

Effect of Muscular Exercise Program with and without Antioxidant on Serum Level of S100B Protein in Male Albino Rats as a Marker of Central Exhaustion and Some Related Parameters

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Abstract: The study targeted effect of muscular exercise alone on plasma level of s100B protein level as a marker of central exhaustion which increases permeability of blood brain barrier, and some related factors as serum lactic acid, malondialdehyde (MDA) and total antioxidant capacity. Also, to investigate the effect of muscular exercise with the use of antioxidant on plasma level of s100B protein level and some related factors as serum lactic acid, malondialdehyde (MDA) and total antioxidant capacity. Twenty seven adult male albino rats were divided into 3 equal groups; Group 1: control, Group 2: exposed to muscular exercise program only and Group 3: exposed to muscular exercise program and take vitamin C. The results showed that there were statistically significant differences of measured markers between resting and after the exhaustive muscular exercise bout before the application of training program in all groups, After the application of a 12 weeks training program; During rest, There were statistically significance differences between different groups regarding S100B and TAC only, while the other markers (MDA and LAC) didn't show any significant changes. After the exhaustive exercise bout: There were statistically significant differences between group 1 and the other two groups as regards to LAC, MDA and TAC level. There was no statistically significant difference among the 3 groups in serum level of S100B. But in groups 2 & 3 there were statistically significant differences between resting and exhaustion after application of training program regards all markers (S100B, LAC, TAC, and MDA).

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1. Introduction

S100B is a beta homodimeric protein within the S100 family of calcium-binding proteins, with each beta monomer approximately 10.5 kDa in size (*Gonc et al., 2008*). However, S100B is not specific to the brain, as it has been detected in extracerebral cell types such as adipocytes (*Gonc et al., 2010*), melanocytes, skeletal myofibers and myoblasts (*Tubaro et al., 2010*). S100B can also be found in tissues of the bladder, liver, kidney, colon, lungs, pancreas, tonsil and stomach via immune histochemical staining with antibody detection (*Pham et al., 2010*). Intracellular and extracellular functions of S100B, including a recent review by Michetti et al. (*Michetti et al., 2012*). The intracellular effect of S100Bas it regulates cytoskeleton and cell proliferation and regulates levels of cytosolic Ca² and/ or transducing the intracellular Ca² signal (*Donato, 2003*). Extracellular effect of S100B appears to have a neurotropic role in both development and repair (*Sorci et al., 2010*). It stimulates neurite outgrowth, glial proliferation, and the regeneration of injured nerves (*Iwasaki et al., 1997*). S100B also has a neuro-protective function, in enhancing neuronal cell maintenance, preventing

motor neuron degeneration and enhancing the survival of neurons (*Bhattacharyya et al., 1992*).

However, the possibility of a cerebral origin of S100B in this study still cannot be excluded, as vitamin E may play a role in the maintenance of neurological structure and function. (*Pace et al., 2003*).

A possible relationship between S100B and vitamin E was suggested by *Bialowas-McGoey et al., 2008*, who reported that vitamin E can alter the oxidative state of S100B and affect the expression of an S100B receptor to cause increases in microglial activation. However, the significance of this, and its relationship with S100B levels are still not well studied (*Khanna et al., 2005*).

The aim of this study was to examine the effect of vitamin C as an antioxidant supplementation on serum level of S100B and oxidative stress in rest and after an exhaustive exercise bout pre and after a scheduled exercise training program.

3. Materials and Methods

This study was carried out on 27 adult male albino rats which divided into 3 equal groups as follow; Group 1: Included 9 for control, Group 2: Included 9 exposed to muscular exercise program

only and Group 3: Included 9 exposed to muscular exercise program and take vitamin C.

Serum were collected from all groups before and after the application of exercise program for estimating the level of S100B, Serum MDA, Serum Lactic Acid, Serum TAC.

3. Results and Discussion

The obtained results are shown in tables 1-14 and figures 1-12.

For many years, a physically active lifestyle has been promoted to decrease risks to several chronic diseases by stimulating respiratory, cardiovascular, and immune systems. However, studies regarding the effects of exercise on central nervous system have gained increased interest in applied and basic research. Regular exercise decreases the risks of cognitive impairment related to age and Alzheimer disease in humans. (*Laurin et al., 2001*).

Conversely, it has been postulated that exercise in humans, instead of benefit effects, can induce brain damage. To confirm this hypothesis, the measurement of blood concentration of S100B protein has been acknowledged as a promising tool. An increased number of studies have suggested that the measurement of S100B level in blood and cerebrospinal fluid could be used in clinical practice to evaluate brain injury involving reactive astrogliosis, astro-cytic death, and/or blood-brain barrier dysfunction. (*Portela et al., 2002*).

However, to our knowledge, there are few studies about exercise and S100B peripheral levels. Exhaustive exercise, especially when sporadic, can produce muscle damage and initiate an acute inflammatory response, evidenced by increased cytosolic enzymes in the blood, and sarcolemma and line disruption (*Gomez-Cabrera et al., 2007*).

Growing body of evidence suggests that antioxidant supplementation can be a useful strategy to reduce the harmful effects of exercise induced oxidative stress (*Malaguti et al., 2013*).

Vitamin C is the major water-soluble antioxidant present within the cell and extracellular fluids (*Close et al., 2006*).

The aim of this study was to examine the effect of vitamin C as an antioxidant supplementation on serum level of S100B and oxidative stress in rest and after an exhaustive exercise bout pre and after a scheduled exercise training program.

Serum level of S100B and other oxidative stress markers (MDA, TAC, and lactic acid) was evaluated in all groups, and the relation of their level to rest state and after exhaustion before and after exercise training program with and without the use of Vit.C was assessed.

The obtained data showed a statistically significant high in the level of S100B after exhaustion in pre and post exercise program in the all groups compared to the resting state.

Despite of there was no head trauma occurs during exercise, this increase may be due to direct effect of exercise on BBB causing disruption in permeability or due to activation or exercise-induced damage of peripheral tissues containing S100B will likely lead to an increase in serum S100B (*Marchi et al., 2003*).

The obtained data nearly coincides with the results of *Otto et al.(2000)*, *Straume-Naesheim et al.(2008)*, *Bailey et al.(2011)*, *Michetti et al.(2011)*, *Neselius et al.(2012)* and *Stocchero et al.(2014)*.

However, several studies have also failed to show this relationship as *Saenz et al.(2006)*, *Chevront et al.(2008)* and *Stavrinou et al.(2011)*.

There is large variability in the studies conducted to investigate this relationship with regards to their hypotheses, type of exercise activity, sample collection methods. etc. Furthermore, changes in S100B have been attributed to different causes and are not limited to changes in BBB permeability alone. These causes may include:-

Firstly:- trauma to the head (in contact sports like boxing, soccer, basketball, etc.) (*Otto et al., 2000*, *Straume-Naesheim et al., 2008*, *Graham et al., 2011* and *Neselius et al., 2012*).

Secondly:-Chronic exposure to hypoxic conditions in breath-hold divers (*Andersson et al., 2009*).

Thirdly:-Decompression after scuba diving (*Stavrinou et al., 2011*) and,

Fourthly:-Its release from extra-cranial sources (*Hasselblatt et al., 2004*).

The obtained data showed also non-significant differences in the level of S100B between resting pre-exercise program state and resting post-exercise program state. In group 2, exposed to muscular exercise program only the mean level of S100B at pre exercise rest was 3.11 ± 0.70 respectively and after post exercise rest was 2.76 ± 0.37 respectively. In group 3 which exposed to muscular exercise program and take vitamin C, the mean of S100B level at pre exercise rest was 3.12 ± 0.52 and after post exercise rest was 2.59 ± 0.36 respectively. This finding may contributes to antioxidant effect of exercise program and use of Vit.C in group 3.

This result doesn't matche with the result founded by *St°alnacke et al. (2003)* and *Michetti et al. (2011)* who noticed high S100B baseline concentrations that exceeded the upper limit for a normal healthy adult population in professional sportsmen. This implies that the disruption in BBB as a result of exercise may be greater in professional sportsmen.

On comparing between pre and post exercise exhaustion markers in 2 groups, the obtained data showed a statistically significant low in the level of S100B after exercise program as follow: in group 2: pre exercise exhaustion was (3.56 ± 0.54) ng/ml and post exercise exhaustion was (3.32 ± 0.47) in group 3: pre exercise exhaustion was (3.89 ± 0.50) ng/ml and post exercise exhaustion was (3.05 ± 0.34) ng/ml. This decrease may be due to effect of exercise training on oxidative stress state that generally decreased following exhausting exercise and the use of antioxidant in group 3, as will be discussed later.

In comparison between group 2 and group 3 according to S100B level, the obtained data showed that degree of increase in S100B level in group 3 which received Vit.C is less than that of group 2. This result is matched with *Schulpis et al. (2007)* who used a-tocopherol (vitamin E) supplementation as an antioxidants there was reduction in the response of S100B to exercise. Vitamin E is known to be an antioxidant, shown to reduce the oxidative stress caused by exercise. The S100B response to exercise seems to be partially attenuated after 30 days of vitamin E supplementation, suggesting that higher levels of antioxidant may determine the levels of S100B released during exercise. While the S100B response is reduced after vitamin E supplementation, baseline S100B was unaffected. A possible relationship between S100B and vitamin E was also suggested by *Bialowas-McGoey et al. (2008)* who reported that vitamin E can alter the oxidative state of S100B and affect the expression of an S100B receptor. However, the significance of this, and its relationship with S100B levels are still not well studied.

There is still much discussion revolving around the exact mechanisms of S100B release during exercise and its implications (*Donato et al., 2013*).

Four main causes of S100B release are under investigation (increases in BBB permeability, head injury, skeletal muscle injury and lipolysis), and it is acknowledged that each of these may contribute to S100B elevations in varying proportions and in different situations. Care must therefore be taken when interpreting S100B levels after exercise. In the absence of substantial head injury, skeletal muscle injury and lipolysis, S100B elevations might then be a good marker for the detection of increased BBB permeability. Implications of this S100B increase are also unclear, but it has been pointed out that S100B is not simply a marker for brain injury, or a compound involved in neurotoxic processes. It is recognized that S100B is also involved in several extracellular signaling pathways that may be beneficial in initiating repair and regeneration processes in the brain

(*Schulte et al., 2013*) or in injured skeletal muscle (*Donato et al., 2013*).

The obtained data showed that both pre-exercise and post-exercise exhaustion significantly increased serum MDA levels, thereby confirming the role of exercise in free radical production and consequent oxidative stress. Lipid peroxidation can cause decrease in cell membrane fluidity, inability to maintain ionic gradient, cellular swelling, and tissue inflammation. All these changes modulate a variety of cellular processes, eventually leading to limited muscle contraction (*Thirumalai et al., 2011*).

Samples of pre exercise program showed a statistically significant increase mean level of serum MDA in the 3 groups after an acute bout of exercise until exhaustion pre-exercise program. In group 1 the mean MDA level before exhaustion was (11.4 ± 2.2) $\mu\text{mol/L}$ and after exhaustion was (21.04 ± 2.8) $\mu\text{mol/L}$, in group 2 before exhaustion was (11.3 ± 1.1) $\mu\text{mol/L}$ and after exhaustion was (21.1 ± 3.5) $\mu\text{mol/L}$, and in group 3 before exhaustion was (12.1 ± 1.3) $\mu\text{mol/L}$ and after exhaustion was (22.7 ± 3.2) $\mu\text{mol/L}$. This is matched with *Kostka et al. (2000)* and *Miyazaki et al. (2001)*. Similarly, *Child et al. (2000)* found an increase in MDA of about 40% immediately after a half marathon. *Lekhi. (2007)* who found a significant increase in the content of MDA and uric acid in serum after an exhaustive endurance exercise. Also, *Mohammad et al. (2014)* found statistically significant increase in the levels of serum MDA after the bout of intense exercise (p value < 0.05). *Spirlandeli et al. (2014)* and *Ljiljana et al. (2015)* found increase of MDA after acute exercise.

However *Marta et al. (2015)* found that the physical exercise was unable to change MDA level. This difference might be explained by the type, time, and intensity of exercise.

Not all studies reported increases in MDA in response to exercise *Viinikka et al. (1984)*, *Niess et al. (1996)* measured plasma levels of MDA in trained and untrained individuals at rest, before and after an exhaustive bout of exercise. They found no significant increases in MDA in either group following a treadmill test to exhaustion, either at 15 min post-exercise or 24 h post-exercise.

Also, *Duthie et al. (1990)* and *Dufaux et al. (1997)* found that moderately trained subjects who ran for 2.5 h on a treadmill showed no change in plasma MDA. Similarly, there were no documented changes at rest, before or after 4 weeks of high intensity rowing training plasma MDA levels as found by *Dernbach et al. (1993)* in athletes, and *Alessio et al. (2000)* found no change in plasma MDA after repeated isometric contractions. In addition, *Dixon et al. (2006)* has studied the effects of moderate-intensity whole-body resistance exercise

and found that it had no effect on serum MDA concentration.

The obtained data in the present study showed a significant difference in the increase of MDA in group 2 and 3 before and after exercise training. In group 2, the level of MDA after post exercise exhaustion was (18.3 ± 2.4) $\mu\text{mol/L}$ and after pre exercise exhaustion was (21.1 ± 3.5) $\mu\text{mol/L}$. In group 3 the level of MDA after post exercise exhaustion was (15.9 ± 2.18) $\mu\text{mol/L}$ and after pre exercise exhaustion was (22.7 ± 3.2) $\mu\text{mol/L}$. This mean that exercise program improve of the level of MDA in group 3 with a difference of

This is in agreement with Miyazaki et al. (2001) who reported a decrease in MDA after regular exercise program. There was a smaller increase in erythrocyte MDA in response to the exercise bout post-training compared to pre-training. Also, Aysun et al. (2013) in his study, the MDA was found to decrease after regular exercise.

Moreover, decreased levels of MDA in response to exercise have also been reported in highly trained skiers and runners immediately following exercise to exhaustion Hubner-Wozniak et al. (1994) and Rokitzki et al. (1994a). So, strenuous endurance training was shown to reduce indices of oxidative stress following exhausting exercise.

In group 3, which received Vit.C, there was significant decrease in the mean level of MDA (15.9 ± 2.18) $\mu\text{mol/L}$ compared with group 2 exposed to exercise program only (18.3 ± 2.4) $\mu\text{mol/L}$. This result was in agreement with some studies done by Babaei et al. (2009), Mandana et al. (2014), Popovic. (2015) and Leila and Mohammad. (2016). There was a significant reduction in MDA serum concentration of the training group treated with anti-oxidants supplementation compare to control group not received antioxidant supplementation.

However, Goldfarb. (2005) did not observe any significant difference between the MDA level of the exercise group receiving vitamin C supplementation and its control group.

A variety of sports activities lead to oxidative stress and oxidative damage to lipids, and subsequent production of lipid peroxidation through increased production of reactive species (Fisher-Wellman and Bloomer., 2009). Long-term exercise decreases blood cholesterol and plasma LDL. Therefore, part of the reduction in the levels of MDA may be due to reduced availability of fatty acids (Michalczyk et al., 2008).

In general, three factors may be involved in this reduction:

firstly: a decrease in the production of free radicals in the body.

secondly: an increase in the activity of antioxidant enzymes.

and thirdly: a balancing of oxidants against antioxidants in the body.

In addition, by reviving free radicals and turning them into ascorbic acid radicals, vitamin C prevents oxidative stress and MDA increase, resulting in a negative correlation between plasma ascorbic acid (PAA) and MDA (Belviranli and Gokbel., 2006).

The obtained data noticed that mean serum level of TAC statistically significantly increased after an acute exhaustive bout before training exercise in all groups. In group 1 before exhaustion was (1.11 ± 0.13) mg/dl and after exhaustion was (2.25 ± 0.41) mg/dl. In group 2 before exhaustion was (1.08 ± 0.11) mg/dl and after exhaustion was (2.38 ± 0.62) mg/dl. In group 3 before exhaustion was (1.09 ± 0.17) mg/dl and after exhaustion was (2.35 ± 0.64) mg/dl. This increase may contribute to uric acid, which is considered as an important component of the antioxidant system, and is responsible for one third of the TAC increase. Another possible explanation for the elevation of TAC is changes in other antioxidants, for example, GSH, melatonin, or the antioxidant enzymes (Yu, 1994).

The previous result matching with that of **Sekineh et al. (2014)** who observed that plasma TAC levels increased significantly after the eccentric exercise. he explain this increase probably due to cellular damage caused by eccentric exercise, release of intracellular- muscle enzymes to the blood and high concentrations of GSH.

Nuttakaan et al. (2005) found that athletes who have regular exercise showed the benefit of exercise by gaining a higher total antioxidant capacity, lower oxidation of protein and lipid and moreover, against the free radicals from exhaustive exercise better than in non-exercised people.

The increase in total antioxidant capacity of plasma after the exercise was also consistent with that of Child et al. (1998). They found that plasma total antioxidant capacity increased from 475 ± 84 (mmol/l) to 564 ± 11 (mmol/l) after the half-marathon in 17 male runners. The increase in plasma TAC levels is due to the increase in the levels of creatine kinase in the blood which consequently leads to high concentration of GSH as one of TAC factors. Also, **Möller et al. (1996)** and Liu et al. (1999) observed increased levels of plasma total antioxidant capacity after acute or exhausting exercise.

Diaz et al. (2011) stated that a short duration bout of submaximal exercise elicited a significant increase in TAC levels 60 and 120 minutes post-exercise.

However TAC has been shown to remain unchanged following submaximal bouts of aerobic

exercise in some studies **Tozzi-Ciancarelli et al. (2002)**, **Waring et al. (2003)** and **Watson et al. (2005)**.

In this study, there is elevation in the level of TAC in both groups 2 and 3 exhaustion after training compared with exhaustion before training. In group 2 TAC level after pre-exercise exhaustion was 2.38 ± 0.62 and after post exercise exhaustion was 3.17 ± 0.61 . In group 3 the mean level after pre-exercise exhaustion was 2.35 ± 0.64 and after post exercise exhaustion was 3.41 ± 0.27 .

In matching with our result **Brites et al. (1999)** and **Mukherjee and Chia. (2009)** reported that regular sports training can increase plasma total antioxidant capacity as seen in their two independent studies with soccer athletes.

The increase in serum antioxidant capacity (TAC) after regular exercise is also confirmed by other studies as **Sahlin et al. (2010)**, who noticed that after one week of exercise training a progressive decreasing on oxidative stress was observed, whereas the TAC remained at higher levels.

Also **Czepluch et al.(2011)** found that exposure of healthy sedentary subjects to strenuous physical exercise has been associated with inhibition of macrophage migration due to TGF- β activation, and this inhibitory effect on migratory activities of macrophages was associated with increased total antioxidant capacity.

Some studies show the effects of moderate physical activity on increasing TAC or activity of antioxidant enzymes as **Lesgards et al.(2002)**, **Fatouros et al.(2004)** and **Nuttakaan et al.(2005)** who concluded that athletes, who have regular exercise, showed the benefits of exercise by gaining a higher total antioxidant capacity.

Cesari et al. (2004) also found a positive correlation between the concentration of antioxidants in the serum and the level of physical performance in the elderly.

Also, both single intense exertion (half-marathon run) and regular endurance training were connected with higher antioxidant capacity **Child et al.(1998)** and **Child et al.(1999)**.

The results of other studies do not confirm any positive relation between physical activity and TAC, as with **(Kostka et al., 1998 and Kostka et al., 2000)**. Also, **Afzalpour et al. (2008)** did not report any increase in TAC, but observed positive association between TAC and VO₂max in a study with untrained subjects submitted to anaerobic training over eight weeks. **Jackson et al.(2010)** also found that exercise decrease level of TAC.

The obtained data showed that the increase in the level of TAC post exercise exhaustion in group 3 exposed to exercise program with receiving Vit.C

was 3.41 ± 0.27 more than that of group 2 which exposed to exercise program only was 3.17 ± 0.61 .

The obtained data is in agreement with **Leila and Mohammad. (2016)**, who found supplementation of vitamin C in the control group, resulted in a significant increase in the content of TAC compared to the no-supplement training group.

Also, this increase agreed with the results of **Babaei et al.(2009)**, **Nour-Shahi et al. (2010)** and **Sari-Sarraf.(2013)**.

Vitamin C supplementation increases the levels of serum ascorbic acid and subsequently increased TAC (**Lee et al., 2009**). Taking a vitamin C supplement can lead to an increase in TAC by donating electrons to vitamin E and restoring this fat-soluble antioxidant. This antioxidant action of vitamin C occurs following oxidative stress and prevents further lipid peroxidation on the surface of the cell membrane (**Evansm, 2000**).

Also **Bohlooli et al. (2015)** studied the effect of spinach supplementation as antioxidants on exercise-induced oxidative stress. They found that TAC significantly elevated after supplementation ($P < 0.05$) and lowering level of MDA ($P < 0.05$).

The obtained data point to a significant increase in the lactic acid concentration after the exercise in all groups.

This result corroborating with previous studies **Thomas et al.(2004)**, **Beneke et al.(2005)**, **Van.(2010)**, **Machado et al.(2013)** and **Filipe et al.(2015)**. This increase represents the utilization of the glycolytic system, for the lactate presents itself as an important mediator of the anaerobic metabolism of energy resynthesizes (**Robergs et al., 2004**).

There was decrease in the level of lactic acid exhaustion after training compared with exhaustion before training. This decrease may be due to antioxidant effect of regular exercise on ROS production. Generally muscles always produce lactate, even at rest ($0.8 - 1.5$ mmol/L), but lactate increases incrementally with exercise intensity. At a certain intensity lactate increases exponentially, this is called the lactate threshold. Training-induced adaptations include a lower blood lactate concentration at any given workload (**Overgaard et al., 2010**).

The obtained data showed that the degree of increase of lactic acid after post exercise exhaustion in group 3 who received vit.c supplementation was less than that of group 2. Previous reports have demonstrated an increased level of ROS during high-intensity exercise (**Viña et al., 2000**). Another study found that ingestion of an antioxidant substance inhibited the production of ROS (**Zembron-Lacny et al., 2006**). Although the details of the mechanism behind the correlation between the blood lactate and

ROS levels remain unclear, we hypothesize that the antioxidant function of Vit. C suppresses the increase in the blood lactate concentrations during high-intensity exercise.

The obtained data matched with **Morimasa et al. (2013)** who investigated the anti-fatigue effects of *A. sieboldianus* (antioxidant substances) by subjecting rats to exercise at several levels of intensity and found that *A. sieboldianus* intake before high-intensity exercise decreases the blood level of lactate. This finding suggests that *A. sieboldianus* intake could help to alleviate fatigue during high-intensity exercise. Their result demonstrates a novel activity of *A. sieboldianus* in addition to its previously reported antioxidant and hyperglycemia-

suppressing properties (**Tabuchi et al., 2004; Yamada et al., 2005**). They infer that inhibiting blood lactate elevation during performing high-intensity exercise by ingesting this plant as a dietary supplement could suppress production of reactive oxygen species (ROS), improve metabolic capability, and prevent an excessive stress response.

Exercise is one of the internally generated sources of free radicals. Endurance exercise can increase oxygen utilization from 10 to 20 times over the resting state. This greatly increases the generation of free radicals. Prompting concern about enhanced damage to muscles and other tissues, for that athletes need to take extra antioxidants to defend against the increased free radicals resulting from exercise.

Table (2): Comparisons of the differences in the measured markers during rest before the application of the exercise training program between the different groups. (n = 9).

Variables	resting, pre training program			p-value	Sig.
	G1 (Control)	G2 (ms.train.)	G3 (ms.train. +vit C)		
	Mean ± SD	Mean ± SD	Mean ± SD		
S100B (ng/mL)	2.9±0.49	3.11±0.70	3.12±0.523	0.8	NS
LAC(mmol/L)	1.26±0.20	1.14±0.12	1.09±0.22	0.2	NS
MDA(μmol/L)	11.4±2.2	11.3±1.1	12.1±1.3	0.5	NS
TAC(mg/dl)	1.11±0.13	1.08±0.11	1.09±0.17	0.9	NS

N= number of rats in each group NS = non-significant

Table 2: Illustrates that there were no statistically significance differences (p-value > 0.05) between different groups in all markers (S100B, LAC, MDA, and TAC level) in rest before the application of the exercise training program.

Table (3): Comparisons of the differences in the measured markers after the exhaustive muscular exercise bout before the application of the exercise training program between the different groups. (n=9).

Variables	Exhaustive exercise, pre training program			p-value	Sig.
	G1 (Control)	G2 (ms.train.)	G3 (ms.train.+vitC)		
	Mean ± SD	Mean ± SD	Mean ± SD		
S100B (ng/mL)	3.57±0.24	3.56±0.54	3.89±0.50	0.2	NS
LAC(mmol/L)	4.23±0.38	4.34±0.88	4.53±0.44	0.6	NS
MDA(μmol/L)	21.04±2.8	21.1±3.5	22.7±3.2	0.4	NS
TAC(mg/dl)	2.25±0.41	2.38±0.62	2.35±0.64	0.9	NS

N= number of rats in each group. NS = non-significant

Table 3: Illustrates that there were no statistically significance differences (p-value > 0.05) between different study groups in all markers (S100B, LAC, MDA, and TAC level) after the exhaustive muscular exercise bout.

Table (4): Comparisons of differences in the measured markers in resting and exhaustive conditions before the application of the training program in group 1. (n=9)

Variables	Group 1		% change	p-value	Sig.
	Resting Mean ±SD	Exhaustive exercise Mean±SD			
S100B(ng/dl)	2.9±0.49	3.57±0.24	23.1	0.03	S
LAC(mmol/L)	1.26±0.20	4.23±0.38	235	<0.001	S
MDA(μmol/L)	11.4±2.2	21.04±2.8	84.56	<0.001	S
TAC(mg/dl)	1.11±0.13	2.25±0.41	102.7	<0.001	S

N= number of rats in each group S = significant

Table 4: Illustrates that there was statistically significant increase (p-value <0.05) in serum level of different markers (S100B, LAC, MDA, and TAC level) after the exhaustive exercise bout when compared to resting condition in group 1 (control) before the application of training program.

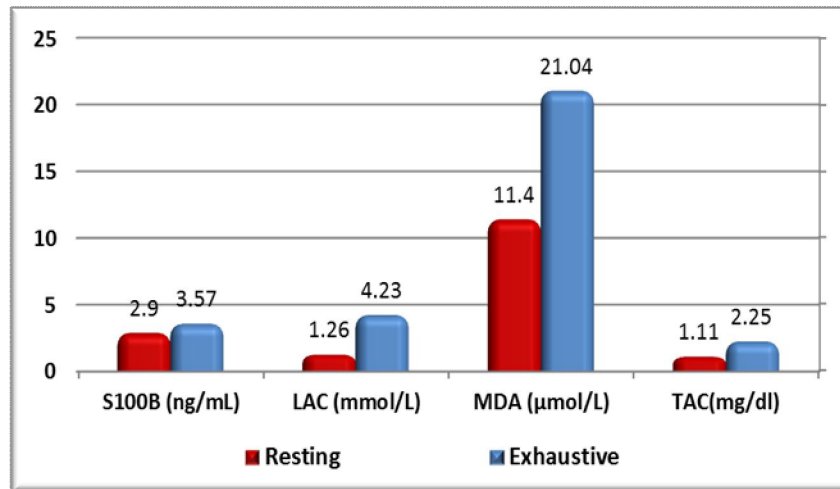


Figure (3): Comparison of different markers under resting and exhaustive conditions in group 1 before the application of training program.

Table (5): Comparisons of differences in the measured markers in resting and exhaustive conditions before the application of the training program in group 2. (n=9)

Group 2 Variables	Resting	Exhaustive exercise	% change	p-value	Sig.
	Mean ±SD	Mean ±SD			
S100B (ng/mL)	3.11 ± 0.70	3.56 ± 0.54	14.47	0.02	S
LAC (mmol/L)	1.14 ± 0.12	4.34 ± 0.88	280.7	<0.001	S
MDA (μmol/L)	11.3 ± 1.1	21.1 ± 3.5	86.73	<0.001	S
TAC (mg/dl)	1.08 ± 0.11	2.38 ± 0.62	120	<0.001	S

N= number of rats in each group S = significant

Table 5: Illustrates that there was statistically significant increase (p-value <0.05) in serum level of different markers (S100B, LAC, MDA, and TAC level) after the exhaustive exercise bout when compared to resting condition in group 2 before the application of training program.

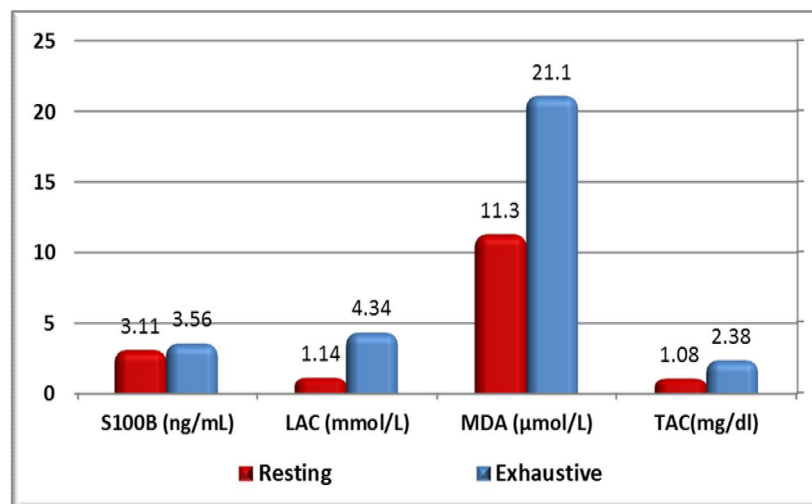


Figure (4): Comparison of different markers under resting and exhaustive conditions in group 2 before the application of training program.

Table (6): Comparison of differences in the measured markers in resting and exhaustive conditions before the application of the training program in group 3. (n=9)

Group 3 Variables	Resting	Exhaustive exercise	% change	p-value	Sig.
	Mean ±SD	Mean ±SD			
S100B(ng/dl)	3.12±0.52	3.89±0.50	24.68	0.02	S
LAC(mmol/L)	1.09±0.22	4.53±0.44	315.6	<0.001	S
MDA(µmol/L)	12.1±1.3	22.7±3.2	87.6	<0.001	S
TAC(mg/dl)	1.09±0.17	2.35±0.64	115.6	<0.001	S

N= number of rats in each group S = significant

Table 6: Illustrates that there was statistically significant increase (p-value <0.05) in serum level of different markers (S100B, LAC, MDA, and TAC level) after the exhaustive exercise bout when compared to resting condition in group 3 before the application of training program.

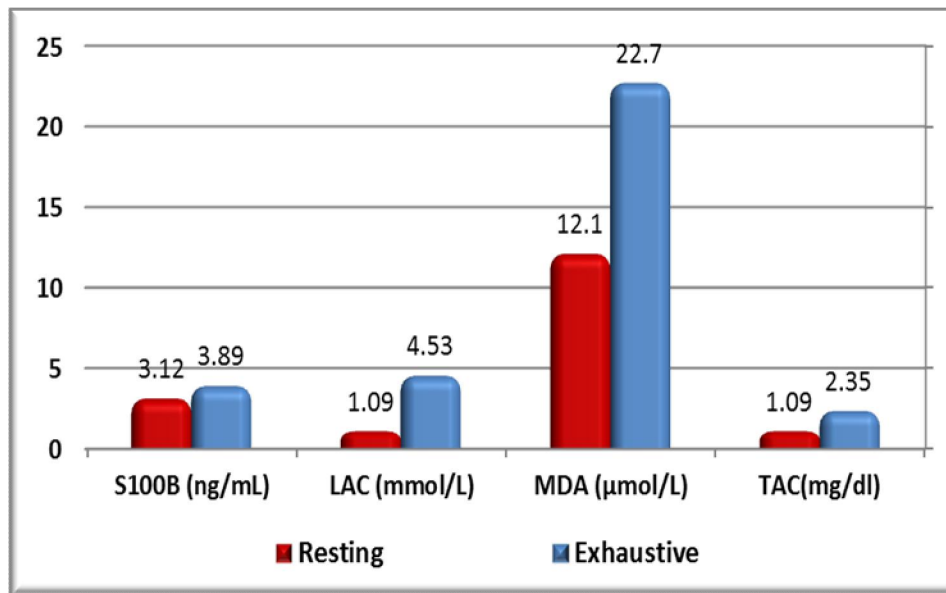


Figure (5): Comparison of different markers under resting and exhaustive conditions in group 3 before the application of training program.

Table (7): Comparison of the changes in the measured markers during rest after the application of the exercise training program between the different groups. (n = 9).

Variables	Resting, post training program			p-value	Sig.
	G1 (Control)	G2 (ms training)	G3 (ms.train+vitC)		
	Mean ± SD	Mean ± SD	Mean ± SD		
S100B ng/ml	2.9±0.49	2.76±0.37	2.46±0.36	0.05	S
LAC(mmol/L)	1.26±0.20	1.34±0.31	1.12±0.19	0.1	NS
MDA(µmol/L)	11.4±2.2	11.1±1.4	11.2±2.04	0.9	NS
TAC(mg/dl)	1.11±0.13	1.36±0.24	1.36±0.27	0.02	S

N= number of rats in each group NS = non-significant S = significant

Table (7): Illustrates that there were statistically significance differences (p-value <0.05) between groups 2 and 3 in S100B and TAC levels during rest after application of the training program. There were no statistically significant differences (p-value >0.05) between different study groups in LAC, and MDA levels during rest after application of the training program.

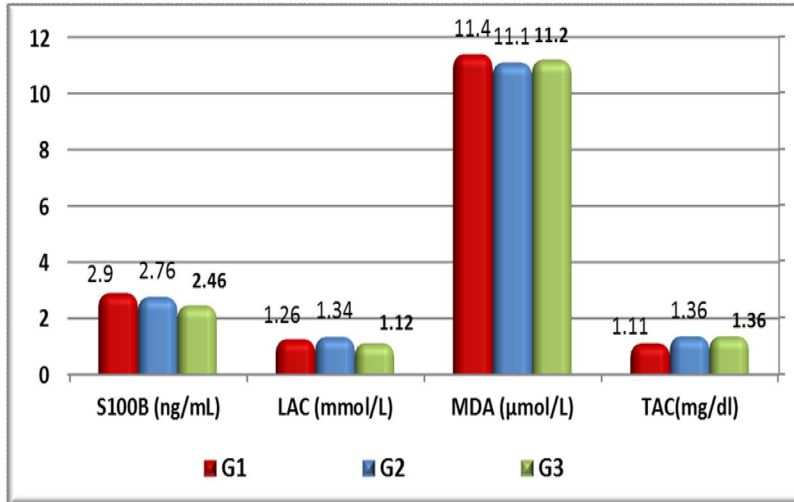


Figure (6): Comparison of the changes in the measured markers during rest after the application of the exercise training program between the different groups.

Table (8): Comparison of the changes in the measured markers following the exhaustive exercise bout after the application of the exercise training program in the different groups. (n = 9).

Variables	Exhaustive exercise, post training program			p-value	Sig.
	G1 (Control)	G2 (ms.train)	G3 (ms.train+vitC)		
	Mean ± SD	Mean ± SD	Mean ± SD		
S100B (ng/mL)	3.45±0.17	3.32±0.47	3.05±0.34	0.06	NS
LAC (mmol/L)	4.25±0.33	3.68±0.46	3.48±0.52	0.003 ^{a,b}	S
MDA (μmol/L)	21.5±2.6	18.3±2.4	15.9±2.18	< 0.001 ^{a,b}	S
TAC (mg/dl)	2.24±0.34	3.17±0.61	3.41±0.27	< 0.001 ^{a,b}	S

N= number of rats in each group NS = non-significant S = significant

a: significance between G2, and G1 b: significance between G3, and G1

Table (8): Illustrates that there were statistically significant differences (p-value <0.05) in different groups (after exhaustive exercise bout after application of training program) for measured markers (p-value <0.05) between group 1 and each of groups 2, and 3 as regards to LAC, MDA and TAC level.

On the other hand there were no statistically significance differences (p-value >0.05) as regards SB100 level.

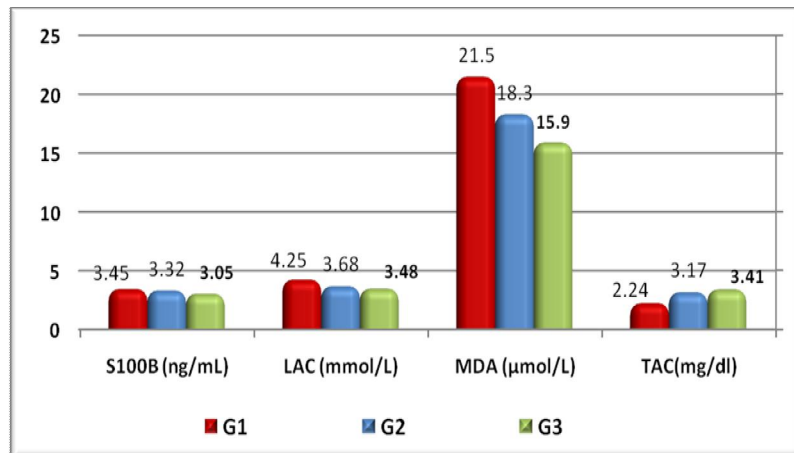


Figure (7): Comparisons between exhaustive after application of training program markers in different groups.

Table (9): Comparison of differences in the measured markers in resting and exhaustive conditions after the application of the training program in group 2. (n=9)

Variables	Resting	Exhaustive exercise	% change	p-value	Sig.
	Mean±SD	Mean±SD			
S100B (ng/mL)	2.76±0.37	3.32±0.47	20.3	0.003	S
LAC(mmol/L)	1.34±0.31	3.68±0.46	174.62	<0.001	S
MDA(μmol/L)	11.1±1.4	18.3±2.4	64.86	<0.001	S
TAC(mg/dl)	1.36±0.24	3.17±0.61	133.1	<0.001	S

N= number of rats in each group S = significant

Table 9: Illustrates that there was statistically significant increase (p-value <0.05) in serum level of different markers (S100B, LAC, MDA, and TAC level) after the exhaustive exercise bout when compared to resting condition in group 2 after the application of training program.

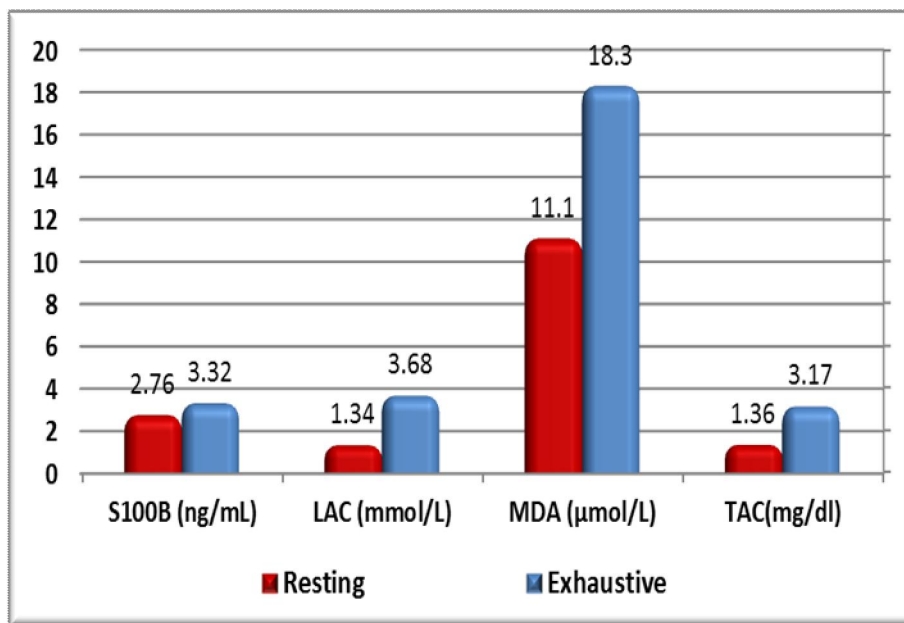


Figure (8): Comparison of different markers under resting and exhaustive conditions in group 2 after the application of training program.

Table (10): Comparison of differences in the measured markers in resting and exhaustive conditions after the application of the training program in group 3. (n=9)

Variables	Resting	Exhaustive exercise	% change	p-value	Sig.
	Mean ±SD	Mean ±SD			
S100B (ng/mL)	2.59±0.36	3.05±0.34	17.76	0.001	S
LAC(mmol/L)	1.12±0.19	3.48±0.52	210.71	<0.001	S
MDA(μmol/L)	11.2±2.04	15.9±2.18	41.96	0.001	S
TAC(mg/dl)	1.36±0.27	3.41±0.27	150.7	<0.001	S

N= number of rats in each group S = significant

Table 10: Illustrates that there was statistically significant increase (p-value <0.05) in serum level of different markers (S100B, LAC, MDA, and TAC level) after the exhaustive exercise bout when compared to resting condition in group 3 after the application of training program.

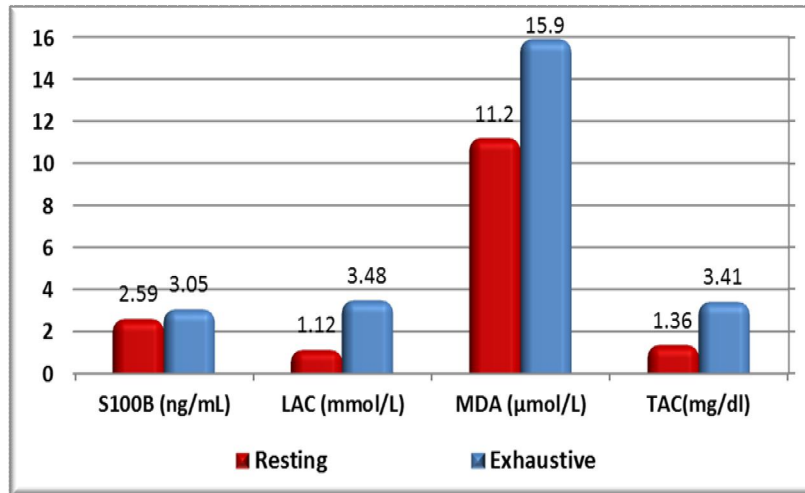


Figure (9): Comparison of different markers under resting and exhaustive conditions in group 3 after the application of training program.

Table (11): Follow up of the changes in different markers before and after application of the training program in group 2. (n=9)

Variables	Resting pre training program	Exh. exc. pre training program	Resting post training program	Exh. exc. post training program	p-value	Sig.
	Mean± SD	Mean± SD	Mean± SD	Mean± SD		
S100B (ng/mL)	3.11±0.70	3.56±0.54	2.76±0.37	3.32±0.47	<0.001	S
LAC (mmol/L)	1.14±0.12	4.34±0.88	1.34±0.31	3.68±0.46	<0.001	S
MDA (μmol/L)	11.3±1.1	21.1±3.5	11.1±1.4	18.3±2.4	<0.001	S
TAC (mg/dl)	1.08±0.11	2.38±0.62	1.36±0.24	3.17±0.61	<0.001	S

N= number of rats in each group S = significant

Table 11: Illustrates that there were statistically significant differences (p-value <0.05) in different markers (S100B, LAC, MDA, and TAC level) follow up at resting and after exhaustion before training program with increase levels after exhaustion. Then decrease in the level at resting after training program then increase again exhaustion after training program with decrease in the level of (S100B, LAC and MDA) and increase in the level of TAC in group 2.

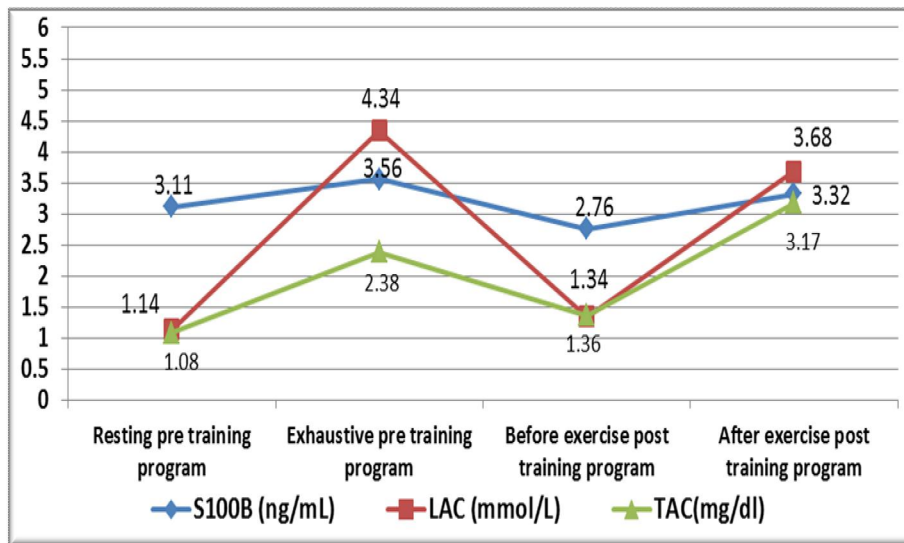


Figure (10): Different markers before and after application of the training program follow up in group 2.

Table (12): Follow up of the changes in different markers before and after application of the training program in group 3. (n=9)

Variables	Resting pre training program	Exhaustive pre training program	Resting post training program	Exhaustive post training program	p-value	Sig.
	Mean± SD	Mean± SD	Mean± SD	Mean± SD		
S100B (ng/mL)	3.12±0.52	3.89±0.50	2.59±0.36	3.05±0.34	<0.001	S
LAC (mmol/L)	1.09±0.22	4.53±0.44	1.12±0.19	3.48±0.52	<0.001	S
MDA (µmol/L)	12.1±1.3	22.7±3.2	11.2±2.04	15.9±2.18	<0.001	S
TAC (mg/dl)	1.09±0.17	2.35±0.64	1.36±0.27	3.41±0.27	<0.001	S

N= number of rats in each group S = significant

Table 12: illustrates that there was statistically significance decrease with p-value <0.05 in different markers (S100B, LAC, MDA) between resting and exhaustive pre training program and resting and exhaustion post training program with use of vit c. there was also statistically significance increase in level of TAC level between resting and exhaustive pre training program and resting and exhaustion post training program with use of vit.c.

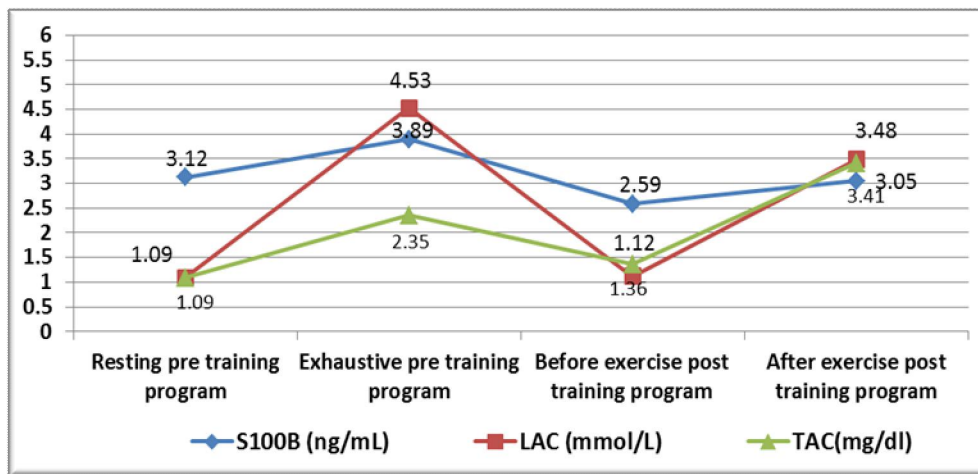


Figure (11): Different markers follow up among group 3 (taking vitamin C).

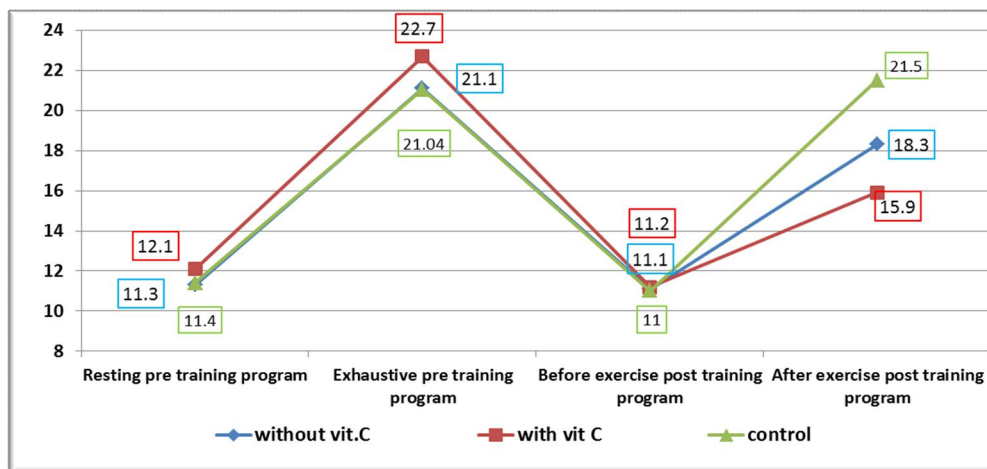


Figure (12): MDA marker follows up in different study groups.

Table (13): Comparisons of different change between resting and exhaustive pre and post training program markers in group 2. (n=9)

Variables	Difference in pre training program	Difference in post training program	p-value	Sig.
	Mean \pm SD	Mean \pm SD		
S100B (ng/mL)	0.36 \pm 0.11	0.25 \pm 0.1	0.04	S
LAC (mmol/L)	2.84 \pm 0.82	1.88 \pm 0.75	0.02	S
MDA (μ mol/L)	0.89 \pm 0.26	0.66 \pm 0.20	0.05	S
TAC (mg/dl)	1.1 \pm 0.40	1.52 \pm 0.45	0.05	S

N= number of rats in each group S = significant

Table 13: Illustrates there were statistically significance differences (p-value <0.05) in markers (LAC, S100B, Lactic acid and MDA level) as regards to percent change exhaustive and post training program with low mean difference post training.

Table (14): Comparisons of different change different markers resting and exhaustive before and after application of training program in group 3. (n=9)

Variables	Difference in exhaustion	Difference in exercise	p-value	Sig.
	Mean \pm SD	Mean \pm SD		
S100B (ng/mL)	0.40 \pm 0.32	0.17 \pm 0.08	0.05	S
LAC (mmol/L)	3.31 \pm 1.1	2.2 \pm 0.81	0.03	S
MDA (μ mol/L)	0.86 \pm 0.19	0.54 \pm 0.31	0.02	S
TAC (mg/dl)	1 \pm 0.59	1.60 \pm 0.52	0.04	S

N= number of rats in each group S = significant

Table 14: Illustrates there were statistically significance differences with p-value <0.05 in (MDA, S100B, TAC and Lactic acid) as regards to percent change between resting and exhaustive before and after application of training program.

Conclusion and Recommendation

The present work showed that endurance exercise promoted an increase in serum S100B levels independent of CNS injury. Several mechanisms related to central and/or peripheral S100B sources may be involved. However, more studies are needed to clarify the influence of serum S100B protein in exercise physiology, as well as the mechanism involved in its secretion during physical activity.

Vitamin C is an essential component of the diet and may reduce the adverse effects of exercise-induced reactive oxygen species, including muscle damage, immune dysfunction, and fatigue. However, reactive oxygen species may mediate beneficial training adaptation that vit.C attenuates and provide other health benefits without impairing training adaptation. In addition, the positive health benefits of using vitamins E and C may suggest an additive or synergistic effect when combined with regular exercise. New research initiatives should examine the following:

- The effects of vitamin E and C together on the adaptive response to strength training.
- The combined effects of exercise and vitamin C on diabetes risk factors.

- The combined effects of exercise and vitamin C on lipoprotein status and risk factors for cardiovascular disease.

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