Phytochemical screening, antibacterial and antiplasmodial activities of chrozophora oblique and launaea nudicaulis

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Abstract: The present study showed the different Phytochemicals screening in chrozophora obliqua and luanaea nudicaulis plants and also correlates the antibacterial and antiplasmodial activity. The results of the study show that the crude and all fractions having zeroed or negative affects to phytochemicals such as alkaloids and saponnins tests, whereas the Flavonoids were present in crude extract and all fractions. While the Tannins test showed the positive results against the crude extracts, chloroform and ethyle acetate fractions, whereas the Crude extracts were completely active against all the bacterial strains. Further from the results of the study found that the Hexane and crude extract having better results against Staphylococcus aureus, Escherichia coli and salmonella typhi. Furthermore, from the results of the study noted that the Crude hexane, Chloroform, E. acetate, butanol and H2O having negative result against Escherichia coli. Out of all fractions the methanolic fractions were most remarkable fractions obtained against malarial activity, while the n-hexane showed better result in 0.025 µg/ml drug dose level. Chloroform, E-acetate and n-butanol were found to be negative effects against all drug dose level, while the Chloroquine was taken as positive control for drug dose amount for known chemo factor which do not showed the increase in number of schizonts.

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1. Introduction

The exploration of novel Phytochemicals needs a lot of research. Nature has bestowed the wild flora with valuable chemistry [1]. Medicinal herbs are globally used for various purposes such as food, health care, fuel and medicines. Medicinal plants and their constituents are the potential sources of both traditional as well as allopathic drugs [2]. The Medicinal power of these plants lies in Phytochemical constituents which cause definite Pharmacological actions on the human body [3]. Majority of Phytochemicals have been known to bear valuable therapeutic activities such as insecticides [4], antibacterial, antifungal [5], anticonstipative [6], spasmolytic [7], antiplasmodic activities [8]. The genus Chrozophora belongs to family euphorbiaceae with 30 species. The Chrozophora obiqua is used in food and traditional medicines for the treatment of infectious diseases, whooping cough, appetite and as a stimulant [9].

Launaea is an important genus of family asteraceae with 59 species. The plant is popularly used in folk medicine for the treatment of fever, swelling, cuts, itches, ulcer, toothache and eczema eruption [10]. The World Health Organization (WHO) currently encourages, recommends and promotes traditional remedies in health care programs as they are easily available at low cost, comparatively safe and are culturally acceptable [11]. Limited information is available regarding antibacterial and antiplasmodail activity of chrozophora obliqua and launaea nudicaulis. Therefore, present study was carried out to investigate the antibacterial and antiplasmodail activity against various bacterial and plasmodial species. Preliminary phytochemical studies of these extracts are also undertaken to find out bioactive compounds having antibacterial and antiplasmodial activity.

2. Materials And Methods

2.1 Plant Material

The amount of desired plants samples of Chrozophora obliqua and Launaea nudicaulis were collected from the areas of Pakistan. These plants were recognized by the faculty of Botany Department, Kohat University of Science and Technolgy, Pakistan. **2.2 Preparation of extracts**

The plants samples were appropriately rinsed with distilled water to removed soil, sand and other surface like bacteria and virus etc. and then were placed in shadow for drying at temperature of 25 - 30 0C. The dried plants were grinded individually and then stored in dirt free, dried synthetic bags for additional biological and chemical processes.

The weight of dried plants (chrozophora obliqua and launaea nudicaulis) up to 2 kg was saturated in methanol for 15 days individually and were extracted and then filtered. The methanol filtrate was passed through (Vacuum Rotary Evaporator) VRE at 50 C0 to obtained dehydrated extracts of methanol. The extracted methanol were suspended in water and consecutively partitionized with n-hexane. chloroform, ethyl acetate and n-butanol to get nhexane-soluble, chloroform-soluble, ethyl-acetatesoluble, n-butanol soluble and dihydrogen oxide fractions, correspondingly. The isolated material and successive further solvent fractions were after that subjected to phytochemical study, antibacterial and antiplasmodial biological test.

2.3 Phytochemical screening

The extracts were subjected to preliminary phytochemical screening for possible presence of bioactive antimicrobial compounds by the methods of harborne [12] and sofowora [13] for identifying alkoloids, saponins, flavons, terpens and tannins.

A. Alkaloids: Dilute 70 % Hydrochloric acid solutions were mix up with extracts, fractions and filtered. The solution of Hager's reagent was inserted to the filtrate. Yellow color precipitate shows alkaloids existence [14].

B. Saponins: Extracts, fractions were individually heated with 10 mL of dihydrogen oxide (distilled) for about 15 minutes and mix materials was then cleaned and permitted to cool. Then 3 mL of filtrates were collected and mix with 15 mL sterilized water and vibrate dynamically for 4 minutes. Froth formation showed the occurrence of saponins in the filtrate [15].

C. Tannins: Extracted material and fraction was heated individually with 20 mL of H2O (sterilized) for 10 minutes in a water bath and then cleaned. Then 2 mL of the filtrates were combined with 10 mL dihydrogen oxide and 4-5 drops of 20 percent Ferric tri-chloride solution were inserted. A blue colour or brown-green precipitate shows tannins occurrence [16].

D. Flavonoids: Extracted material and fraction was mix up to 15 mL of ethyl acetate. Solution was heated for 5 minutes and then cleaned. Mix 1 mL of dilute NH3 solution to 5 mL filtrate. Yellowish color precipitate shows flavonoids occurrence [17].

E. Terpenes: Mix solution 0.5 mg of extract and fraction individually to 6 mL dihydrogen oxide and then mixed with 5-6 ethyl acetate drops. Occurance of bright green color shows terpenes [17].

3. Antibacterial activity

Extracts and various fractions of *chrozophora obliqua* and *launaea nudicaulis* were studied for antibacterial assay using gram negative, gram positive

bacteria. Three bacterial strains E. coli, Staphylococcus aureus and Salmonella typhi were used by Agar Well Diffusion Susceptibility Method [18]. In Dimethyl Sulfoxide (DMSO) solutions the extracts and fractions were prepared at the same amount of 3 μ g/ μ L and DMSO was used as a solvent, because it cannot demonstrate any type of activity against bacteria. During the investigation, 28 g/L nutrient agar was used and in the solution media, petri plates and borer were cleaned for 20 minutess at very elevated pressure and at the temperature of 130 °C in autoclave. Agar media were poured in Petri dishes for laminar flow. The 6 mm of sterile discs were dig in bacterial media by means of plastic borer. Every well were given a named special. The cultured bacterial consequent to 10^4 to 10^6 (colony forming unit) were speckled on the solid media surface. The Solutions of methanolic extracts and different fractions of same amount 2 $\mu g/\mu L$ were prepared. The stock solution of each 40 mL was further added into the applicable wells. The zones of inhibition were calculated once a day for incubation at 40 °C in the incubator. Amoxicillin standard of 5 µg/µL were used for positive control and DMSO was used as a negative control. The inhibition zones of methanolic extracts and other soluble fractions were matched with given drugs as inhibition zones i.e. amoxicillin. The quantity of bacterial growth in each well was calculated.

3. Antiplasmodail activity of Chrozophora oblique 3.1Antiplasmodial bioassay

The modified method of kerharo and adam (1974) was followed. 100 µL of blood sample were used for the patient of malaria in 8 wells of micro titer plates and were subjected equally amount for the media culture, Roswell park memorial institude-1640. 200 (IU) pencillin and 15 µL kanamycin were subjected into micro titer plate well for the protection of bacterial contamination. Next the micro titer plates were shaked continousely to obtained same phase and standrize suspension in every well. 20 µL drugs for each dose level i.e. 0.025 µg/mL and 0.050 µg/mL were transferred into apprehensive well and chloroquine were used for positive control. The drug amount of famous chemo factor was placed in well number [7]. The negative control in well 8 was kept at the end of the test. The micro titer plates were placed in incubator at 40 °C for the next five days. The duration of incubation, every day different research slides of each wells were made for micro titer plate. Every day all slides are noticeable for detection. Each research Slides were dehydrated and set with CH3OH. When the slides were dried, every slide was marked with the giemsa solution (methylene blue) and examined with the help of microscope at 10 x, 40 x and 100 x specification. In this way the growth of schizont were detected and recorded.



Figure 1. Photographic view of antibacterial activity of Chrozophora obliqua and Launaea nudicaulis

4. Results And Discussion

4.1 Phytochemical screening of chrozophora oblique and launaea nudicaulis

The following results showed the detection of various group of phytochemicals in different methanolic extracts and various organic fractions of chrozophora obligua and launaea nudicaulis as shown in tables 1. It shows the phytochemical examination of methanolic crude extract and various chemical fractions of chrozophora obligua plant. The methanolic crude extract and all chemical made fractions give zero or negative results to phytochemical like alkaloids and saponins tests. These two phytochemicals were absent in the given tested samples. Flavonoids were present in methanolic crude extract and in all chemical fractions. Terpenes were present in all fractions except n-butanol and aqueous fractions. Tannins were present in methanolic crude extract, chloroform and E. acetate fraction. The table shows the phytochemical examination of methanolic crude extract and the various chemical fractions of

Launaea nudicaulis plant. The methanolic crude extract and all the chemical fractions give zero or negative results to alkaloids and saponins tests and hence were observed absent in the tested samples.

4.2 Antibacterial Activity

The extracts and fractions of *Chrozophora* oblique and *Launaea nudicaulis* plants were same concentrations ration of 2 μ g/ μ L for getting duplicate results carried various inhibition zones. *In vitro* antibacterial activities of crude extract and chemical fractions of *chrozophora obliqua* and *launaea nudicaulis* were subjected, these activities were matched with the given antibiotics (Amoxicillin). In the following antibacterial activity three bacterial strains E. coli, S. aureus and S. typhi was analyzed by AWDSM "Agar Well Diffusion Susceptibility Method". The inhibition zone for antibacterial activity of Chrozophora obliqua is displayed in table 2.

4.3 Antibacterial activity of Chrozophora oblique and launaea nudicaulis

| Plants | Extracts | Alkaloids | Flavonoids | Terpenes | Tannins | Saponins |
|---------------------|------------|-----------|------------|----------|---------|----------|
| | Crude | - | + | + | + | - |
| | n-Hexane | - | + | + | + | - |
| Chrozophore obligue | Chloroform | - | + | + | + | - |
| Chrozophora obliqua | E-acetate | - | + | + | - | - |
| | n-Butanol | - | + | - | - | - |
| | Aqueous | - | - | - | - | - |
| | Crude | - | + | + | + | - |
| | n-Hexane | - | + | + | + | - |
| Launaaa nudiaulia | Chloroform | - | + | + | + | - |
| Launaea nuulcuns | E-acetate | - | + | + | - | - |
| | n-Butanol | - | + | + | - | - |
| | Aqueous | - | + | + | - | - |

Table 1. Phytochemical screening of chrozophora obliqua and launaea nudicaulis

| | Inhibition zone (mm) | | | | | | | |
|-----------------------|----------------------|--------------------|------------|------------|-----------|---------|-----------|--|
| Organisms | Chrozophora oblique | | | | | | | |
| _ | Crude | n-hexane | Chloroform | E. Acetate | n-butanol | Aqueous | Standards | |
| Escherichia Coli | 9 | 7 | 7 | 6 | - | - | 19 | |
| Salmonella typhi | 14 | 16 | 17 | 10 | 9 | 9 | 21 | |
| Staphylococcus Aureus | 11 | 16 | 17 | 15 | 12 | 7 | 21 | |
| | Launa | Launaea nudicaulis | | | | | | |
| Escherichia Coli | 6 | 6 | - | - | - | - | 19 | |
| Salmonella typhi | 12 | 18 | 15 | 12 | 13 | 10 | 21 | |
| Staphylococcus Aureus | 15 | 17 | 12 | 11 | 10 | 9 | 21 | |

Table 2. Antibacterial activity of Chrozophora obliques and Launaea nudicaulis

| S. No | Drug | Drug dose level | Drug Concentration (µg⁄ml) | Isolation of parasites in different fractions (Schizonts) | | | | | |
|----------|------|-----------------------|-------------------------------|---|--------------|------------|---------------|---------------|---------|
| | | | | Methanol | n- Hexane | Chloroform | E. Acetate | n- Butanol | Aqueous |
| 1 | ٨ | A ₁ | 0.025 | 01 | 02 | 03 | 03 | 04 | 05 |
| 1 | А | A ₂ | 0.050 | 0 | 01 | 02 | 03 | 03 | 04 |
| 2 | п | B_1 | 0.025 | 0 | 03 | 03 | 04 | 04 | 05 |
| 2 1 | в | B ₂ | 0.050 | 0 | 03 | 03 | 03 | 04 | 05 |
| 2 | C | C ₁ | 0.025 | 01 | 03 | 02 | 04 | 03 | 04 |
| 3 | C | C ₂ | 0.050 | 0 | 02 | 01 | 03 | 02 | 04 |
| 4 | D | D | Chloroquine | No Growth | | | | | |
| 5 | Е | Е | Distilled water media | Hundred percent growth | | | | | |

Table 3. Antiplasmodail activity of chrozophora obliqua

Table 4. Antiplasmodail activity of launaea nudicaulis

| C C | Drug | Drug dose level | Drug Concentration (µg⁄ml) | Isolation of parasites in different fractions (Schizont) | | | | | | |
|------------|------|-----------------------|----------------------------------|--|--------------|-------------|-----------|---------------|---------|--|
| s. No | | | | Methanol | n- Hexane | Chloroform | E.Acetate | n- Butanol | Aqueous | |
| 1 | • | A ₁ | 0.025 | 01 | 02 | 03 | 03 | 04 | 05 | |
| 1. | А | A_2 | 0.050 | 0 | 01 | 02 | 03 | 03 | 04 | |
| 2 | р | B ₁ | 0.025 | 0 | 03 | 03 | 04 | 04 | 05 | |
| 2 В | D | B ₂ | 0.050 | 0 | 03 | 03 | 03 | 04 | 05 | |
| 3 C | C | C ₁ | 0.025 | 01 | 03 | 02 | 04 | 03 | 04 | |
| | C | C ₂ | 0.050 | 0 | 02 | 01 | 03 | 02 | 04 | |
| 4 | D | D | Chloroquine | No growth | | | | | | |
| 5 | Е | Е | Distilled water media | Hundred p | ercent grow | <i>r</i> th | | | | |

Methanolic Crude extract was completely active throught out all bacterial strains. Hexane and the crude extract showed good results against S.aureus, *E.coli* and *s typhi*. Chloroform, E. acetate, n- butanol and H_2O soluble fraction show negative result against E.coli.crude, n-hexane and chloroform showed maximum inhibition (19mm). n- hexane and chloroform fraction showed good activity against *salmonella typhi* and *staphylococcus aurens*. Methanolic Crude extract and n- hexane were throught out active against salmonella *typhi* and *staphyloccus aureus*. Hexane and crude extract show good activity against *S.aureus*, and *s typhi*. methanolic Crude hexane Chloroform, E. acetate, butanol and H_2O show negative result against *E.coli*.

4.3 Antiplasmodail activity

4.3.1 Antiplasmodail activity of Chrozophora oblique and launaea nudicaulis

The table 3 demonstrates the results of antiplasmodial activity. Out of all fractions, the *CH*₃*OH* fraction of *Chrozophora obliqua* is the most remarkable fraction obtained against malarial activity. n-hexane revealed good result in A2 drug dose level. Chloroform fraction exibited good result in C2 drug dose level.. E-acetate and n- butanol showed negative results against all drug dose level (≤ 1 level is

significant). Chloroquine were taken as positive control for drug dose amount for known chemo factor which do not showed the increase in number of schizonts. The table 4 shows the results of antiplasmodial activity of launaea nudicaulis. Out of all fractions, the CH_3OH fraction of launaea nudicaulis are most remarkable fractions obtained against malarial activity. n-hexane showed good result in A2 drug dose level. Chloroform, E-acetate and nbutanol showed negative result against all drug dose level. (≤ 1 level is significant). Chloroquine were taken as positive control for drug dose amount for known chemofactor which do not showed the increase in number of schizonts.

5. Conclusion

The results proved that the plants store different phytochemicals classes of with different concentration. These phytochemicals are responsible for different biological activity by inhibiting the reproduction of different microbes and destroy them. The antibacterial and antiplasmodial results of chrozophora oblique and launaea nudiculis indicate that crude extracts, ethyl acetate, chloroform and nbutanol fraction of all parts were the most active fraction. It is recommended that chrozophora oblique and launaea nudiculis are important plants from medicinal point of view and can be potent candidates for further in-vivo bioassays which would leads to the synthesis of safe herbal drugs with or less side effects of global interest.

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