Studies On Adsorbent Dosage, Particle sizes and pH constraints on Biosorption of Pb(II) and Cd(II) ions from Aqueous Solution using Modified and Unmodified *Crasstrotrea Gasar* (bivalve) Biomass.

*Osu Charles . I. and ¹S.A. Odoemelam

*Department of Pure and industrial chemistry, University of Port Harcourt, PMB, 5323, Port Harcourt, Rivers State. Nigeria. ¹Department of Chemistry, Michael Okpara University Of Agriculture, Umudike, Nigeria charsike@vahoo.com

Abstract: Biosorption of Pb(II) and Cd(II) ions from aqueous solution did not improve by chemical modification of Crasstrotrea Gasar biomass with oxalic acid and ethanol. Unmodified Crasstrotrea Gasar biomass was proved to be very effective adsorbent for the removal of Pb (II) and Cd(II) ions from aqueous solution. As the particle sizes increases from 106-500µm, the percentage removed increases. The particle size of 500µm showed the highest removal. Also the percentage metal ions sorbed increases, as the dosage of the adsorbent increases from 0.2g to 1.0g and further increase of the adsorbent did not provide more increment in the percentage removal of Pb(II) and Cd(II) ions was at maximal at pH 10. Since the Crasstrotrea Gasar biomass is cheap, abundantly available and environment friendly, so it can be used for metal ion removal from aqueous solution using a simple and cost effective treatment procedure.

[Osu Charles . I. and S.A. Odoemelam. Studies On Adsorbent Dosage, Particle sizes and pH constraints on Biosorption of Pb(II) and Cd(II) ions from Aqueous Solution using Modified and Unmodified *Crasstrotrea Gasar* (bivalve) Biomass. *Rep Opinion* 2014;6(12):67-72]. (ISSN: 1553-9873). <u>http://www.sciencepub.net/report.</u> 9

Key words: Biosorption, heavy metals, agricultural by-product

1.Introduction

The concentration levels of heavy metals in the environment are continually changing due to man's activities. These have generated a lot of interest in pollution studies. Major natural sources of these metals are rock-weathering, releases from terrestrial and submarine sediments. Heavy metals are often found in crude oils in small quantities are removed during the refining process (Cutler, 2007).

Heavy toxic metals entered into the environment through wastewater from metal plating industries and industries of Pb, Cd - Ni batteries, phosphate fertilizer, mining, pigments electroplating, Corrosion, and other industrial waste (Ahalya, et al.,2005). The discharge of heavy metals into the environment has become a matter of concern because of their hazardous nature. Heavy metals are toxic and pose a threat to man and the environment. Adsorption has now been recognized as an effective and economic method for the removal of pollutants from the environment (Martins, et al 2006). Adsorption is operative in most natural, physical, biological and chemical system, and is widely used in industrial applications such as activated charcoal, synthetic resins and water purification. Similar to surface tension, adsorption is a consequence of surface energy (Abdul-Ghaffar, 2006).

Many adsorbents, which include, pillared clay (Vinoid and Anorudhan 2001); Cassava waste

(Ahalya et al. 2005), Caladium bicolor (Igwe, and Abia, 2006), Maize cob and husk (Igwe and Abia, 2003) have been reported.

In this present study, Pb(II) and Cd(II) ions removal from aqueous solution using modified and unmodified *Crasstrotrea Gasar* biomass were examined. The constraints of pH, particle sizes and dosage of the adsorbent were also studied.

2.Materials And Methods

Collection and preparation of samples.

The marine animal shells was collected from Okirika village of Rivers state of Nigeria and was washed thoroughly, clean of adhering dirt, rinsed thoroughly with de-ionized water and dried in the oven at 105 °C for two days. The process helped to remove moisture present in the material, which was different from the chemisorbed water normally, released during carbonization reactions of elevated temperature. Higher temperature of 150 °C was used to ensure complete dehydration. The purpose of dehydration was to effect de-sorption of physically absorbed water which may catalyze the decomposition of the char carbon thereby producing a very low carbon.

 $H_2O + C - H_2 + CO$

After oven drying, the sample was macerated into powdered form. The powdered form of the sample obtained was first sieved through a 1000 μm mesh and then through 500 $\mu m,$ 250 $\mu m,$ and 106- μm meshes.

Chemical activation of the adsorbent.

The sieved sample was soaked in 0.3 M HNO_3 for 24 hours at room temperature. The adsorbents was then filtered through what man no. 41 filter paper and rinsed thoroughly with de- ionized water to maintain a pH of 7.4. The rinsed adsorbent was kept in an oven at 100°C for 12 hours for the moisture and finally stored in a tight plastic container.

The treatment of the absorbent with $0.3 \text{ M HN}0_3$ aids to oxidize the adhering organic material, removal of any debris or soluble bio-molecules that might interact with the metal ions during the sorption. This process is called chemical activation.

Chemical modification of the absorbent.

The oven dried chemical activated sample was weighed and divided into two parts. 200g of first part was left untreated was labeled the unmodified sea animal shell A (USASA) where A is shell of Crasstrotrea Gasar(Bivalve) .400g of second part was treated with 5 dm³ of 3 M Oxalic acid solution for 24 hours at 28° C in a well ventilated place according to the method of Okieimen and Okieimen, (2001). The biosorbent was then esterified (Krishnakumar et al., 2007; Drake et al., 1996) by suspending the adsorbent in 2.6 L of ethanol and 240 Ml of concentrated hydrochloric acid. The mixture was shaken for 5 hours at 150 rpm, filtered and the residue was washed thoroughly with distilled water to maintain pH of 6.9 and finally oven dried at 100° C, stored and was labeled modified sea animal shell A (MSASA). Surface functional group can be introduced via chemical modification (Okieimen et al., 1991; Drake et al., 1996; Krishnakumar et al., 2007).

All reagents used were analytical grades, purchased and used without further purification.

Effect of particles size of the absorbents.

Initial concentration of 4 Mg/L of the metal ions (Pb²⁺ and Cd²⁺) solution were prepared. 50 MI of each these metal ions solution were mixed and put in a conical flask containing 1 g of each of the 850 μ m, 500 μ m, 250 μ m, and 160 μ m particles sizes of modified and unmodified absorbent. They were corked and left to stand for 10 minutes in a rotary shaker at a constant speed of 150 rpm, temperature of 29° C and optimum pH of 7.4. At the end of the time, the mixtures were filtered. The various metal ions content of the filtrates were determined by an air acetylene flame atomic absorption spectrophotometer (UNICAM SOLAAR 32).

Effect of pH

The pH of 50 Ml of metal ions solutions with initial concentration 4 Mg/ L were adjusted to values 2, 4, 7, 10 and 12 by adding 0.1 M HCl or 0.1 M NaOH solutions. Then 1 g of each of the adsorbent (Both modified and unmodified) with particle size 500 μ m was added and agitated in a shaker for 60 minutes at 29°C. Then, the suspensions were filtered and the metal ions content of the filtrates were determined using an air acetylene atomic absorption spectrophotometer (UNICAM SOLAAR 32).

Effect of adsorbent dosage

Initial concentration of 4 Mg/L of the two metal ions (Pb²⁺ and Cd²⁺⁾ solution was prepared. 50 MI of each these metal ions solution were mixed and put in a conical flask containing 0.2 g, 0.4 g, 0.6 g, 0.8 g, 1.0 g, and 1.2 g respectively of the 500 μ m, particles size of modified and unmodified absorbent. The mixtures was corked and left to stand for 10 minutes in a rotary shaker at a constant speed of 150 rpm at a temperature of 29° C and optimum pH of 7.4. At the end of the time, the mixtures filtered. The various metal ions content of the filtrates were determined by an air acetylene flame atomic absorption spectrophotometer (UNICAM SOLAAR 32).

3. Results





Figure 2: Effect of pH on the Biosorption of Pb(II) and Cd(II) lons from Aqueous Solution on USASA and MSASA





4. Discussion.

The experimental results for the adsorption of Pb(II) and Cd(II) ions on the unmodified and modified adsorbent with different particle sizes were plotted and reported above. Figure 1 show the plot of the percentage of the metal ions removed against the particle sizes of the adsorbent. From the plots, it was observed that as the particle size increases from 106 - 500μ m, the percentage removed increases and went down at particle size 850 µm. Although, one would expect that the smaller particle size should give a greater percentage removal because of the surface area, but as the particle size increased, the number of micro pores also increases.

The increase in micro pores increases the number of the accessible sites, hence increases the percentage adsorbed. This is similar to the results of Eneida et al., (2005), who reported that there was no influence of bio-sorbent size on adsorption capacity. They reasoned that the two particle sizes of adsorbent they tested were actually of the same thickness (dimension which determines the diffusion distance). This is so because size grading of ground biomass particle by standard sieves works on the length and width dimensions. Leusch et al., (1995), showed that larger biomass particles of sargassum fluitans and Ascophylum nodosum, had higher metal uptake than smaller particles. Igwe and Abia (2006), showed that as the particle size increase from 0.15 mm to 0.85 mm, the amount adsorbed also increased, but from

particle size 0.85 mm to 1.00 mm, the amount adsorbed decreased.

From the results of this study, the percentages of metal ion removed on unmodified sorbent were higher than modified sorbent. This means that modification of the adsorbent did not improve the adsorptive capacity of the adsorbent used. In this study, ethanol in the presence of concentrated HCl was used for the esterification of the carboxylic acids present on the cell wall. Thus, the introduction of ester groups such as $RCOOCH_2CH_3$ alters the surface characteristics of the adsorbents; hence affect the percentage of metal ion removed (Motoyuki, 1990; Krishnakumar et al., 2007).

Metal uptake by bio-sorption is reported to occur through interactions with functional groups native to the biomass cell wall (Goksungur et al., 2005). It is metabolism-independent and proceeds rapidly within some minutes by any one or combination of the following metal binding mechanisms; coordination, complexation, ion exchange, physical adsorption (e.g. electrostatic) or inorganic micro -precipitation. The mechanism of metal bio-sorption is complicated, the status of (living and non-living), biomass types of biomaterials, properties of metal-solution chemistry, ambient/environmental condition such as pH, will all influence the mechanism of metal bio-sorption (Wang and Chen, 2006).

The decrease in the percentage adsorbed on modification is probably as a result of steric hindrance caused by the attachment of the ester group on the adsorbent surface (Igwe et al., 2005). Therefore, this reduces the ease with which the hydrogen ions were lost, hence, decreasing the contribution by columbic interaction of the metal ions with the surface groups to the adsorption capacity. This was similar with the work of Beveridge and Murray, (1980); Jianlon, (2002); Chen-Shoel et al., (2002), but not similar to the work carried out by Tiemann et al., (1999).

The adsorption trend is Pb^{2+} (99.99 ± 0.004 %) > Cd^{2+} (99.50 ± 0.006). Thus, the influence of bio-sorbent size on the metal uptake seems to be a function of both the type of biomass and metal ion. Other researches have also reported the influence of particle size on the biomass adsorption (Yang and Volesky, 1999; Igwe and Abia, 2003).

The adsorption of Pb(II) and Cd(II) ions from aqueous solution on modified and unmodified sea animal shells in this research were found to be a highly pH dependent process. pH of the solution, in fact determines the chemistry, degree of ionization and speciation of metal ions and also affects the surface charge of the adsorbent. In this experiment, the adsorption behaviors of the aforementioned metal ions were studied at different pH values. The results obtained are shown in figure 2. From the plots, it was observed that the percentage of metal ion removed had a characteristic change as pH changes. The percentage of metal ion removed increases as pH increase from 2 - 7 and increased to maximum at pH 10, then decreased at pH 12 for the metal ions. This is similar to work carried out by Abdul-Ghaffar (2006). At low pH, the cell surface sites are closely linked to the H⁺ ions, thereby making these unavailable for other cations. However, with an increase in pH, there is an increase in ligands with negative charges which results in increase binding of cations (Krishnakumar et al., 2007).

pH is an important parameter for adsorption of metal ions from aqueous solution because it affects the solubility of the metal ions concentration of the counter ions on the functional groups of the adsorbate and the degree of ionization of the adsorbate during reaction. It is now established that heavy metals are taken up from water predominantly by ion exchange and complexation reactions (Eneida et al., 2005).

According to Gardea-torresdey (1996), pH dependent binding suggests that metal ions are adsorbed by biomass through carboxyl, carbonyl or hydroxyl ligands. This is similar to the findings with the adsorbents used. The addition of NaOH raises the pH before equilibration, enhanced sorption of metal ions by the biomass. NaOH hydrolysis further showed that metal ion adsorption occur through carboxylic or hydroxylic ligands of the adsorbent (Gardea-Torresdey et al., 1998d).

At higher pH, the coordinating ligands may be deprotonated and available to adsorb positively charged ions such as Pb(II) and Cd(II) ions through an ion exchange mechanism. At lower pH, the hydroxyl groups may become protonated and can no longer attract the positively charged ions.

The effect of pH on metal bio-sorption has been studied by many researches and the results demonstrated the increasing pH values, as algae biomass (Kratochvil et al., 1998), as coconut fiber and sawdust (Igwe et al., 2005), as maize cob (Igwe and Abia, 2003), as activated carbon (Ansari and Sadegh, 2007).

Igwe et al.,(2005), reported that the extent of hydrogen ion exchange would depend on the relative concentrations of the exchangeable hydrogen and hydrogen ion concentration of the medium. According to Low et al.,(1995), at low pH values, the surface of the adsorbent would be closely associated with hydronium ions, which hinder the access of the metal ions to the surface functional groups. Consequently, the percentage of metal ion removed decrease at low pH.

Figures 3 showed the variation of distribution coefficients (Kd) of Pb(II) and Cd(II) sorption between unmodified and modified adsorbents as a function of pH. It was further observed that as the pH increased the distribution coefficient, K_d , which indicates the amount of metal bound to the biomass, also increases with or without modification, with higher binding of the metal ions occurring between pH 8 – 10.

The effect of dosage of the adsorbents for the adsorption of metal ions studied was shown on figures 4. From the figure, it was observed that the percentage removal of the metal ions increases with increasing the adsorbents doses from 0.2 g to 1.0 g and further increase of the adsorbent doses did not provide more increment in the Percentage of the metal ion removed. Therefore, 1.0 g dose of the adsorbents were used in all the studies.

The increase in the percentage of the metal ion removal with increase in adsorbent dose is due to the greater availability of the exchangeable sites or surface area at higher concentration of the adsorbent. Non-significant increase observed when the adsorbent doses were increased from 1.0 g to 1.6 g, suggests that after a certain dose of adsorbent, the maximum adsorption is attained and the amount of ions bound to the adsorbent and the amount of free ions remains constant even with further addition of the dose of adsorbent. This is similar to studies reported by Abdel-Ghani et al., (2007), Murat Teker and Mustafa Imamoglu (1999) and Alok Mittal, (2006).

Conclusion

Bio-sorption of Pb(II) and Cd(II) ions from aqueous solutions depend significantly on pH, particle size and dosage of the adsorbent used. Crasstrotrea Gasar biomass, cost effective and environment friendly adsorbent was proved to be a very good adsorbent for the removal of Pb(II) and Cd(II) ions from aqueous solutions. At pH 10, adsorption was at maximal with 500µm particle size and 1.0 g of the adsorbent. Adsorption cannot be improved by chemical modification with oxalic acid and ethanol (esterification).

Corresponding Author:

Dr. Osu, Charles Ikenna Department of pure and Industrial Chemistry, University Of PortHarcourt, Rivers State, Nigeria. E-mail: <u>charsike @yahoo.com</u>

References

- 1. Abdel-Ghani, N. T., M. Hefray, G. A. F. EL-Chaghaby. Removal of lead from aqueous solution using low cost abundantly available adsorbent. Int. J. Environ. Sci. Tech. 2007; 4(1): 67:73.
- 2. Abdul-Ghaffar. Removal and stabilization of chromium metal ions from industrial effluents. EJEAFCHE, 2006; 5 (2): 1286-1295.
- 3. Abia, A. A., Horsfall, M. Jnr. And O. Didi. The use of chemically modified and unmodified cassava waste for the removal of Cd, Cu, and Zn ions from aqueous solution. Bioresour. Tech., 2003; 90:345-348.
- Ahalya, N., Kanamadi, R. D. and Ramachandra, T. V. Biosorption of chromium(vi) from aqueous solutions by the husk of Bengal from (cicer arientinum) Electron. Journal Of Biotechnol. 2005; 8(3): 257-264.
- 5. Alok Mittal. Removal of the dye, amaranth from waste water using hen feathers as potential adsorbent. Electron. J. Environment. Agric. Food Chem. 2006; 5(2): 1296-1305.
- Ansari, R. and M. Sadegh. Application of activated carbon for the removal of arsenic ions from aqueous solution. E-Journal of Chem. 2007; Vol. 4(1): 103-108.
- Beveridge, T. J. and Murray, R. G. E. Sites of metal deposition in the cell wall of bacillus subtilis. J. of Bioethenol. 1980; Vol. 141, no 2: 876-887.
- Cohen-Sheol, N. Ilzycer, D. Gilath, I. and Telor, E. The involvement of pectin in Sr(II) ion biosorption by Azolla. Water Air and Soil Pollution. 2002; Vol. 135(1): 1-4.
- Cutler, J. C. Hydrocarbon Chemistry: In: Encyclopedia of Earth. Eds. (Washington, D. C. Environmental Information Coalition, National Council for Science and the Environment). 2007
- Drake, Lawrence, R., Lin, Shan., Rayscon, Garry D. and Jackson, Paul J. Chemica modification and metal binding studies of Datura innoxia. Environmental Science and Toxicol. 1996; Vol. 30 (1): 110-114.
- Eneida, S. C., Celia, R. G. T. and Teresa, M. K. R. Biosorption of chromium(III) by sargassum sp. Biomass. Electron. J. Biotechnol. 2005; 5:1-7.
- Gardea-Torresday, J. L., Tongan, D. J. M., Salvador. Copper adsorption by esterified and unesterified fractions of sphagnum peat moss and its different hormic substances. J. Hazard. Matter 1996; 48:1191-206.
- 13. Gardea-Torresdey, J. L., Tiemann, K. J. Dokken, K. and Gamez, G. Investigation of metal binding in Alfalfa Biomass through

chemical modification of Amino and sulfhyrydryl ligands. Proceedings of the 1998 Conference of Hazardous Waste Research, Texas, U.S.A. 1998d; 239-248.

- Goksungur, Yekta, Uren, Sibel and Guvenc, Ulgar. Biosorption of Cadmium and lead ions by ethanol treated waste baker's yeast biomass. Bioresource Technol. 2005; Vol. 96(1): 103-109.
- 15. Igwe, J. C. and A. A. Abia. Sorption kinetics and intra-particlate diffusivity of As(III) bioremediation from aqueous solution, using modified and unmodified coconut fiber Ecl. Quim., SaO. Paulo, 2006; 31(3): 23-29.
- Igwe, J. C. and A. A. Abia. Maize cob and husk as adsorbents for removal of Cd, Pb, and Zn ions from waste water. Physical Sci. 2003; 2:83-94.
- 17. Igwe, J. C., Ogunewe, D. and Abia A. A. Africa J. of Biotechnol. 2005; Vol. 4(10): 1113-1116.
- Jianlong, Wang. Biosorption of Copper(II) by chemically modified biomass of saccharomyces cerevisal. Process Biochemistry. 2002; vol. 37, no. 8; 847 – 850.
- 19. Kratochvil, D., P. Pimentel and B. Volesky. Removal of trivalent chromium by seaweed biosorbent. Environ. Sci. Technol., 1998; 32:2693-2698.
- KrishnakumanParvathi, Ramachandramurtty Nagendran and Redhakrishnan Nreshkumar. Lead biosorption onto waste beer yeast byproduct, a means to decontaminate effluent generated from battery manufacturing industry. Electron. J. Of biotechnol. 2007; Vol. 10. No. 92-104.
- 20 Leusch, A., Z. R. Holan and B. Volesky, Biosorption of heavy metals (Cd, Cu, Ni, Pb, Zn) by chemically reinforced biomass of

6/18/2013

marinealgae. J. Chem. Technol, Biotehcnol., 1995, 62: 279- 288.

- 21 Low, K. S., C. K. Lee and A. C. Leo. Removal of metals from electroplating waste using banana pith. Bioresour. Technol., 1995; 51: 227-231.
- 22 Martins, Bruno, L., Curz, Clando, C. V., Luna Aderval, S. and Henriques, Cristrane A. Sorption and desorption of Pb(II) ions by dead sargassum sp. Biomass. Biochemical Engineering Journal, 2006; Vol. 27 no. 3; 310-314.
- 23 Motoyuki, S. Adsorption Engineering. Elsevier Sci. Publishers. 1990; .5-61.
- 24 Murat Teker and Mustafa Imamoglo and Onger Saltabas. Adsorption of copper and cadmium ions by activated carbon from rice hulls. Turk. J. Chem. 1999, 23:185-191.
- 25 Okieimen, C. O., Okieimen, F. E. Enhanced metal sorption by groundnut (Arachis hypugea) husks modified with thioglycolic acid. Bulletin of Pure and Applied Sciences, 2001; 20c(i), 13-20.
- 26 Tiemann, K. J., Gardea torresdey, J. L. Sias, S., Gamez, G., Rodringuez, O., Renner, M. W. and Furenlid, L. R. Study of the ligands involve in the metal binding to Alfalfa biomass. Environ. Sci. tehncol. 1999; Vol. 31, 182A – 186A.
- 27 Vinoid, V. P. and T.S. Anorudhan,.. J. Chem. Technol. Biotechnol. 2001; 7; 92-101.
- 28 Wang, Jianlong, and Chen, Cann. Biosorption of heavy metal by Sacharomyces Cerevisiae. A review. Biotechnology Advances, 2006; Vol. 24 (5): 427-451.
- 29 Yang, J. and B. Volesky. Biosorption of uranium on Sargassum biomass. Water Res. 1999; 333:3357-3363.