre content was found to be  $34.25\pm0.01\%$ . The carbohydrate content is very high; 79.18 $\pm0.64\%$ . This high carbohydrate content and crude fibre values are in accordance with the report of IARC (2004). These values suggest that the seeds could serve as source of

roughage and the suitability of compounding it in animal feed (Abighor *et al.*, 1997). The protein content of ACSF is very low  $4.20\pm0.08\%$ . It is lower in comparison to the values of  $(11-27.8\pm0.04\%)$  reported for wheat flour and defatted wheat germ flour (Muhammad *et al.*, 2007). The ash content,  $1.76\pm0.02\%$  is lower than the value reported for *A. heterophyllus* and *T. africana* (Ajayi *et al.*, 2008), and greater than the values determined for seeds such as kolanut, coconut, and melon seeds (Onyeike & Acheru, 2002). Addition of ACSF resulted in an increase in the ash values of the feed compounded up to  $15.61\pm0.02\%$ , crude protein content up to  $20.06\pm0.13\%$  and fat content up to  $13.69\pm0.03\%$ . Higher values were obtained at the 10% inclusion.

# Feed intake and body weight changes

There was a gradual increase in the quantity of feed consumed by rats in the 4 groups (0%, 10%, 20%) and 30%) throughout the feeding trial that lasted for eight weeks (Figs. 1 & 2). There was steady but not rapid increment in the body weight of the rats during the experiment period. The control (0%) was found to have the highest weight gain followed by the 10% when compared to 20 and 30%.

#### **Physical appearance**

The physical appearance of the rats was normal in all the groups throughout the eight weeks of the experiment. The eyes, mouth and hair of the animals used in their respective groups appeared to be normal throughout the period of the study.

#### Organ weights

The weight of the seven organs collected for tissue pathology did not differ significantly from each other in both groups. An average kidney weight of  $0.99\pm0.19$ g;  $0.84\pm0.5$ g;  $0.85\pm0.10$ g and  $0.82\pm0.09$ g was obtained for the rats in the 0%, 10%, 20% and 30% groups while that of the heart was  $0.60\pm0.15$ g,  $0.62\pm0.27$ g,  $0.48\pm0.15$ g and  $0.48\pm0.07$ g respectively (Fig. 3).

### Haematological and biochemical parameters

The results of the haematological and the biochemical studies of the rats used for the experiment were presented. Platelets had the highest value of 166,428.57±57,616 in 10% against the observed  $149,714.28\pm70,554$  in the control (0%) while all other haematological parameters such as neutrophil, WBC, RBC, HB, PVC, MCHC are also with higher values in 10% incorporation when compared to other levels of incorporations but are similar to that control. The absence of significant differences as shown among the control and 10% groups for the haematological parameters is an indication that ACSF may not be toxic and had no adverse effect on the blood of the rats under study at 10% incorporation. The haematological values from this study are similar to the report given by Vishnu et al. (2010) for the toxicity study of Garcinia mangostana pericarp extract in rats. The serum protein values are comparable to each other in the experimental groups but found to be lower than the control. The AST, ALT, ALP values appear to be increased gradually from 10% inclusion to the highest level of inclusion. These values in 10% inclusion though lower than others but are still comparable to the values obtained for the control group. Alanine aminotransferases enzyme activity examined was 80.33±6.67 in the control group feed as against 79.00±3.80, 85±16.52 and 91.33±9.02 that was obtained for the 10%, 20% and 30% groups respectively. The increase in the activities of ALT, AST and AST in 20% and 30% may probably be attributed to the dietary treatment effect of higher ACSF inclusion levels. It could be that as the inclusion level increased, the tendency of increase in the antinutritional factor in the experimental ingredient increased. Ewuola and Egbunife (2008) observed in the separate studies that increase in ALT and AST are clinical indication of diagnosing state of damage done to visceral organ by toxic substance or infection. Michael et al. (2012) reported a similar case with rabbit fed with kenaf seed meal at different levels of inclusion.

### Histopathology

The histopathology study of the liver and the kidney of the rats used as experimental model revealed no major complications in the tissues of all the rats in the 10% groups (Table 5). For the other test groups, there is moderate periportal and diffuse cellular infiltration by macrophages and few lymphocytes in the liver of the 30% experimental group while no visual lesion were observed for both the 0%, 10%, and 20% respectively. There were severe portal and central venous congestion in the kidney of the 20% group while no lesion was observed in the 10% group. The 10% group was found to be normal during this study

since no visual lesion was observed in both the kidney and liver.

#### Conclusion

ACSF may find usefulness in rat feed for livestock due to its high carbohydrate content. The feed however has to be supplemented with other high protein residue such as groundnut cake because it is in low protein. The presence of tannins, flavonoids and alkaloids in ACSF ascertained that it contains some major compounds that have remarkable biological activities hence it might be helpful in preventing various diseases. The metals that are found in ACSF are all useful in making the body strong. The incorporation of ACSF at 10% level into rat diet did not produce any significant changes in haematological parameters as well as in the heart and kidney of the rats; it can therefore be concluded that ACSF might be a good additive in rat feed at 10% level of inclusion.

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