

# Toxin Production By Fungi Isolated From Rotten Pawpaw Fruits In Parts Of Imo And Abia States Of Nigeria

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**Abstract:** Culture filtrates of *B. theobromae*, *A. niger*, *A. flavus* and *R. oryzae* isolated in parts of Imo and Abia states of Nigeria contained toxic metabolites which elicited electrolyte leakages and discoloration on young leaf discs of pawpaw. Procedures for extraction of the metabolites based on solvent extraction are described. The sensitivity of the pawpaw leaf discs to the toxic metabolite preparations corresponded to the conductances and discolorations inferring their susceptibility to the pathogens in the bioassays conducted.

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**Key words:** Toxin, Fungi, PawPaw, Imo, Abia, Nigeria

## 1. INTRODUCTION

A great deal of information on toxins has been obtained from investigations of relatively serious from investigations of relatively serious plant pathogen relationships. Fungi noted for their saprophytic activity and weak parasites has received little attention. However some toxic metabolites cause disease symptoms on host plants and host plant parts in bioassay. *Aspergillus niger* have been known to produce Aflatoxins (Wogan, 1973) which induced electrolyte leakage in living plant tissues and altered cell permeability, (Kohmoto et. al; 1976). The principle of conductivity has been used in toxin detection due to the increase in electrolyte presence caused by the leakage of ions from host plant tissues attributed to the metabolite (Vidhyase Karan et. al; 1986).

In a survey of the prevalence of post-harvest fruit rots of *Asmilia tribola* (pawpaw) in Imo and Abia States of Nigeria, *Aspergillus flavus*, *Aspergillus niger*, *Botryodiplodia theobromae* and *Rhizopus oryzae* were constantly isolated from affected fruits and individually produced fruit rots in inoculation exercise (Ezeibekwe, 1993).

The production of toxins by these isolates was studied using the principle of electrolyte conductivity in bioassay.

## 2. MATERIALS AND METHOD

### Culture Medium and Extraction of Toxic Metabolite

Pawpaw 200g was washed with clean water, peeled and sliced for preparation of pawpaw broth. The slices were boiled until they become soft and then strained through cheese cloth. Dextrose 20g was added and the extract made up to one litre with distilled water. The medium (30ml) was dispensed in Erlen

meyer flasks, and sterilized in the autoclave at 121°C for 15 minutes.

The medium was inoculated with mycelial discs 8mm with each of *B. theobromae*, *R. oryzae*, *A. niger* and *A. flavus* isolated from infected pawpaw fruits. Potato dextrose agar discs 8mm were put into the control flasks. Each set up was in three replicates. After two weeks of incubation at 25°C, the cultures were filtered with No. 1 filter paper and the filtrate stored in MacCarthy bottles at 10°C.

The culture filtrate was extracted with two volumes of acetone. The acetone supernatant was decanted and the precipitate left in the test tube. Any acetone left on the precipitate was removed by evaporation at 30°C. The supernatant was evaporated in a water bath at 30°C and the precipitate redissolved in methanol (10mls).

The supernatant and the precipitate were each diluted with 5ml of deionized water and tested for toxin activity. The controls were similarly treated.

### Test For Toxin Activity (Bioassay)

Young fully expanded leaf, 21 days old was used. The leaf was cut into small discs 1 cm by 5mm in size and rinsed in several changes of distilled water. The extract 1ml from each isolate was diluted with 50ml of deionized water and put in sterile test tubes.

A leaf disc was put in each test tube and incubated for 30 minutes at 25°C on a shaker set at 100 strokes per minute. Five hours after, conductance of the ambient solution was determined with a conductivity meter.

The control and water blank were subjected to the same screening. The conductance in umhos of the blank (water) was subtracted from those of the control and the toxin infiltrated leaves to determine the increase in the electrolyte leakage induced by toxin. All tests were conducted in three replications.

**Extraction Of Toxin From Infected Pawpaws**

Healthy pawpaw fruits were surface sterilized by dipping in 10% sodium hypochlorite solution for 10 minutes, followed by washing with distilled water. The fruits were cut into 10mm diameter discs and placed in a sterile container. The discs were then inoculated with the pathogens *Botryodiplodia theobromae* (BT), *Rhizopus oryzae* (RO), *Aspergillus niger* (AN), and *Aspergillus flavus* (AF). The discs were incubated in a humid chamber at 25°C for 72 hours. The supernatant was extracted from the discs and used for bioassays.

**Fig. 1: Bioassay with Pawpaw Leaf Discs Conductivity of Acetone Precipitate**

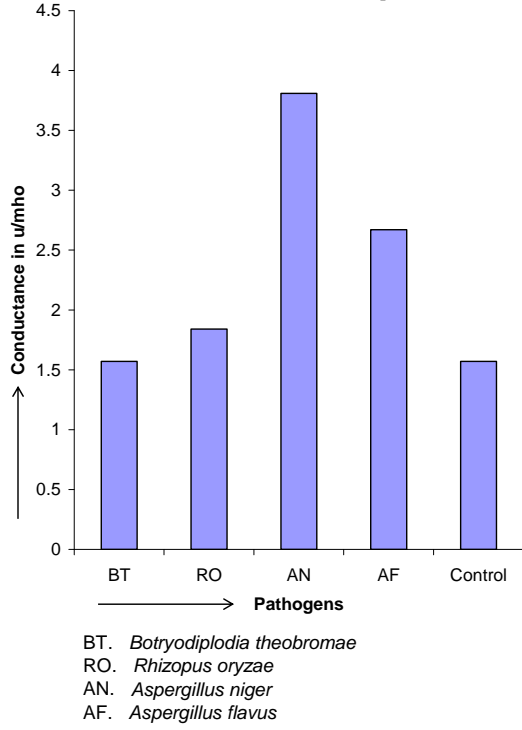


Fig. 1: Bioassay with Pawpaw Leaf Discs Conductivity of Acetone Precipitate

**Extraction of toxins from infected pawpaw fruits**

Infected pawpaw fruits were surface sterilized and cut into 10mm diameter discs. The discs were inoculated with the pathogens *Botryodiplodia theobromae* (BT), *Rhizopus oryzae* (RO), *Aspergillus niger* (AN), and *Aspergillus flavus* (AF). The discs were incubated in a humid chamber at 25°C for 72 hours. The supernatant was extracted from the discs and used for bioassays.

**Fig. 2: Bioassay with Pawpaw Leaf Discs Conductivity of the Acetone supernatant**

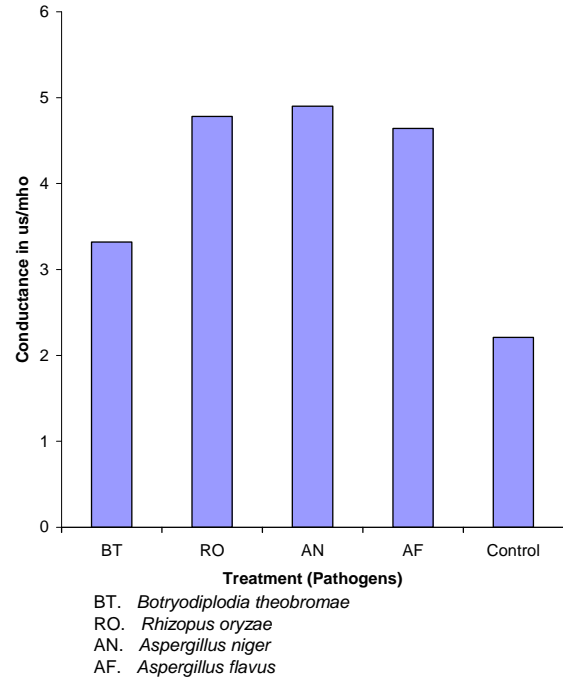


Fig. 2: Bioassay with Pawpaw Leaf Discs Conductivity of the Acetone supernatant.

**Table 1: Effect of Metabolite Extracted from Pawpaw Fruit Inoculated with *Asperillus niger* on Pawpaw Leaf Discs**

Electrolyte leakage test by conductance						
Anova table						
Source of Variation	df	ss	ms	f	f	
Acetone Supernatant	2	0.84	0.42	2.4	0.05*	0.01**
Acetone Precipitate	2	0.56	0.28	0.23	0.05*	0.01**
Error	4	0.70	0.175			
<b>Total</b>	<b>8</b>	<b>2.10</b>				

\* non significant ( $P < 0.05$ ,  $V_1 = 2$ ,  $V_2 = 4$  df)\*\* Highly insignificant ( $P > 0.01$ ,  $V_1 = 2$ ,  $V_2 = 4$  df)**Table 2: Effects of Metabolite Extracted from Pawpaw Fruit Inoculated with *A. flavus* on Pawpaw Leaf Discs Electrolyte leakage test by conductance**

Anova table						
Source of Variation	df	ss	ms	f	f	
Acetone Supernatant	2	0.90	0.45	1.2	0.05*	0.01**
Acetone Precipitate	2	0.11	0.55	0.10	0.05*	0.01**
Error	4	2.20	0.55			
<b>Total</b>	<b>8</b>	<b>3.21</b>				

\* non significant ( $P < 0.05$ ,  $V_1 = 2$ ,  $V_2 = 4$  df)\*\* Highly insignificant ( $P > 0.01$ ,  $V_1 = 2$ ,  $V_2 = 4$  df)

#### 4. DISCUSSION

The results of the investigation on bioassays involving the four pathogens show a significant variation in their toxic effects as revealed by the electrolyte leakages, figures 1 and 2. *A. niger* and *A. flavus* generally appeared to produce more toxic effect than *B. theobromae* and *R. oryzae*. Fig. 1 and 2. The result of the bioassay work using extracts from the various zones on and around the inoculation points show observable differences in toxicity effect between replicates in both the experimental and the controls. Vidhyase Karen et. al; (1986) stated that the maximum dilutions at which visible symptoms were noticed was considered as the dilution and point of activity of the toxin and the minimum concentration required, to induce brown spot symptoms on rice by toxin extracted from *Helminthosporium oryzae* was 1.04 ug/ml when purified with chloroform and 0.63 ug/ml when purified with charcoal. In the same work they held that in electrolyte leakage bioassays, the effectiveness of each step in removing contaminating materials from the toxins whose action was seen to have increased with each step of purification the conductance of a solution is the sum of the contributions of all the ionic species present, and hence not only depend on the species for

which one is analyzing. This explains part of the factors that likely affected the purity and the effect of the target metabolite phytotoxins.

The toxic metabolite extracts showed an ability of eliciting electrolyte leakages in leaf discs bioassay at varying depress. The invitro extracts from the pathogens revealed significant differences in toxicity. There were no significant differences in toxicity between the zonal extractions in the invivo tests,  $P = 0.05$ .

#### 5. CONCLUSION

The production of toxic metabolites by the fungal associates of pawpaw rots appeared to be important in the mechanism of disease spread. Although the toxic extracts need to be subjected to further purification, Aflatoxin is known to be produced by *A. flavus*.

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