

Effect of Organic manure and Mycorrhizal on the Growth and Yield of *Capsicum annum* (Hot Pepper).Tolulope Yetunde Akande¹, Olajire Fagbola², Kehinde Olajide Erinle³, Tope Daniel Bitire², Joseph Urhie¹¹ College of Resources and Environment, Northeast Agricultural University, Harbin 150030, P. R. China;² Agronomy Department, Faculty of Agriculture & Forestry, University of Ibadan, Ibadan Nigeria;³ School of Agriculture, Food and Wine, University of Adelaide, Adelaide SA 5005.

Corresponding Author: fagbola8@yahoo.co.uk

Abstract: A screen house experiment was conducted to investigate the growth and nutrient uptake as influenced by organic manure and arbuscular mycorrhizal fungi (AMF) in *Capsicum annum*. The treatments were inoculated with arbuscular mycorrhizal (M+), without arbuscular mycorrhizal (M-) and with application of organic manure; with pig dung (PD+), without pig dung (PD-), with poultry manure (PM+), without poultry manure (PM-). The experiment was laid out in factorial combination using complete randomized design with three replicates. The result of this study showed that at 5 weeks after planting plant height increased with the application of pig dung and poultry manure. Stem diameter increased across the weeks but at 5 weeks after planting mycorrhizal inoculation, with pig dung and poultry manure increased. The application of Pig dung increased the root and shoot biomass of plant when compared with the inoculated plants. The combined application of pig dung and mycorrhizal inoculation significantly increase Mg uptake when compared with control. The concentration of nutrients especially phosphorus must be considered in the soil before inoculation with AMF and application of organic manure maintained the soil fertility.

[Akande TY, Fagbola O, Erinle KO, Bitire TD, Urhie J. **Effect of Organic manure and Mycorrhizal on the growth and Yield of *Capsicum annum* (Hot Pepper)**. *N Y Sci J* 2018;11(5):1-9]. ISSN 1554-0200 (print); ISSN 2375-723X (online). <http://www.sciencepub.net/newyork>. 1. doi: [10.7537/marsnys110518.01](https://doi.org/10.7537/marsnys110518.01).

Keywords: Arbuscular mycorrhiza, Organic manure, Vegetative Growth, Nutrient Uptake.

Introduction.

Degradation of agricultural lands is escalating worldwide and most especially in the tropics. Mbagwu *et al.*, (2003) described it has temporary or permanent destruction of the soil productive capacity which is caused by man-induced activities. As a result of fertility decline, there is a low level of organic matter in soil associated with physical and biological properties and buildup of toxic concentrations in the soil. However, Swift and Palm (2000) suggested that it is more helpful to describe soil fertility as an ecosystem concept integrating the diverse soil functions, including nutrient supply which promotes crop production. These can be achieved by the use of appropriate management practices such as green manure, mulching, cover cropping and organic manure which have continuous effect on the soil in the long-run. Hence, there is need to understand the soil environment, recognize the limitation of that environment and ameliorate where possible without damaging the soil quality for efficient crop production.

Duruigbo *et al.*, (2013) described Organic agriculture as the use of natural resources and focus on environmental protection including the maintenance of soil structure and fertility, water resources and biodiversity). Application of organic residues to soils increase soil organic matter, buffer soil, improve aggregate stability and enhance water retention capacity of soils (Spaccini *et al.*, 2002). Organic

agriculture is environmental friendly and excludes use of inorganic fertilizer. Manure has been used effectively as organic fertilizer for centuries and their nutrient content varies depending on source and nutrient content (Egbuchua and Enujoke, 2013). Organic manures are efficient in increasing soil nutrient contents, ensuring positive residual effects and enhancing soil's physical and chemical characteristics (Ayeni *et al.*, 2010). The utilization of organic manures is an integral part of agriculture that has been exploited overtime and across ages because of its ability to restore soil fertility, supply major plant nutrients, such as N, P, K, Ca, Mg and also stabilizer soil pH (Sanchez and Miller, 1986).

Many agricultural and horticultural crop species form a mutualistic symbiotic relationship with AMF in the root of 95% of all vascular plant (Smith and Read, 1997). Plant host benefits from increase AMF abundance and or functionality due to soil amendments (Rillig and Mummey, 2006; Warnock *et al.*, 2010). Plants inoculated with AMF are more efficient in acquisition of nutrients which results into improved plant growth (Oseni *et al.*, 2010), improve P uptake (van der Heijden *et al.*, 2006; Smith *et al.*, 2015), and under certain conditions they can also contribute to nitrogen (N) uptake (Hodge *et al.*, 2010; Whiteside *et al.*, 2012) and micronutrients (Perner *et al.*, 2007). Fagbola *et al.*, (1998) also reported increased in P absorption due to more efficient mining

of soil P. According to Javaid and Riaz (2008), root colonization with AMF enhances plant tolerance to growth inhibition factors, thus improving crop growth and productivity. Hence, amended soil improves AMF functionality (Warnock *et al.*, 2010). Mycorrhizae are of immense importance in maintaining soil fertility and these are influenced by climatic and seasonal changes in the physico-chemical properties of soil.

Pepper (*Capsicum species*) is one of the most varied and widely used foods in the world. Hot pepper (*Capsicum annum*) is widely cultivated and seen has traditional vegetable or spice (Grubben and El-Tahir, 2004). The major constraints of pepper production are infertile soil, weeds, pests and diseases. Reinder (2007) reported that the use of organic manure in horticultural crops production enhances genetic expression of such crop in term of quality and quantity of the crop. Comparatively, yield in developing countries is about 10-30% of that in developed countries (Grubben and El-Tahir, 2004). Capsicum in combination with herbs is believed by people to cure certain ailments and diseases such as measles, fever and cold. (Grubben and El-Tahir, 2004). The objective of this study is to determine the response of *Capsicum annum* (Hot Pepper) to organic manure and mycorrhizal inoculation in topsoil.

2.0 Materials And Methods

Soil collection and analysis.

The experimental soil was collected on the agricultural land site of the University of Ibadan, Nigeria (7°24'N; 3°48'E) and pot experiment conducted in the screen house of the Department of Agronomy, University of Ibadan, and Ibadan, Nigeria. The collected soil sample was collected at a depth of 0-15cm. These soils were air dried and sieved with 5mm mesh size to remove debris before weighing into 5kg pots. The bulk soil sample was air dried at room temperature and grounded to pass through 2mm and 0.5mm diameter sieve before analysis. The particle distribution analysis was done by Hydrometer method (Gee and Bauder, 1986). The pH was determined in a 1:2 soil/water suspension using digital pH meter. Organic carbon was determined by Walkley-Black dichromate wet oxidation method (Nelson and Sommers, 1982). Total nitrogen was by micro-Kjeldahl distillation technique as described by Bremner and Mulvaney (1982). Available phosphorus was determined by Bray No 1 method as described by IITA (1982). Exchangeable K, Ca and Mg were extracted using Ammonium Acetate, K was determined using flame photometer, Ca and Mg determined using Atomic Absorption Spectrophotometer (AAS) (Table 1).

Table 1. Soil physicochemical properties

Soil depth (cm)	pH (H ₂ O)	Organic C (g/kg)	Total N	Available P (mg/kg)	Exchangeable Bases				Sand (g/kg)	Silt	Clay
					K (cmol/kg)	Na	Ca	Mg			
0-15	7.1	25.52	1.39	15.5	2	0.8	1.9	9.8	838	160	2

Experimental Design

The experiment was a factorial combination conducted in a completely randomised design with three replicates. The factors include: two levels of pig dung (with or without), two levels of poultry manure (with or without), and two levels of mycorrhizal inoculation (with or without).

Manure analysis and mycorrhizal treatments.

Pig and poultry manure were incorporated into the soil at the rate of 10t/ha. Air dried manure were characterized by digestion with nitric and sulfuric acid, analyzed for macronutrients by standard procedures (Kaira and Maynard 1991; Cater 1993) (Table 2).

Arbuscular mycorrhizal (*Glomus clarum*) inoculum (20g) was applied at third-quarter central top of the soil 24 hours before transplanting pepper seedlings (Carling *et al.*, 1978). Pepper seedlings (Roma) were obtained from Institute of Agricultural Research and Training Ibadan, Nigeria. Mycorrhizal colonization was quantified after clearing root sample in 10% KOH, acidified in 1% HCl for 30 min and then stained with 0.05% trypan blue in lactoglycerol (1: 1: 1 lactic acid, glycerol and water) at room temperature (Phillips and Hayman, 1970). The stained roots, inside a gridline plate (Giovanetti and Mosse, 1980) were observed under a dissecting microscope.

Table 2. Poultry and pig manure chemical properties

	pH (H ₂ O)	Organic C (%)	Total N (%)	Available P (%)	Exchangeable Bases			
					K (cmol/kg)	Na	Ca	Mg
Poultry manure	9.1	68.26	0.52	0.32	0.8	0.2	0.2	1.9
Pig dung	5.6	63.61	0.31	0.06	0.4	0.0	0.3	0.1

Data collection and harvest.

The growth parameters observed on the seedlings were: height (cm) and stem diameter (cm). The number of fruit, roots and shoot dry weight biomass, N, P, K, and Ca, Mg of leaf and soil, and percentage mycorrhizal colonization were determined. Leaf, shoot and root dry weights were taken after oven drying the samples at 70°C for 48 h. The plant samples were digested in triple acid as described by Juo *et al.*, (1974). The P in solution was determined by ascorbic acid (Murphy and Riley, 1962). Nitrogen in the leaf was determined by digesting dried and ground leaf samples in hot tetraoxosulphate (vi) acid solution with selenium catalyst (Novozamskey *et al.*, 1983). The exchangeable cations (K, Ca and Mg) were determined using Atomic Absorption Spectrophotometer.

Statistical analyses.

All data were analyzed using analysis of variance (ANOVA) with the GENSTAT and Duncan's Multiple Range Test (DMRT) at $P < 0.05$ was used to separate the means.

3.0. Results.

Effect of organic manure and mycorrhizal inoculation on vegetative growth of pepper plant

Plant height under pig dung and poultry manure application was significantly high when compared with poultry manure thus contributing 25.7% increase to plant height at 5 WAP (Table 3). Similar treatment trend was observed at 7 WAP, with pig dung and poultry manure contributing 21.6% increase to plant

height in soil. Pig dung application contributed 6.9% increase to plant height when compared with mycorrhizal inoculation and pig dung while with mycorrhizal inoculation and poultry manure application contributed 11.6% increase when compared with poultry manure application under screen house condition. There was no significant difference in plant height at 9 WAP. Although, pig dung and poultry manure application contributed 10.7% increase to plant height when compared with mycorrhizal inoculation, pig dung and poultry manure application. At 11 WAP, there was no significant difference although with pig dung and without mycorrhizal inoculation contributed 10.9% increase than with mycorrhizal inoculation and pig dung; with pig dung and poultry manure contributed 8% increase compared with mycorrhizal inoculation, pig dung and poultry manure in the soil. Stem girth of pepper plant at 5WAP indicated with pig dung and poultry manure was significantly high when compared with mycorrhizal inoculation and poultry manure; with pig dung and without mycorrhizal inoculation; with poultry manure and without mycorrhizal inoculation; control which were significantly low. There was no significant difference at 7WAP, although pig dung contributed (4.10%) an increase to stem girth when compared with mycorrhizal inoculation and pig dung whereas with pig dung contributed 25.0% increase to stem girth than with poultry manure. There was no significant difference at 9 and 11 WAP (Table.3).

Table 3. Plant height and Stem girth of pepper plant as affected by mycorrhizal inoculation, pig dung and poultry manure application

Mycorrhizal inoculation	Pig dung application	Poultry manure application	Plant Height				Stem Diameter			
			Weeks after Planting				Weeks after Planting			
			5	7	9	11	5	7	9	11
M+	PD+	PM+	9.03ab	11.43ab	14.60a	16.10a	0.15a	0.22a	0.34a	0.45a
M+	PD+	PM-	8.50ab	11.57ab	14.43a	16.26a	0.13ab	0.23a	0.42a	0.64a
M+	PD-	PM+	9.00ab	12.00ab	14.57a	16.28a	0.11b	0.16a	0.29a	0.51a
M+	PD-	PM-	9.60ab	12.07ab	14.20a	15.70a	0.12ab	0.22a	0.39a	0.57a
M-	PD+	PM+	10.36a	13.53a	16.36a	17.50a	0.12ab	0.17a	0.34a	0.52a
M-	PD+	PM-	9.13ab	12.43ab	15.86a	18.26a	0.11b	0.24a	0.38a	0.64a
M-	PD-	PM+	7.70b	10.60b	14.16a	15.56a	0.10b	0.18a	0.37a	0.56a
M-	PD-	PM-	9.83ab	13.03ab	15.53a	17.33a	0.10b	0.20a	0.41a	0.51a
ANOVA										
M			NS	NS	NS	NS	*	NS	NS	NS
PD			NS	NS	NS	NS	*	NS	NS	NS
PM			NS	NS	NS	NS	NS	NS	NS	NS
M*PD			NS	NS	NS	NS	NS	NS	NS	NS
M*PM			NS	NS	NS	NS	NS	NS	NS	NS
PM*PD			*	NS	NS	NS	NS	NS	NS	NS
M*PM*PD			NS	NS	NS	NS	NS	NS	NS	NS

Means in columns followed by the same letter (s) are not significantly different according to Duncan's Multiple Range Test at ($P < 0.05$), (PD+)=With Pig dung; (PD-)= Without Pig dung; (PM+)=With Poultry Manure; (PM-)=Without poultry manure; (M+)=With mycorrhizal; (M-)=Without mycorrhizal; * $P < 0.05$. NS=Not significant. M=mycorrhizal; PD=pig dung; PM=poultry manure

Organic manure and mycorrhizal inoculation on dry weight biomass (root and shoot) number of fruit yield and mycorrhizal colonization.

The root dry weight under soil amendments showed no significant difference although with pig dung contributed 17.1 % increase when compared with mycorrhizal inoculation and pig dung, 22.8% and 15.6% increase when compared with poultry manure; control respectively. The shoot dry weight showed no significant difference although; pig dung contributed 16.9% increase when compared with mycorrhizal inoculation and pig dung, 39.2% increase when compared with control (Table 4). Pig and poultry manure application contributed 10.5% increase compared with mycorrhizal inoculation, pig dung and poultry manure. There was no significant difference in number of fruit yield under screen house condition although control performed better with 57.8% increase when compared with mycorrhizal inoculation, pig

dung and poultry manure application and 21% increase when compared with pig dung and poultry manure (Table 4). Pig dung and mycorrhizal inoculation contributed 26.6% increase when compared with pig dung whereas poultry manure contributed 71.5% increase when compared with mycorrhizal inoculation and poultry manure. Mycorrhizal colonisation was significantly high at with mycorrhizal inoculation, pig dung and poultry manure when compared with mycorrhizal inoculation and poultry manure application; with mycorrhizal inoculation; with pig dung and poultry manure; with pig dung; with poultry manure and control under screen house condition. However, with mycorrhizal inoculation, pig dung and poultry manure; with mycorrhizal inoculation and pig dung contributed 49.8% and 45.8% increase compared with control respectively.

Table 4. Effect of mycorrhizal inoculation, pig dung and poultry manure application on dry weight biomass of root and shoot, number of fruit yield and mycorrhizal percentage colonisation.

Mycorrhizal Inoculation	Pig application	Dung Poultry application	manure	Dry Root Biomass	Dry Shoot Biomass	Number of Fruit Yield	Mycorrhizal Colonization (%)
				------(g/plant)-----			
M+	PD+	PM+		0.96a	1.70a	2.67a	58.73a
M+	PD+	PM-		2.43a	3.37a	5.00a	54.43ab
M+	PD-	PM+		2.56a	1.80a	1.33a	32.57de
M+	PD-	PM-		2.10a	2.46a	1.67a	40.33c
M-	PD+	PM+		1.36a	1.90a	5.00a	41.67c
M-	PD+	PM-		2.93a	4.06a	3.67a	36.5cd
M-	PD-	PM+		2.26a	2.63a	4.67a	50.9b
M-	PD-	PM-		2.47a	2.47a	6.33a	29.47e
ANOVA							
M				NS	NS	NS	**
PD				NS	NS	NS	NS
PM				NS	NS	NS	NS
M*PD				NS	NS	NS	*
M*PM				NS	NS	NS	NS
PM*PD				NS	NS	NS	NS
M*PM*PD				NS	NS	NS	NS

Means in columns followed by the same letter (s) are not significantly different according to Duncan's Multiple Range Test at (P<0.05), (PD+)=With Pig dung; (PD-)= Without Pig dung; (PM+)=With Poultry Manure; (PM-)=Without poultry manure; (M+)=With mycorrhizal; (M-)=Without mycorrhizal; *P<0.05, **P<0.01, NS=Not significant. M=mycorrhizal; PD=pig dung; PM=poultry manure

Nutrient uptake as influenced by organic manure and mycorrhizal inoculation.

N uptake showed that with pig dung and poultry manure was significantly high when compared with pig dung; with poultry manure; with mycorrhizal inoculation and pig dung application were significantly low. Exception was with mycorrhizal

inoculation, pig dung and poultry manure; control; with mycorrhizal inoculation and without pig dung and poultry manure; with mycorrhizal inoculation and poultry manure which showed no significant difference (Table 5). There was no significant difference in P uptake of pepper plant although with mycorrhizal inoculation and pig dung contributed 68.7%

increase when compared with pig dung and 64.1% when compared with control. Similar trend was observed in K uptake, although poultry manure contributed 27.7% when compared with mycorrhizal inoculation and poultry manure in the soil and 64.1% increase when compared with control. With mycorrhizal inoculation, pig dung and poultry manure contributed 38.5% increase compared with pig dung and poultry manure increase in K uptake. Ca uptake showed no significant difference although with

mycorrhizal inoculation and pig dung showed 18.8% increase when compared with control. With mycorrhizal inoculation, pig dung and poultry manure contributed 57.1% and 38.8% increase compared with pig dung and poultry manure application and control. Mg uptake indicated with mycorrhizal inoculation, pig dung and poultry manure application was significantly high when compared with control which was significantly low (Table 5).

Table 5. Nutrient uptake in soil as affected by mycorrhizal inoculation, pig dung and poultry manure application

Mycorrhizal inoculation	Pig Dung application	Poultry Manure application	N	P	K	Ca	Mg
			(g/kg)	(mg/kg)	------(mg/kg)-----		
M+	PD+	PM+	1.08ab	0.43a	1.66a	0.98a	0.31ab
M+	PD+	PM-	0.89b	1.31a	1.38a	1.17a	0.38a
M+	PD-	PM+	1.47ab	0.47a	1.25a	0.70a	0.22ab
M+	PD-	PM-	1.24ab	0.66a	1.35a	0.97a	0.28ab
M-	PD+	PM+	1.65a	0.36a	1.02a	0.42a	0.17ab
M-	PD+	PM-	0.95b	0.41a	1.34a	0.95a	0.25ab
M-	PD-	PM+	1.06b	0.74a	1.73a	0.82a	0.23ab
M-	PD-	PM-	1.22ab	0.47a	1.01a	0.60a	0.13b
ANOVA							
M			NS	NS	NS	NS	NS
PD			NS	NS	NS	NS	NS
PM			NS	NS	NS	NS	NS
M*PD			*	NS	NS	NS	NS
M*PM			NS	NS	NS	NS	NS
PM*PD			NS	NS	NS	NS	NS
M*PM*PD			NS	NS	NS	NS	NS

Means in columns followed by the same letter (s) are not significantly different according to Duncan's Multiple Range Test at ($P < 0.05$), (PD+)=With Pig dung; (PD-)= Without Pig dung; (PM+)=With Poultry Manure; (PM-)=Without poultry manure; (M+)=With mycorrhizal; (M-)=Without mycorrhizal; * $P < 0.05$, NS=Not significant. M=mycorrhizal; PD=pig dung; PM=poultry manure

4.0 Discussion

Effect of organic manure and mycorrhizal inoculation on vegetative plant growth.

The result obtained from our study showed that pig dung and poultry manure application increase plant height when compared with poultry manure application especially at 5 and 7 week after transplanting. This supports the observations of Adesina *et al.*, (2014) who reported increase in plant height of pepper with manure application. Awosika *et al.*, (2015) also reported that the application of pig manure only at low rates increased agronomic properties of tomato plant (*Lycopersicon esculentum*). Inoculation with mycorrhizal and poultry manure application also showed increase when compared with poultry manure application. The growth increase is suggested to be attributed to mycorrhizal interactions

which enhanced plant to acquire nutrients. According to Crews *et al.* (1978) and Lovato *et al.*, (1995) inoculation with AMF increases the survival and growth rates of plants and cuttings in screen houses and natural conditions. Olawuyi *et al.*, (2014) reported that mycorrhizal inoculation with cow dung increased the agronomic characteristics of pepper plant. AMF aids in plant growth by increasing metabolic activities and accelerating transfer of nutrients from fungus to plants. Since, most crops species perform better with mycorrhizal inoculation, we suggest it enhanced the growth of pepper plants. Ewulo *et al.*, (2008) and Ikeh *et al.*, (2012) also reported increase in plant height of tomato and pepper respectively when soil was amended with poultry manure. Manure serves as source of organic matter which is easily mineralized and readily available for plant utilization. There was

general increase in stem girth of pepper plant with organic manure application and AM in the soil. Our study indicated that soil treatments with pig dung and poultry manure application significantly increase stem diameter of pepper plant at 5 weeks after transplanting when compared with mycorrhizal inoculation and poultry manure application and control. This supports a report by Senjobi *et al.* (2010) who reported that addition of farmyard manure improved all the growth parameters of leafy vegetable. Similarly, Ewulo *et al.* (2007) reported that cow dung applied at 7.5t/ha increase stem diameter of pepper plant. Our study showed no significant effect between mycorrhizal inoculated plants and non-mycorrhizal inoculated plants for root dry weight biomass. However, application of pig dung showed increase when compared with mycorrhizal inoculation and pig dung for dry root biomass of pepper plant. Erinle *et al.* (2017) reported that application of manure significantly increase dry weight of root and shoot. This is in accordance with Fernando and Linderman (1997), who reported that root dry weight of mycorrhizal inoculated plants are not significantly different from each other. Grey and William (2006) also reported an increase in the supply of nutrients following organic manure application. The shoot dry biomass increased with application of manure. This may likely be as a result of initiating different conformational changes in growth of different plants varieties. There was no significant difference in number of fruit yield. In our findings, inoculated plants did not show a better yield performance. This must have been due to the concept of nutrient imbalance in tropical soils (Akinrinde, 2006). In addition, enhanced mycorrhizal infection may account for enhanced plant growth through improved nutrient uptake.

Effect of AM fungi, organic manure on nutrient uptake and mycorrhizal colonisation.

AM fungi have been considered to have wide range of host in which species are more effective with particular host plant in increasing nutrient uptake. According to Smith and Read (1997), AM fungi are able to colonize most crop species by improving plant growth and nutrient uptake. The contribution of AMF to plant nutrient uptake is often particularly evident in plant that are deficient in certain nutrients mainly phosphorus. In our study, inoculation with mycorrhizal and pig dung application aided uptake of P. This agrees with George *et al.* (2000) that AM symbiosis increases P uptake and growth of their plant host and mycorrhizal inoculated roots have higher phosphorus absorption capacity compared to non-mycorrhizal root (McArthur and Knowles 1993; Ortas *et al.*, 1996). Subbarao *et al.* (2001) reported that organic manure sustained food production and P

released from organic manure mineralization increases its availability in soil (Gopalakrishnan, 2007). This increase in uptake may be due to increase surface area of soil contact, increased movement of nutrients into mycorrhizae, a modification of the root environment and increased storage. Mycorrhizae can therefore be much more efficient than plant roots at taking up phosphorus in the soil. The application of organic manure contributed to N uptake in the soil which was significantly high with pig dung and poultry manure application. Hedge (2001) stated that plants seem to be better than fungi at short circuiting nitrogen cycle. Manures applied to soil enhance N concentration in the soil and tend to initiate different conformational changes in growth of different plants varieties. This shows the effectiveness of organic manure application in the soil. Ca and Mg uptake showed inoculated plants increase than the control. The mycorrhizal inoculation and pig dung application showed an increase in Ca and Mg uptake. This is in agreement that AM fungi is not a fertilizer base but can only aid nutrient uptake such as N, P, K, Ca and Mg from soluble pool of nutrients available for plant absorption (Bagayote *et al.*, 2000; George *et al.*, 2000 and Ortas *et al.*, 1996). Manure handling and storage has being widely studied, however the quality is difficult to quantify due to differences in the quality of the sources. Mycorrhizal colonization rates differed depending on the host plant and the AMF species. Our study indicated a strong colonisation level with mycorrhizal inoculation, pig dung and poultry manure when compared with pig dung and poultry manure application and control. Faisal *et al.* (2010) reported that the concentration of P in plant shoots increase with increasing level of inoculum. Studies have proven that AM colonization is influenced by nutrient availability (especially P) in soil. Hence, percentage colonization of roots is as a result of increased acquisition of P by the mycelial network. We suggest that AMF becomes less effective in colonisation of roots when P concentration in the soil is high. However, mycorrhizal colonization rates differed depending on the host plant and the AMF species.

5.0 Conclusion.

The role of organic manure with or without mycorrhizal inoculation in this study resulted to increase in plant height and stem girth of pepper plant. The organic soil amendments have strategically different approach that relies on long-term solutions in the soil. The combined application of pig dung and mycorrhizal inoculation increase uptake of nutrients most especially in uptake of P. The application of organic materials to nutrient depleted soils goes beyond just increasing or replenishing nutrient element content of amended soils. Thus, a concomitant

use of mycorrhizal fungi with organic amendments is required to enhance flow of nutrients to crops. Lastly, high quality AM fungal strains with effective potential is required to have appropriate symbiotic association.

Acknowledgement.

The authors wish to thank the Soil Analytical Laboratory of the Agronomy department in the Faculty of Agriculture and Forestry, University of Ibadan for assistance in sample analyses.

Corresponding Author:

Professor O. Fagbola
Agronomy Department, Faculty of Agriculture and Forestry, University of Ibadan, Ibadan Nigeria.
E-mail: fagbola8@yahoo.co.uk.

References

- Mbagwu SC, Aggregate Stability and Soil Degradation in the Tropics. University of Nigeria Nsukka, Nigeria Lecture.2003.
- Swift MJ, Palm, CA, Soil Fertility as an Ecosystem Concept.2000. Pp 64-74.
- Duruigbo CI, Okereke-Ejiogu EN, Nwokeji EM, Peter-Onoh CA, Ogwudire VE, Onoh PA, Integrated remediation strategies for sustaining agrobiodiversity degradation in Africa. IOSR Journal of Agriculture and Veterinary Science (IOSR-JAVS) 2013. 3(4): 16-23.
- Spaccini RA, Piccolo JSC, Mbagwu TA, Zena, Igwe CA, Influence of the addition of organic residues on the carbohydrate content and structural stability of some highland soils in Ethiopia. Soil Use Management. 2002. 18:404-411.
- Egbuchua CN, Enujeke EC, Growth and yield responses of ginger (*Zingiber officinale*) to three sources of organic manures in a typical rainforest zone, Nigeria. Journal of Horticultural Forest 2013.5(7): 109-114.
- Ayeni LS, Omole T, Adeleye JO, Ojeniyi SO, Integrated application of poultry manure and NPK fertilizer on performance of tomato in derived savanna transition zone of South-West Nigeria. Science Nature 2010.8(2):50-54.
- Sanchez PA, Miller RH, Organic matter and soil fertility management in acid soils of the tropical soils. XVIII Congress of International Soil Science Society. 1986. P: 10.
- Smith SE Read DJ, Mycorrhizal Symbiosis. Academic Press, London United Kingdom. 1997.
- Rillig MC, Mummey DL, Mycorrhizas and soil structure. *New Phytology* 2006. 171: 41–53.
- Warnock DD, Daniel L, Mummey DD, McBride B, Julie-Major J, Lehmann J, Rillig MC, Influences of non-herbaceous biochar on arbuscular mycorrhizal fungal abundances in roots and soils: Results from growth-chamber and field experiments. *Applied Soil Ecology*. 2010.46: 450–45.
- Oseni TO, Shongwe NS, Masarirambi MT, Effect of arbuscular mycorrhiza (AM) inoculation on the performance of tomato nursery seedlings in vermiculite. *International Journal of Agriculture Biology*. 2010.12: 789–792.
- Van der Heijden MG, Streitwolf-Engel R, Riedl R, Siegrist S, Neudecker A, Ineichen K, Boller T, Wiemken A, Sanders IR, The mycorrhizal contribution to plant productivity, plant nutrition and soil structure in experimental grassland. *New Phytologist* 2006. 172:739-752.
- Smith SE, Anderson IC, Smith AF, Mycorrhizal associations and phosphorus acquisition: from cells to ecosystems. In: Plaxton W, Lambers H (Eds.), Annual Plant Reviews Volume 48: Phosphorus Metabolism in Plants, first ed. John Wiley & Sons, Ltd. 2015. pp. 409-440.
- Hodge A, Helgason T, Fitter AH, Nutritional ecology of arbuscular mycorrhizal fungi. *Fungal Ecology*. 2010. (3) 267-273.
- Whiteside MD, Garcia MO, Treseder KK, Amino acid uptake in arbuscular mycorrhizal plants. *Plos One*. 2012. (7): 147-643.
- Perner H, Schwarz D, Bruns C, Mader P, George E (2007) Effect of arbuscular mycorrhizal colonization and two levels of compost supply on nutrient uptake and flowering of pelargonium plants. *Mycorrhiza* 17:469–474.
- Fagbola O, Osonubi O, Mulongoy K, Contribution of arbuscular mycorrhizal AM) fungi and hedgerow trees to the yield and uptake of cassava in an alley-cropping system. *Journal of Agricultural Science (Cambridge)* 1998.131: 79-85.
- Javaid A, Riaz T, Mycorrhizal colonization in different varieties of Gladiolus and its relation with plant vegetative and reproductive growth. *International Journal of Agriculture and Biology* 2008.10: 278–282.
- Grubben GJH, El-Tahir IM, Capsicum species, In: Grubben GJH, Denton OA. (Editors). Plant Resources of Tropical Africa 2. Vegetables PROTA foundation, Wageningen, Netherlands. 2004. Pp: 154-163.
- Reinder HP, Learning together for organic farming: In magazine on low external input and sustainable agriculture, March, 2007, 23 No. 1.
- Gee GW, Bauder JW, Particle-size analysis. In *Methods of soil analysis; Part 1: Physical and mineralogical methods* (ed) Klute A, Madison, WI: Agronomy Society of America-Soil Science

- Society of America Publication Inc. SSSA 1986.383-411.
22. Nelson DW, Sommers LE, Organic carbon In: Page A. L., Miller, and Keeney, D. R (eds.) Methods of soil analysis Part 2. Agronomy 9 Madison WI. 1982: 538-580.
 23. Bremner JM, Mulvaney CS, Total nitrogen, In: Page Miller AL, RH, Keeney D. R (Ed): Methods of soil analysis Part 2. Agronomy. 9. Madison, WI, USA. 1982: pp. 149-157.
 24. IITA-International Institute of Tropical Agriculture, Automated and semi-automated of and plant analysis. Manual series. IITA, Ibadan, Nigeria.1982. pp 7.
 25. Kaira YP, Maynard DG, Method manual for forest soil and plant analysis (NOR-X-319). Edmonton, Canada: Northwest Region, Northwest Forestry Center. 1991.
 26. Cater MR, Soil sampling and methods of analysis. Ontario: Canadian Society of Soil Science. Chemical properties, growth, yield and nutrient status of tomato. African Journal of Agricultural Research 1993. (3):612-616.
 27. Carling DE, Riehle WG, Brown MF, Johnson DR, Effect of vesicular arbuscular mycorrhizal fungus on nitrogen reductase and nitrogenase activities in nodulating and non-nodulating soybeans. *Phytopathology* 1978.68: 1590-96.
 28. Phillips JM, Hayman DS, Improved procedures for clearing roots and staining parasitic and vesicular-arbuscular mycorrhizal fungi for rapid assessment of infection. *Transactions of British Mycological Society*. 1970. 55: 158-61.
 29. Giovanetti M, Mosse, An evaluation of techniques for measuring vesicular arbuscular mycorrhizal infection in roots. *New Phytol*. 1980(84): 489-500.
 30. Juo ASR, Moormen FR, Maduakor HO, Forms and pedogenic distribution of extractable Fe and Al in selected soils of Nigeria. *Geoderma* 1974.11:167-76.
 31. Murphy J, Riley JP, A modified single solution method for the determination of phosphate in natural waters. *Analytical Chimimicta Acta*. 1962. 27: 31-36.
 32. Novozamskey I, Houba VJG, van Eck R, van Vark W, A novel digestion technique for multi-element plant analysis. *Communication in Soil Science and Plant Analysis*. 1983. 14: 239-48.
 33. Adesina JM., Sanni KO, Afolabi LA, Eleduma AF, Academia Arena, 2014, 6(1): 9-13.
 34. Awosika OE, Awodun MA, Ojeniyi SO, Comparative effect of pig manure and NPK fertilizer on Agronomic performance of Tomato (*Lycopersicon esculentum* Mill). American Journal of Experimental Agriculture, 2015. 4(11): 1330-1338, 2014.
 35. Crews CE, Johnson CR, Joiner JA, Benefits of mycorrhizae on growth and development of three woody ornamentals. *Horticultural Science* 1978. (23):213-215.
 36. Lovato PE, Schuepp H, Trouvelot A, Gianinazzi S, Application of arbuscular mycorrhizal fungi (AMF) in orchard and ornamental plants. In: Varma A., Hock B. (eds.), *Mycorrhiza*. Springer-Verlag Berlin, 1995. pp. 443-467.
 37. Olawuyi OJ, Odebode AC, Babalola BJ, Afolayan ET, Onu CP, Potentials of Arbuscular Mycorrhiza Fungus in Tolerating Drought in Maize (*Zea mays* L). *American Journal of Plant Sciences*. 2014. 5: 779-786.
 38. Ewulo BS, Ojeniyi SO, Akanni DA, Effect of Poultry Manure on Selected Soil Physical and Chemical Properties, Growth, Yield and Nutrient Status of Tomato. *African Journal of Agricultural Research* 2008. (3): 612-616.
 39. Ikeh AO, Udoh EI, Uduak GI, Udounag PI, Etokeren UE, Response of cucumber (*Cucumis sativus* L.) to different rates of goat and poultry manure on an ultisol. *Journal of Agricultural Social Research* 2012. 12(2):132-139.
 40. Senjobi BA, Peluola CO, Senjobi CT, Lawal IO, Ande OT, Salami BT, Performance of *Cochorus olitorius* as influenced by soil type and organic manure amendments in Yewa North Local Government Area, Ogun State. *African Journal of Biotechnology*. 2010.9(33): 5309-5312.
 41. Ewulo BS, Hassan KO, Ojeniyi SO, Comparative Effect of Cowdung Manure on Soil and Leaf Nutrient and Yield of Pepper. *International Journal of Agricultural Research*. 2007. (2): 1043-1048.
 42. Erinle KO, Akande TY, Urhie J, Bitire TD, Effect of Manure compost on heavy metal translocation and bio-concentration factors in soils from an old municipal dumpsite. *New York Science Journal*. 2017: (10)4.
 43. Fenado WGD, Linderman RG, The effect of mycorrhiza (*Glomus intradices*) colonization on the development of root and stem rot (*Phytothora vignae*) of cowpea. *Journal of National Science Country*. Sri Lanka 1997.25(1):39-47.
 44. Grey C, William V, The Right away to Grow Fresh Vegetable all Year round. 2006. Macmillan publ. London.
 45. Akinrinde EA, Issues of Optimum Nutrient Supply for Sustainable Crop Production in Tropical Developing Countries. *Pakistan Journal of Nutrition* 2006 5(4):377-387 Asian Network for scientific information.

46. George CD, Nutrient Uptake In: Arbuscular Mycorrhizal: as Physiology and Function (Y. Kapulnik and D. D. Douds. Jr. eds). Kluwer Academic Publishers, Dordrecht, Netherlands. 2000. pp.307-343.
47. McArther DAJ, Knowles NR Influence of VAM and phosphorus nutrition on growth, development and mineral nutrition of potato. *Plant Physiologist* 1993.102:771 – 782.
48. Ortas I, Harris PJ, Rowell DR, Enhanced uptake of phosphorus by mycorrhizal sorghum plant as influenced by forms of nitrogen. *Plant and Soil* 1996. 184:255 – 264.
49. Subbarao GV, Wheeler RM, Levine LH, Stutte GW, Glycinebetaine accumulation, ionic and water relations of red-beet at contrasting levels of sodium supply. *Journal of Plant Physiology*. 2001. 158: 767-776.
50. Gopalakrishnan G, Vegetable crops Publisher, New India Publishing, ISBN, 8189422413, 9788189422417. Length. 2007. p. 343.
51. Hodge, A., 2001. Arbuscular mycorrhizal fungi influence decomposition of, but not plant nutrient capture from, glycine patches in soil. *New Phytologist* 151:725 – 734.
52. Bagayoko M, George E, Romheld V, Buerkert A, Effects of mycorrhizae and phosphorus on growth and nutrient uptake of millet, cowpea and sorghum on a West African soil *Journal Agricultural Science* 2000.135, 39–407.
53. Faisal M, Ahmad T, Srivastava NK, Influence of different levels of *Glomus macrocarpum* on growth and yield of chilli (*Capsicum annum L.*). *Indian Journal of Scientific Research*. 2010. (1):97–99.

5/18/2018