

## Evaluation of genetic diversity among some wild populations of *Achillea bieberstenii* Afan. from Iran using morphological and agronomical traits

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**Abstract:** In this study, genetic diversity among 25 populations of *Achillea bieberstenii* Afan. collected from 12 provinces of Iran using a total of 34 morphological and agronomical characteristics including 30 quantitative and 4 qualitative traits was evaluated as a important step for possible use in the breeding programs of this medicinal plant. For this purpose, of each population 6 plants were studied. According to the results of analysis of variance, there were significant differences among the studied populations for most of traits. Also, significant positive and negative correlations were observed among evaluated traits. Based on the constructed dendrogram, all populations were clearly divided into 11 main clusters. Groups mainly have differences in yield and yield components. The importance of this work on the breeding potential of populations in question is discussed.

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### 1. Introduction

Morphological variability is a characteristic of all organisms and one of the basic characteristics in the world. Measurement, description and analysis of variations are fundamental steps to answer questions of biological adaptability (Ge and Hong, 1995). Morphological characters are the outside exhibition of organism, and in the natural habitats, they are not only affected by the genetic background of the species, but also by the environmental conditions as well. Plants have the potential to response to the changed environments by changing their morphology and there for, the intra-specific variation in plant characteristics is usually regarded as the adaptive mechanism to different environments (Mal and Doust, 2005). Studies on the morphological variations of plant species according to the habitat differences suggest their relationships with environmental factors clearly, and help us understand the manner, mechanism and influencing factors of plant adaptation and evolution (Yang, 1991).

The genus *Achillea* L. (commonly known as yarrow) belongs to the Aster family (Asteraceae) and comprises more than 100 species worldwide (Rahimmalek et al., 2009). These often medicinal and rhizomatous perennial plants are native to Europe, Western Asia and North Africa although they are also found in Australia, New Zealand and North America (Huber Morath, 1986; Chevallier, 1996). In traditional systems of medicine, *Achillea* species have a long

history of use as medicinal plants mainly due to their anti-inflammatory, anti-spasmodic, diaphoretic, diuretic, carminative, tunic, vermifugal and emmenagogic properties and are used as a cure for hemorrhage, pneumonia, rheumatic and abdominal pains, stomach-ache and wounds (Zargari, 1996; Baris et al., 2006; Esmaeili et al., 2006). Nowadays, different medicinal properties of these plants such as spasmolytic, choleric, anti-inflammatory and wound healing are documented (Benedek, 2007). In recent years, the anticancer activity of essential oils isolated from some *Achillea* species has been reported and shown that they can modulate macrophages activities (Paulo, 2005). Due to hair growth promotion property, yarrow is used in cosmetic industries for production of hair shampoos as well creams (Karlova, 2001; Lewis, 2006).

Iran represents a significant source of germplasm of different medicinal plant species, particularly for the genus *Achillea* (Zargari, 1996). In the flora of Iran, the genus *Achillea* is represented by 19 species, of which seven are endemic (Huber Morath, 1986). One of these species is *A. bieberstenii* Afan. which occurs naturally in many parts of the country in the central, North, Northwest, West and Northeast with the local name of "Bumadarane Zard" (Huber Morath, 1986; Mozaffarian, 1996). This plant is a perennial villose herb with 10-100 cm height and radiate heads which are borne in large dense compound corymbs on the erect stems (Huber Morath, 1986).

*A. biebersteinii* is used in folk medicine of Jordan because of its carminative properties, while in Turkey the plant is also used for abdominal pain, stomach-ache and for wound healing (Bader et al., 2003). To date, many investigations considered the volatile oil and extract of *A. biebersteinii* from the chemical constituents to biological activities points of view (Rustaiyan et al., 1998; Bader et al., 2003; Sokmen et al., 2004; Baris et al., 2006; Esmaili et al., 2006; Kordali et al., 2009; Rahimmalek et al., 2009). Based on the results of these studies, the plant has considerable different biological activities including antibacterial, antifungal, antioxidant, insecticidal, herbicidal and wound healing.

In Iran, most of the studies on the genus *Achillea* were done using one species originated from a limited geographical area and there is especially no comprehensive research considered variation in morphological characteristics of wild populations of *A. biebersteinii*. Since Iran is one of the important origins of genus *Achillea* and there is substantial diversity in *A. biebersteinii* plants in the country, therefore, the main aim of the present study was to

expand the knowledge on morphological and agronomical characteristics of 25 populations of *A. biebersteinii* growing wild in the different parts of Iran in order to find promising populations of this species which have the potential to be used as initial materials for breeding programs.

## 2. Material and Methods

### 2.1. Plant materials

*A. biebersteinii* plants were collected at the full flowering stage from their natural habitats in different parts of Iran between April and May 2009 (Fig 1). Sites of collections were determined based on previous collections cited in Flora Iranica (Huber Morath, 1986) and local information as well. Geographical and climatic conditions of each habitat were obtained from the nearest meteorology station (Table 1). Plant materials were identified at the herbarium of department of horticultural sciences, University of Tehran, Karaj, Iran.

Figure 1. Collection sites of studied *Achillea biebersteinii* populations. For codes see Table 1.

Table 1. Origin, geographical, climatic of natural habitats of 25 Iranian wild populations of *A. biebersteinii*

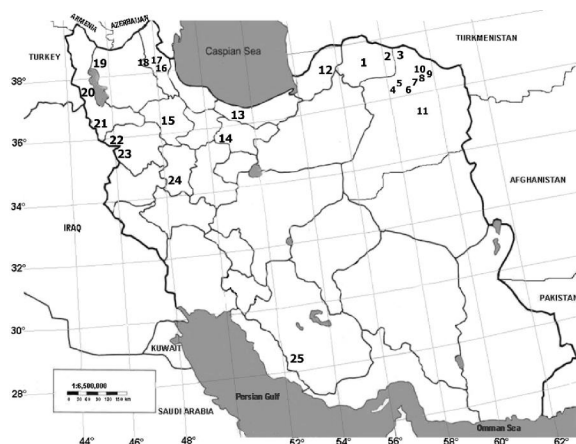
No.	Site name (origin)	Abb.	Longitude	Latitude	Altitude (m)	T <sub>min</sub>	T <sub>max</sub>	H <sub>min</sub>	H <sub>max</sub>	H <sub>total</sub>	P <sub>annual</sub>	Sun
1	Havar, North Khorasan	H	E 57°11'	N 37°28'	2980	6.8	19.7	0.4	0.8	0.4	272.4	2714
2	Goolool, North Khorasan	GOL	E 58°11'	N 37°37'	2100	6.8	17.5	0.4	0.7	0.53	252.7	2714
3	Chelmir, Khorasan-e-Razavi	CH	E 58°34'	N 37°31'	1584	6.8	19.7	0.4	0.8	0.58	272.4	2714
4	Adag, Khorasan-e-Razavi	AD	E 58°53'	N 36°11'	1260	6.7	21.8	0.3	0.7	0.49	239.8	3072.2
5	Akhlamad, Khorasan-e-Razavi	AK	E 58°59'	N 36°11'	1155	6.7	21.8	0.3	0.7	0.49	239.8	3072.2
6	Buzhan, Khorasan-e-Razavi	BU	E 59°03'	N 36°61'	1600	6.7	21.8	0.3	0.7	0.49	239.8	3072.2
7	Golmakan, Khorasan-e-Razavi	GLM	E 59°13'	N 36°29'	1315	6.6	20.2	0.35	0.69	0.48	212.6	2898.2
8	Azghad, Khorasan-e-Razavi	AZ	E 59°24'	N 36°19'	1800	7.1	21.1	0.37	0.74	0.55	255.2	2892.4
9	Goojgi, Khorasan-e-Razavi	GO	E 59°56'	N 36°31'	2100	7.1	21.1	0.37	0.74	0.55	255.2	2892.4
10	Ortokand, Khorasan-e-Razavi	O	E 59°51'	N 36°48'	1480	7.1	21.1	0.45	0.75	0.55	255.2	2892.4
11	Aman Abad, Khorasan-e-Razavi	AM	E 59°32'	N 35°58'	1210	7.1	21.1	0.37	0.74	0.55	255.2	2894.4
12	Tangehgo, Golestan	T	E 55°49'	N 37°23'	220	11.7	23.9	0.55	0.82	0.68	564.1	2439.1
13	Siahbisheh, Mazandaran	SI	E 51°33'	N 36°23'	1990	6.3	14.8	0.47	0.8	0.63	503.4	1959.4
14	Mohammadshahr, Tehran	MO	E 50°32'	N 35°48'	1140	7.8	21.2	0.32	0.69	0.47	243.8	2959.7
15	Zanjan, Zanjan	ZN	E 48°45'	N 36°30'	1640	4	18	0.37	0.75	0.54	313.1	2843.2
16	Sardabeh, Ardabil	SAR	E 48°15'	N 38°37'	1840	2.8	15.3	0.53	0.89	0.71	303.9	2454.3
17	Meshkinshahr, Ardebil	ME	E 47°38'	N 38°24'	1394	5.9	15.4	0.45	0.75	0.6	383.9	2503.2
18	Sati, Ardebil	SA	E 47°24'	N 38°15'	1920	5.9	15.4	0.45	0.75	0.6	383.9	2503.2
19	Mishoodagh, East Azarbaijan	MI	E 45°38'	N 38°19'	2450	6.9	18	0.37	0.71	0.54	288.9	2794.3
20	Ghasemloo, West Azarbaijan	GH	E 44°43'	N 37°29'	1340	5.4	17.6	0.42	0.78	0.6	341	2829.3
21	Piranshahr, West Azarbaijan	PI	E 45°04'	N 36°41'	1842	6.2	17.9	0.37	0.71	0.51	672.7	2766.4
22	Nenor, Kordestan	N	E 46°00'	N 35°52'	1830	8.7	18.6	0.34	0.58	0.44	689.3	2884.6
23	Zaribar, Kordestan	ZR	E 46°08'	N 35°32'	1285	5	20.6	0.34	0.77	0.53	991.2	2967.9
24	Eberoo, Hamedan	E	E 48°28'	N 34°41'	2250	3.3	19.1	0.36	0.77	0.54	316.6	2929.1
25	Firooz Abad, Fars	F	E 52°37'	N 28°48'	1600	10.1	26.7	0.36	0.65	0.49	416.6	3358.6

T<sub>min</sub>: Average of minimum temperature in year (C°); T<sub>max</sub>: Average of maximum temperature in year (C°); H<sub>min</sub>: Average of minimum relative humidity in year (%); H<sub>max</sub>: Average of maximum relative humidity in year (%); H<sub>total</sub>: Total relative humidity in year (%); P<sub>annual</sub>: Total of precipitation in year (mm); Sun: total of sunshine hours

### 2.2. Evaluation of morphological and agronomical characters

In this study a total of four qualitative and 32 quantitative morphological traits were assessed (Table 2). Of each population, six plants were selected and

their characters were measured using an appropriate instrument. To measure the characters, five random samples were evaluated from each plant.



In order to evaluate morphological diversity and to establish relationships among studied populations, several statistical procedures were conducted. Quantitative data were computed using the SAS software ver. 9.1 (SAS, 2003) to perform analysis of variance, comparison of mean and to calculated coefficient of variation (CV). Simple correlations, factor and cluster analysis and scatter plots were carried out using SPSS® ver. 11.0. Factor analysis was done by Varimax factor rotation technique. Cluster analysis was also done using SPSS® ver. 11.0 based on the matrix resulted from the Euclidian distances and the Wards method.

### 2.3. Data analysis

Table 2. traits, range of variability, mean and coefficient of variations for qualitative and quantitative traits from 25 populations of *A. biebersteinii*

No.	Trait	Abbreviation	Unit	Min	Max	Mean	CV (%)
1	Shoot diameter (between 1 <sup>st</sup> and 2 <sup>nd</sup> nodes)	SD(1&2)	mm	2.18	4.36	3.21	18.9
2	Shoot diameter (near inflorescence)	SD(NI)	mm	1	2.88	2.10	22.1
3	Number of leaves	NL	-	16.08	37.17	28.26	18.9
4	Leaf length	LL	mm	27.15	63.9	50.26	21.5
5	Leaf width	LW	mm	3.81	10.94	7.21	30.5
6	Leaf length/width ratio	LL/LW	Ratio	4.76	12.8	7.32	24.9
7	Internodes distance	ID	mm	14.53	30.71	22.22	21.0
8	Plant height	PH	cm	22.67	62.54	44.53	25.1
9	Number of inflorescence man rays	NIMR	-	5.25	14	7.60	26.7
10	Number of inflorescence heads (Capitula)	NC	-	77.43	265	177.07	26.4
11	Peduncle length	PL	mm	5.81	33.24	17.36	42.6
12	Length of head peduncle	LP	mm	2.05	10.16	3.78	49.2
13	Inflorescence length	IL	mm	26.79	97.09	61.90	37.6
14	Inner bract length	IBL	mm	2.67	3.57	3.21	7.3
15	Outer bract length	OBL	mm	1.41	2.13	1.75	12.1
16	Inner bract width	IBW	mm	0.58	2.67	1.02	44.3
17	Outer bract width	OBW	mm	0.9	1.77	1.31	14.1
18	Inflorescence width	IW	mm	32.85	69.56	51.49	20.5
19	Capitulum width	CW	mm	2.07	3.72	2.52	17.0
20	Capitulum length	CL	mm	3.283	57.585	6.07	176.9
21	Number of ray florets per capitulum	NRPC	-	4.38	5.46	4.92	5.4
22	Length of bract (outside at the base of inflorescence)	LB	mm	8.9	28.83	17.55	29.9
23	Peduncle diameter	PeduD	mm	1.24	4.28	2.07	32.5
24	Pedicel diameter	PediD	mm	0.2	1.29	0.45	50.6
25	Distance of inflorescence from the upest leaf	DIUL	mm	10.74	48.54	26.01	38.1
26	Number of inner bracts	NIB	-	4.31	6.65	5.34	9.6
27	Number of outer bracts	NOB	-	6.67	9.02	7.65	8.3
28	Disc floret length	DFL	mm	2.37	3.74	3.00	10.9
29	Ray floret length	RFL	mm	1.8	2.69	2.26	10.2
30	Number of main stem	NMS	-	1	4.5	2.29	50.1
31	Leaf pubescence	LP	Code	1	3	-	-
32	Stem color	SC	Code	1	4	-	-
33	Flower color	FC	Code	1	3	-	-
34	Leaf color	LC	Code	1	5	-	-

## 3. Results

### 3.1. Analysis of variance

Comparison of means for quantitative parameters showed that all parameters except for number of

leaves, peduncle length, length of head peduncle, inner bract width, inflorescence width, length of bract (outside at the base of inflorescence), peduncle diameter and pedicel diameter were significant ( $P \leq$

0.01) (Table 3). The highest value for leaf length, leaf width and leaf length/width ratio were observed in the Akhlamad (63.9 mm), Ortokand (10.9 mm) and Havar (13.7 mm) populations, respectively and the minimum observed in populations of Tangehgol (27.2 mm), Mishoodagh (3.8 mm) and Zanjan (5.4 mm), respectively. The longest values for the plant height were determined in the population of Goojgi and the shortest in the population of Havar with 62.5 and 22.7 cm, respectively. The mean of highest values for the number of inflorescence main rays and number of inflorescence heads (capitula) was observed in the populations of Meshkinshahr (14 main rays) and Zaribar (number of 265 capitula) and lowest values for them were observed in the Mohammadshahr (6 main rays) and Tangehgol (77 capitula), respectively. The highest value for the inflorescence length was

observed in the population of Eberoo and the lowest in Mishoodagh being 79.1 and 26.8 mm, respectively. The means of highest inner bract width and outer bract width for each plant were in Zanjan population at about 3.57 mm and 2.15 mm, respectively, and the lowest of these traits in populations of Firuz Abad (2.67 mm) and Golmakan (1.41 mm), respectively. The highest internodes distance was detected in population of Ortokand at about 3.71 mm and lowest percentage in Tangehgol population with 14.53 mm. Mean values of the studied morphological and agronomical parameters showed large variations among the populations for almost of measured parameters. Mean values and range of the variability for the different characters of each population are shown in Table 2.

Table 3. Means comparison of 32 quantitative traits in 25 populations of *A. Bieberstenii*

No.	population	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15
		SD(1&2)	SD(NI)	NL	LL	LW	LL/LW	ID	PH	NIMR	NC	PL	LP	IL	IBL	OBL
1	H	3.00	1.82	22.68	59.00	4.61	13.71	23.43	59.32	7.42	132.47	8.63	8.66	54.95	3.34	2.07
2	GOL	2.70	1.75	34.50	39.74	4.34	9.27	20.26	31.80	5.67	143.32	5.81	2.05	39.50	3.07	1.58
3	CH	3.04	2.19	31.00	59.05	6.88	8.65	29.73	46.79	7.33	181.50	20.82	2.68	90.69	3.22	1.50
4	AD	2.83	2.45	20.77	48.98	9.36	5.74	20.91	39.85	6.83	139.67	12.20	3.15	38.98	3.21	1.76
5	AK	3.45	2.11	29.08	63.90	8.56	7.75	22.47	45.85	6.33	186.67	16.80	3.55	53.18	3.28	1.90
6	BU	3.32	2.35	28.83	47.48	9.98	5.82	17.45	39.93	10.00	182.67	16.15	2.61	96.93	3.27	1.56
7	GLM	3.82	2.88	27.67	42.81	6.24	7.09	26.91	59.95	8.67	192.25	28.70	10.16	78.03	3.40	1.41
8	AZ	4.36	2.50	35.83	54.79	7.78	7.68	26.73	62.50	7.92	178.77	21.48	3.83	46.97	3.57	1.84
9	GO	3.54	2.41	37.17	57.08	9.95	5.82	30.25	62.54	7.17	152.88	27.09	4.81	57.52	3.31	2.02
10	O	4.18	2.83	32.33	61.03	10.94	5.76	30.71	49.90	9.17	236.17	24.30	3.45	97.03	3.51	1.92
11	AM	3.32	2.38	25.08	61.17	6.23	9.99	23.47	39.59	8.83	170.65	23.67	3.78	76.24	3.09	1.98
12	T	3.79	2.11	23.67	27.15	4.60	6.33	14.53	27.42	8.17	77.43	10.90	2.91	27.69	3.39	1.69
13	SI	2.73	2.02	19.40	44.23	6.74	8.73	22.96	44.10	6.00	225.92	15.80	3.64	75.97	3.16	1.83
14	MO	2.26	1.47	31.36	55.63	9.65	5.83	23.93	54.00	5.25	99.08	8.61	2.16	36.57	2.79	1.47
15	ZN	3.78	2.24	26.43	49.19	9.15	5.45	19.93	48.85	6.00	222.25	22.67	2.91	85.85	3.57	2.13
16	SAR	2.50	1.45	30.50	29.84	4.13	7.37	15.47	25.77	7.21	179.55	9.69	2.19	42.06	3.31	1.85
17	ME	3.38	2.11	30.33	44.75	6.28	7.27	24.46	41.37	14.00	156.67	18.46	3.78	41.10	3.13	1.65
18	SA	2.64	1.97	16.08	46.63	6.83	7.50	22.04	43.25	5.92	167.85	25.76	4.87	82.08	3.45	1.76
19	MI	2.18	1.40	24.58	29.88	3.82	7.94	16.74	32.28	6.17	93.50	5.92	2.65	26.79	3.15	1.70
20	GH	2.76	1.71	24.79	43.70	4.30	10.58	15.96	36.60	8.80	187.39	13.25	2.82	59.40	2.81	1.58
21	PI	3.32	2.09	34.50	62.14	10.75	6.25	23.27	52.67	6.47	224.37	13.16	3.81	60.70	3.27	2.12
22	N	3.28	2.10	29.33	48.90	5.83	9.08	26.53	49.52	5.83	248.00	18.66	4.07	74.21	3.06	1.73
23	ZR	4.05	2.40	33.94	62.77	8.25	8.07	22.12	53.17	7.00	265.00	20.34	3.01	79.23	3.31	1.62
24	E	3.71	2.80	25.37	54.46	7.23	8.89	19.54	43.51	11.67	184.82	33.24	4.00	97.10	2.92	1.63
25	F	2.40	1.00	31.17	62.27	7.83	8.00	15.68	22.67	6.25	197.83	12.00	2.96	28.75	2.67	1.44
MSD		2.05	1.48	29.05	33.22	7.30	5.71	13.86	24.26	7.97	175.51	34.25	11.46	87.09	0.71	0.76

MSD: Minimum Significant Difference, If difference between two means to be under MSD, are not significant different at 1% level of probability using Tukey's Studentized Range (HSD) Test

### 3.2. Simple correlations

The bivariate correlations between studied characters are shown in Table 4. The most significant positive correlation was found between the capitulum width and inner bract width ( $r=+0.85$ ), shoot diameter (between 1st and 2nd nodes) and shoot diameter (near inflorescence) ( $r=+0.82$ ), plant height and internodes distance ( $r=+0.78$ ), disc floret length and ray floret length ( $r=+0.75$ ), inflorescence width and capitulum width ( $r=+0.70$ ). Also some vegetative characters were significantly correlated with reproductive ones that are observable in correlation Table 4.

### 3.3. Factor analysis

The aim of factor analysis is determining the number of main factors for reducing the number of effective parameters to discriminate populations. For each factor, a factor loading of more than 0.63 was considered as being significant (Table 5). According to factor analysis, 18 of the morphological characters accounted for 44.4% of the variance as the four first main factors, and the other parameters scattered within six factors determined 85.4% of the total variance. The largest portion of the variance at the first factor belongs to variables, shoot diameter (between 1st and

2nd nodes), shoot diameter (near inflorescence), peduncle length, inflorescence length, length of bract (outside at the base of inflorescence) , peduncle diameter and distance of inflorescence from the uppest leaf that negative effects indicated them 14.6% of the total variance. The second factor with 11.2% of total variance included significant positive parameters of the inner bract length, inner bract width, outer bract

width and number of ray florets per capitulum. Characters such as internodes distance, plant height, number of inner bracts and number of outer bracts were existed in the third factor contributing to 10.2% of the overall variance. The fourth factor with 8.5% of the total variance included three parameters of number of inflorescence heads (capitula), inflorescence width and leaf color.

Table 3 (cont.) Means comparison of 32 quantitative traits in 25 populations of *A. bieberstenii*

No.	population	16	17	18	19	20	21	22	23	24	25	26	27	28	29	30
		IBW	OBW	IW	CW	CL	NRPC	LB	PeduD	PediD	DIUL	NIB	NOB	DFL	RFL	NMS
1	H	1.00	1.35	46.32	2.64	4.22	4.78	12.57	1.77	0.31	13.47	6.22	8.60	3.34	2.41	2.50
2	GOL	1.00	1.49	44.25	2.11	3.28	4.58	13.65	1.24	0.31	17.36	5.27	7.38	2.51	1.98	3.50
3	CH	0.58	1.23	36.24	2.09	3.97	4.89	21.46	1.75	0.21	37.37	5.08	7.58	2.71	1.99	1.33
4	AD	0.81	1.26	47.80	2.48	4.04	4.93	15.84	1.63	0.39	30.88	5.00	6.95	3.20	2.62	2.83
5	AK	0.70	1.41	53.13	2.63	4.11	4.52	19.39	1.78	0.44	27.64	5.00	7.36	3.74	2.64	2.00
6	BU	0.71	1.16	43.65	2.22	3.86	5.33	23.27	3.38	0.32	34.37	5.19	7.50	3.23	2.34	1.17
7	GLM	1.14	1.35	50.57	2.47	4.04	5.31	18.53	2.28	0.38	26.87	4.92	8.15	3.23	2.36	1.00
8	AZ	1.17	1.55	61.06	2.80	4.44	4.94	22.83	2.14	0.43	22.34	5.91	8.39	3.11	2.61	1.17
9	GO	1.03	1.34	56.30	2.70	3.99	4.89	19.00	2.08	0.40	48.54	5.80	9.02	3.34	2.20	3.67
10	O	2.67	1.53	59.99	3.72	3.95	5.46	27.97	2.50	0.41	29.57	5.66	8.59	3.17	2.28	1.00
11	AM	1.01	1.50	49.29	2.64	3.60	4.99	19.68	4.28	0.37	32.59	5.22	7.63	3.27	2.31	2.50
12	T	1.07	1.42	39.85	2.15	3.64	5.06	17.19	2.40	0.32	10.74	4.67	7.76	2.87	2.07	2.00
13	SI	0.68	1.05	48.53	2.28	3.65	4.38	15.30	1.82	0.31	30.87	5.59	7.29	2.48	2.04	1.17
14	MO	0.59	0.90	39.56	2.33	3.63	4.87	14.78	1.28	0.20	19.67	6.65	8.25	2.37	1.81	1.33
15	ZN	2.06	1.77	68.00	3.54	5.33	5.22	18.31	2.55	0.54	21.74	5.71	8.21	3.14	2.49	4.50
16	SAR	1.20	1.21	57.82	2.58	4.04	5.00	12.66	1.54	0.40	14.68	5.44	8.10	3.09	2.26	1.67
17	ME	0.84	1.22	48.68	2.21	4.06	5.33	12.00	1.90	1.29	33.24	4.31	6.67	2.84	2.11	2.67
18	SA	0.74	1.19	42.56	2.08	4.15	4.78	16.70	1.91	0.51	31.19	4.89	7.39	2.90	2.31	4.50
19	MI	0.76	1.27	32.85	2.19	3.83	4.64	8.90	1.32	0.45	13.50	5.00	6.67	2.75	2.30	1.00
20	GH	0.91	1.12	49.50	2.30	3.34	4.80	12.17	1.68	0.87	18.35	5.78	7.00	2.68	2.09	3.00
21	PI	1.21	1.52	69.56	3.11	3.64	5.00	24.93	2.03	0.47	18.56	5.77	7.98	3.26	2.35	4.17
22	N	0.81	1.25	43.27	2.07	3.50	4.98	9.68	2.19	0.33	46.05	5.09	7.13	2.76	1.96	2.00
23	ZR	1.02	1.37	66.98	2.49	3.97	4.76	18.00	2.73	0.49	24.64	5.30	7.36	2.69	2.16	3.33
24	E	0.86	1.19	62.24	2.77	3.95	4.71	28.84	2.20	0.81	30.14	4.99	7.33	3.06	2.69	2.33
25	F	0.95	1.18	69.17	2.44	3.30	5.00	15.20	1.50	0.43	15.83	5.00	6.92	3.24	2.22	1.00
MSD		2.24	2.24	63.53	1.48	1.32	1.12	25.25	3.15	1.29	34.27	2.02	2.42	1.11	0.81	3.51

MSD: Minimum Significant Difference, If difference between two means to be under MSD, are not significant different at 1% level of probability using Tukey's Studentized Range (HSD) Test

Table 4. Correlation matrix among 36 morphological characteristics

		1	2	3	4	5	6	7	8	9	10	11	12
		SD(1&2)	SD(NI)	NL	LL	LW	LL/LW	ID	PH	NIMR	NC	PL	LP
1	SD(1&2)	1											
2	SD(NI)	<b>0.82**</b>	1										
3	NL	0.33	0.06	1									
4	LL	0.30	0.26	0.34	1								
5	LW	0.36	0.40	0.34	<b>0.64**</b>	1							
6	LL/LW	-0.23	-0.30	-0.14	0.11	<b>-0.66**</b>	1						
7	ID	<b>0.42*</b>	<b>0.54**</b>	0.36	<b>0.53**</b>	<b>0.42*</b>	-0.05	1					
8	PH	<b>0.52**</b>	<b>0.56**</b>	0.27	<b>0.52**</b>	<b>0.45**</b>	-0.05	<b>0.78**</b>	1				
9	NIMR	<b>0.40*</b>	<b>0.44*</b>	0.03	-0.03	-0.03	0.00	0.04	-0.02	1			
10	NC	<b>0.40*</b>	0.33	0.23	<b>0.46*</b>	0.36	-0.07	0.29	0.26	-0.01	1		
11	PL	<b>0.64**</b>	<b>0.76**</b>	0.02	0.36	0.35	-0.21	<b>0.51**</b>	<b>0.47*</b>	<b>0.40*</b>	<b>0.45*</b>	1	
12	LP	0.26	0.36	-0.20	0.13	-0.11	0.29	0.37	<b>0.55**</b>	0.14	0.04	0.36	1
13	IL	0.45*	<b>0.65**</b>	-0.09	0.37	0.35	-0.10	<b>0.40*</b>	0.38	0.23	<b>0.64**</b>	0.72	0.20
14	IBL	<b>0.60**</b>	<b>0.53**</b>	0.00	-0.07	0.19	-0.29	0.32	<b>0.42*</b>	-0.01	0.14	0.30	0.29
15	OBL	0.27	0.18	-0.03	0.22	0.23	0.00	0.20	0.31	-0.14	0.16	0.08	0.10
16	IBW	<b>0.51**</b>	0.35	0.23	0.11	0.31	-0.23	0.21	0.15	0.08	0.35	0.25	0.04
17	OBW	<b>0.62**</b>	0.39	0.27	0.16	0.15	-0.03	0.18	0.20	-0.04	0.22	0.20	0.10
18	IW	<b>0.45*</b>	0.20	<b>0.40*</b>	<b>0.47*</b>	<b>0.46*</b>	-0.19	0.01	0.17	0.07	<b>0.62**</b>	0.33	0.01
19	CW	<b>0.52**</b>	<b>0.40*</b>	0.26	<b>0.42*</b>	<b>0.56**</b>	-0.26	0.27	0.35	0.06	0.39	0.33	0.07
20	CL	<b>0.43*</b>	0.37	-0.08	0.08	0.26	-0.25	0.19	<b>0.42*</b>	0.06	0.12	0.38	0.21



21	NRPC	<b>0.43*</b>	0.35	0.24	0.01	0.33	-0.37	0.18	0.09	<b>0.48*</b>	0.14	0.27	0.15
22	LB	<b>0.63**</b>	<b>0.68**</b>	0.25	<b>0.50*</b>	<b>0.65**</b>	-0.39	0.33	0.33	0.31	0.32	<b>0.61**</b>	0.02
23	PeduD	<b>0.56**</b>	<b>0.56**</b>	-0.02	0.26	0.25	-0.09	0.12	0.15	0.36	0.31	<b>0.50**</b>	0.10
24	PediD	0.14	0.08	-0.05	-0.08	-0.12	0.06	-0.14	-0.11	0.71	0.09	0.24	-0.02
25	DIUL	0.26	<b>0.52**</b>	0.10	0.33	0.38	-0.22	<b>0.64**</b>	<b>0.40*</b>	0.20	0.36	<b>0.63**</b>	0.10
26	NIB	-0.07	-0.14	0.25	0.32	0.28	0.09	0.22	<b>0.49*</b>	<b>-0.40*</b>	0.03	-0.19	0.02
27	NOB	<b>0.41*</b>	0.32	0.36	0.25	0.35	-0.13	<b>0.48*</b>	<b>0.61**</b>	-0.13	0.01	0.24	0.35
28	DFL	0.36	0.30	0.08	0.38	0.34	-0.06	0.09	0.18	0.16	0.12	0.29	0.37
29	RFL	0.38	<b>0.42*</b>	-0.17	0.22	0.24	-0.11	-0.07	0.17	0.20	0.10	0.35	0.26
30	NMS	0.07	0.07	-0.07	0.11	0.11	0.01	-0.04	0.13	-0.13	0.12	0.14	-0.02
31	LP	-0.14	-0.22	0.31	-0.08	-0.08	0.02	-0.05	-0.20	-0.34	-0.18	-0.23	-0.05
32	SC	0.10	0.07	0.01	-0.03	-0.17	0.27	0.10	0.07	0.14	0.28	0.10	-0.11
33	FC	-0.02	-0.20	-0.06	0.16	-0.11	0.26	-0.20	-0.09	-0.06	0.21	-0.19	-0.32
34	LC	0.27	0.24	0.24	0.23	0.27	-0.13	0.33	0.24	0.01	<b>0.65**</b>	<b>0.56**</b>	<b>0.44*</b>
		1	2	3	4	5	6	7	8	9	10	11	12
		SD(1&2)	SD(NI)	NL	LL	LW	LL/LW	ID	PH	NIMR	NC	PL	LP

\*\*Correlation is significant at the 0.01 level (2-tailed) \* Correlation is significant at the 0.05 level (1-tailed)

Table 4. (cont.) Correlation matrix among 36 morphological characteristics

		13	14	15	16	17	18	19	20	21	22	23	24
		IL	IBL	OBL	IBW	OBW	IW	CW	CL	NRPC	LB	PeduD	PediD
13	IL	1											
14	IBL	0.29	1										
15	OBL	0.10	<b>0.53**</b>	1									
16	IBW	0.27	<b>0.50*</b>	<b>0.45*</b>	1								
17	OBW	0.13	<b>0.64**</b>	<b>0.59**</b>	<b>0.66**</b>	1							
18	IW	0.18	0.13	0.35	<b>0.52**</b>	<b>0.43*</b>	1						
19	CW	0.31	<b>0.42*</b>	<b>0.63**</b>	<b>0.85**</b>	<b>0.63**</b>	<b>0.70**</b>	1					
20	CL	0.28	<b>0.71**</b>	<b>0.46*</b>	<b>0.40*</b>	<b>0.48*</b>	0.27	<b>0.52**</b>	1				
21	NRPC	0.22	0.30	-0.02	<b>0.55**</b>	0.27	0.20	0.38	0.25	1			
22	LB	<b>0.60**</b>	0.31	0.17	0.38	0.33	0.45*	<b>0.58**</b>	0.22	0.28	1		
23	PeduD	<b>0.56**</b>	0.29	0.24	0.25	0.37	0.18	0.26	0.15	0.43	<b>0.45*</b>	1	
24	PediD	-0.02	-0.16	-0.04	0.03	-0.05	0.24	0.04	0.11	0.17	-0.06	-0.03	1
25	DIUL	<b>0.54**</b>	0.06	0.04	-0.13	-0.11	-0.09	-0.07	0.06	0.13	0.22	0.33	0.05
26	NIB	0.01	0.00	0.34	0.21	-0.01	0.17	0.39	0.10	-0.14	0.07	-0.13	-0.37
27	NOB	0.19	<b>0.52**</b>	<b>0.47*</b>	<b>0.50**</b>	0.36	0.26	0.57	0.38	0.27	0.39	0.16	<b>-0.40*</b>
28	DFL	0.10	0.35	<b>0.48*</b>	0.26	<b>0.45*</b>	<b>0.42*</b>	<b>0.49*</b>	0.36	0.28	<b>0.40*</b>	0.33	0.00
29	RFL	0.16	0.39	<b>0.40*</b>	0.21	<b>0.42*</b>	<b>0.43*</b>	<b>0.49*</b>	<b>0.55**</b>	0.04	<b>0.45*</b>	0.20	0.17
30	NMS	0.10	0.19	<b>0.45*</b>	0.11	0.37	0.27	0.17	0.26	-0.11	-0.01	0.06	0.28
31	LP	<b>-0.43*</b>	0.00	-0.15	0.07	0.23	-0.18	-0.36	-0.27	0.12	-0.09	-0.15	-0.08
32	SC	0.33	0.10	-0.17	0.12	-0.03	0.12	-0.01	0.04	0.01	-0.10	0.08	-0.19
33	FC	0.24	-0.03	0.12	0.10	0.11	0.09	0.18	-0.26	-0.24	0.15	0.02	-0.17
34	LC	<b>0.45*</b>	0.11	0.18	0.37	0.11	<b>0.54**</b>	0.23	-0.02	0.31	0.16	0.46	0.32

Table 4. (cont.) Correlation matrix among 36 morphological characteristics

		25	26	27	28	29	30	31	32	33	34
		DIUL	NIB	NOB	DFL	RFL	NMS	LP	SC	FC	LC
25	DIUL	1									
26	NIB	-0.19	1								
27	NOB	0.03	<b>0.65**</b>	1							
28	DFL	0.10	-0.09	0.31	1						
29	RFL	-0.05	-0.11	0.09	<b>0.75**</b>	1					
30	NMS	0.07	0.04	0.06	0.05	0.10	1				
31	LP	-0.13	-0.28	-0.12	-0.03	-0.20	-0.10	1			
32	SC	0.02	0.23	0.14	-0.15	-0.24	0.07	<b>-0.55**</b>	1		
33	FC	-0.39	0.28	0.21	-0.20	-0.15	0.11	-0.37	0.29	1	
34	LC	<b>0.40*</b>	0.04	0.04	0.14	-0.05	0.22	-0.02	0.26	-0.16	1

\*\*Correlation is significant at the 0.01 level (2-tailed) \* Correlation is significant at the 0.05 level (1-tailed)

Table 5. Eigen values of rotated factors and cumulative variance (%) of 10 factors contributing to 100% of total variance

Factor	1	2	3	4	5	6	7	8	9	10
Cumulative variance (%)	14.59	25.80	35.96	44.43	52.88	59.91	66.70	73.27	79.56	85.36
Eigen value	5.25	4.04	3.66	3.05	3.04	2.53	2.45	2.37	2.26	2.09
Parameters	Factor loading									
SD(1&2)	<b>0.63**</b>	0.50	0.21	0.07	0.19	0.12	-0.06	-0.05	0.25	0.17
SD(NI)	<b>0.84**</b>	0.28	0.19	-0.03	0.16	0.08	-0.05	-0.10	0.17	-0.09
NL	-0.05	0.17	0.48	0.25	-0.14	-0.05	-0.27	-0.10	0.18	<b>0.63**</b>
LL	0.30	-0.27	0.54	0.44	0.38	0.05	-0.07	0.12	-0.04	0.24
LW	0.30	0.08	0.44	0.34	0.24	0.00	<b>-0.65**</b>	-0.05	-0.06	-0.08
LL/LW	-0.17	-0.30	0.02	-0.04	0.05	0.06	<b>0.81**</b>	0.22	0.03	0.26
ID	0.48	0.07	<b>0.67**</b>	0.13	-0.10	-0.01	0.09	-0.35	-0.11	0.08
PH	0.38	0.10	<b>0.80**</b>	0.03	0.09	0.19	0.11	-0.22	-0.05	-0.14
NIMR	0.37	0.08	-0.13	-0.06	0.08	-0.19	0.07	-0.06	<b>0.84**</b>	-0.03
NC	0.37	0.07	0.03	<b>0.81**</b>	0.02	0.10	0.00	0.11	-0.07	-0.03
PL	<b>0.79**</b>	0.07	0.13	0.25	0.16	0.13	0.02	-0.15	0.21	-0.11
LP	0.20	0.14	0.28	0.01	0.31	-0.11	<b>0.65**</b>	-0.31	-0.01	-0.24
IL	<b>0.82**</b>	0.06	0.08	0.31	-0.03	0.04	0.06	0.25	-0.06	-0.22
IBL	0.34	<b>0.72**</b>	0.09	-0.19	0.17	0.26	0.04	-0.18	-0.23	-0.09
OBL	-0.02	0.39	0.22	0.08	0.42	0.51	0.00	0.08	-0.23	-0.14
IBW	0.08	<b>0.81**</b>	0.13	0.32	0.10	0.03	-0.10	0.22	0.05	0.03
OBW	0.18	<b>0.67**</b>	-0.01	0.07	0.31	0.41	0.05	0.04	-0.10	0.41
IW	-0.02	0.26	0.16	<b>0.66**</b>	0.41	0.24	-0.20	0.18	0.23	0.13
CW	0.10	0.58	0.35	0.33	0.43	0.14	-0.22	0.30	0.08	-0.04
CL	0.20	0.52	0.18	-0.11	0.30	0.38	0.02	-0.05	0.06	-0.30
NRPC	0.21	<b>0.63**</b>	-0.01	0.23	-0.02	-0.34	-0.14	-0.24	0.32	-0.01
LB	<b>0.64**</b>	0.16	0.24	0.08	0.36	-0.07	-0.40	0.27	0.10	0.18
PeduD	<b>0.67**</b>	0.25	-0.15	0.17	0.14	-0.01	0.01	0.11	0.00	0.05
PediD	-0.03	-0.07	-0.22	0.15	0.04	0.28	0.03	-0.13	<b>0.85**</b>	-0.13
DIUL	<b>0.65**</b>	-0.25	0.18	0.24	-0.13	0.06	-0.08	-0.44	-0.03	-0.14
NIB	-0.27	0.08	<b>0.77**</b>	0.06	-0.05	0.07	-0.04	0.38	-0.21	-0.12
NOB	0.11	0.53	<b>0.71**</b>	-0.04	0.10	0.01	0.04	0.09	-0.18	-0.01
DFL	0.15	0.23	0.06	0.12	<b>0.86**</b>	-0.03	0.03	-0.14	0.00	0.05
RFL	0.20	0.15	-0.06	-0.08	<b>0.88**</b>	0.19	-0.05	-0.01	0.13	-0.10
NMS	-0.01	0.04	-0.03	0.15	-0.01	<b>0.90**</b>	-0.02	-0.01	0.02	-0.03
LP	-0.19	0.05	-0.25	-0.07	-0.06	-0.08	-0.29	-0.52	-0.33	0.56
SC	0.16	0.15	0.11	0.30	-0.35	-0.01	0.60	0.31	0.11	-0.02
FC	0.02	0.02	-0.04	0.04	-0.12	0.00	0.11	<b>0.87**</b>	-0.22	0.03
LC	0.27	0.12	0.05	<b>0.86**</b>	-0.09	0.18	0.06	-0.18	0.09	-0.05
Factor	1	2	3	4	5	6	7	8	9	10

\*\*Significant factor loadings (considered values above 0.63)

### 3.4. Cluster analysis

In order to reveal relationships among the populations of *A. biebersteinii*, a cluster analysis was performed based on 10 factors in this study. All populations at approximately a distance of 10 out of 25 were grouped into 10 main branches (Fig 2).

Group A: This branch was divided into two sub-clusters consisting of five populations, Adag, Akhlamad, Piranshahr, Azghad and Goojgi. These populations had similarity in some characters such as number of leaves and outer bract width. Members of

the first group had the highest value for these parameters than other populations.

Group B: This cluster contained only one member namely Mohammadshahr. This population has highest values for leaf length/width ratio and number of inner bracts. Also, population of Mohammadshahr has lowest value for number of inflorescence main rays, pedicel diameter, disc floret length and ray floret length among others.

Group C: This branch only include one member namely Meshkinshahr population. This population has highest value for number of inflorescence main rays

and pedicel diameter. Length of head peduncle and number of inner bracts were lowest in this population.

Group D: This branch was consisted of population of Eberoo only. This population has utmost value for peduncle length, inflorescence length, length of bract (outside at the base of inflorescence) and ray floret length, which caused the population to be classified into separate cluster.

Group E: This branch was divided into two sub-clusters including three populations of Buzhan, Ortokand and Golmakan. These populations had a similar shoot diameter (near inflorescence), internodes distance, inner bract length, inner bract width, number of ray florets per capitulum and peduncle diameter in comparison to the other populations.

Group F: This cluster only consist of one member namely Firooz Abad. This population has the highest value for leaf length. The lowest values for shoot diameter (near inflorescence), plant height, inner bract length, outer bract length, capitulum length and number of main was also observed for this population in this study.

Group G: This cluster comprises two populations including Tangehghol and Mishoodagh. Members of this group contained leaves with high degree of pubescence. Also, this group has the lowest value of leaf length, number of inflorescence heads (capitula), peduncle length and inflorescence length.

Group H: This branch divided into two sub-clusters, in which the first sub-cluster consisted of two populations (Sardabe and Ghasemloo), while second sub-cluster was made of Zanzan population. These plants were similar in terms of capitulum width, capitulum length and number of inner bracts.

Group I: Only includes population from Havar. This population was among the populations whit the highest leaf length/width ratios, plant height, outer bract length, number of outer bracts and disc floret length.

Group J: This branch was divided into two sub-clusters including three populations of Sati, Nenor and Siahbishe. They were similar from the points of number of inflorescence heads and distance of inflorescence from the upest leaf.

Group K: This cluster divided into two sub-clusters, the first sub-cluster consisted of three populations of Aman Abad, Zaribar and Chelmir, and the Second sub-cluster included population of Golool. The common features of these plants were some qualitative parameters such as degree of leaf pubescence, stem color and flower color.

#### 4. Discussion

It has widely been accepted that there is a correlation between environmental conditions and morphological variation of a certain plant species, and

that species that can occupy a wide range of habitats are more variable in morphology than species with a narrow range of habitats (Baker, 1974; Sultan, 2001; Richards and Pennings, 2005). Morphological variation also is representative of the adaptability of populations to different habitat; species that has a greater morphological variation would be more adaptive to environment than species with a small morphological variation (Pang and Jiang, 1995).

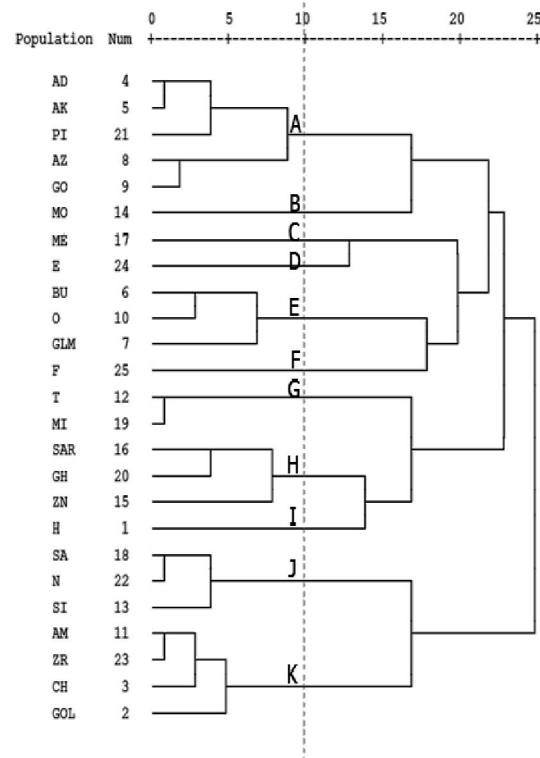


Figure 2. Dendrogram of the similarities among the 25 Iranian wild populations of *A. biebersteinii*, using Wards method based on 10 factors

*Achillea* species have a wide range of distribution in Iran. They differ widely in morphology, phenology, flowering and fruiting patterns (Mozaffarian, 2008). Some of which have significant morphological differentiation within them. It has been shown that there is a close relationship between the leaf area and there essential oil content; this parameters was reported to vary in a great degree among different populations of some *Achillea* species growing wild in different port of Iran (Rahimmalek et al., 2009).

Results of present study showed that there is a high morphological variation within populations of *A. biebersteini* collected from different parts of Iran. As shown in Fig 2 populations of Adag and Akhlamad, Tangehghol and Mishoodagh, Sati and Nenor, Aman



Abad and Zaribar were the most similar than other studied populations and, in country, populations of Adag and Golool have the least similarity. The population of Firooz Abad has the lowest similarity with others, which has different mean value for many characteristics than other populations such as the highest value for leaf length, also it has lowest value for shoot diameter (near inflorescence), plant height, inner bract length, outer bract length, capitulum length and number of main stem.

In conclusion, morphological diversity is the observable physical variation present in populations and includes both genotypic and environmental components (Schlichting and Levin, 1984; Tulig and Clark, 2000; Yeater et al., 2004). Genotypic variability is the component of variation that is due to the genotypic differences among individuals within a population or among populations within a species (Humphreys, 1991; Loos, 1993). Although morphology cannot be directly related to genotype, it has a strong genotypic basis. Therefore, morphological characters can be used as a measure of genetic variations between populations (Schlichting, 2002). In total, we observed in present study that the studied populations of *A. biebersteinii* are diverse and variations between them are high. Therefore, selection of suitable traits for use in breeding programs of this plant is possible.

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#### References

- Bader A, Flamini G, Cioni P L, Morelli I. Essential oil composition of *Achille santolina* L. and *Achillea biebersteinii* Afan collected in Jordan. *Flavour and Fragrance Journal* 2003; 18: 36–38.
- Baker H G. The evolution of weeds. *Annual Review of Ecology and Systematics* 1974; 5: 1–24.
- Baris O, Gulluce M, Sahin F, Ozer H, Kilic H, Ozkan H, Sokmen M, Ozbek T. Biological activities of the essential oil and methanol extract of *Achillea biebersteinii* Afan. (Asteraceae). *Turkish Journal of Biology* 2006; 30: 65–73.
- Benedek B, Kopp B, Melizg M F. *Achillea millefolium* L. s.l. Is the anti-inflammatory activity mediated by proteaseinhibition? *Journal of Ethnopharmacology* 2007; 113: 312–317.
- Chevallier A. *The Encyclopedia of Medicinal Plants*. Dorling Kindersley Publishing Inc, London, 1996.
- Esmaili A, Nematollahi F, Rustaiyen A, Moazami N, Masoudi S, Bamasian S. Volatile constituents of *Achillea pachycephala*, *A. oxyodonta* and *A. biebersteinii* from Iran. *Flavour and Fragrance Journal* 2006; 21: 2353–3256.
- Ge S, Hong D Y. Biosystematic studies on *Adenophora potaninii* Korsh. Complex (Campanulaceae) III. Genetic variation and taxonomic value of morphological characters. *Acta Phytotaxonomica Sinica* 1995; 33 (5): 433–443.
- HuberMorath A. *Achillea*. In: Rechinger K.H. (eds), *Flora Iranica compositae VI-Anthemideae*, Lfg: 49-71, Ackademiche Druck-U, Graz, 1986.
- Humphreys M O. A genetic approach to the multivariate differentiation of perennial ryegrass (*Lolium perenne* L.) cultivars. *Heredity* 1991; 66: 437–443.
- Karlova K. Essential oil from yarrow (*Achillea millefolium* complex), the content and the composition. *Proceeding of 9th international conference of Horticulture, Lednice, Czech Republic* 2001; 2: 339-342.
- Kordali S, Cakir A, Akcin T A, Mete E, Akcin A, Aydin T, Kilic H. Antifungal and herbicidal properties of essential oils and n-hexane extracts of *Achillea gypsicola* Hub-Mor. and *Achillea biebersteinii* Afan. (Asteraceae). *Industrial Crops and Products* 2009; 29: 562-570.
- Lewis M. What is Yarrow-*Achillea millefolium* Essential Oil? [www.Worldwidehealth.com](http://www.Worldwidehealth.com), Alternative Medicine, Complementary Health Directory and Resources, UK, 2006.
- Loos B P. Morphological variation in *Lolium* (Poaceae) as a measure of species relationships. *Plant Systematics and Evolution* 1993; 188: 87–99.
- Mal T K, Doust J L. Phenotypic plasticity in vegetative and reproductive traits in an invasive weed, *Lythrum salicaria* (Lythraceae), in response to soil moisture. *American Journal of Botany* 2005; 92 (5): 819–825.
- Mozaffarian V. *Encyclopedia of Iranian Plants*. Farhang moaser Publication, Tehran, 1996.
- Mozaffarian V. *Compositae: Anthemideae & Echinopeae*. In: *Flora of Iran: 67-105*, Publication of Research Institute of Forest and Rangelands, Tehran, 2008.

17. Pang G C, Jiang D M. Population genetic diversity and data analysis. *Scientia Silvae Sinicae* 1995; 31 (6): 543–550.
18. Paulo S. Effect of the essential oil of *Achillea millefolium* L. in the production of hydrogen peroxide and tumor necrosis factor - $\alpha$  in murine macrophages. *British Journal of Pharmacology* 2005; 41(3): 401-405.
19. Rahimmalek M, Sayed Tabatabaei B E, Etemadi N, Hossein Golid S A, Arzania A, Zeinalie H. Essential oil variation among and within six *Achillea* species transferred from different ecological regions in Iran to the field conditions. *Industrial Crops and Products* 2009; 29: 348-355.
20. Richards C L, Pennings S C, Donovan L A. Habitat range and phenotypic variation in salt marsh plants. *Plant Ecology* 2005; 176: 263–273.
21. Rustaiyan A, Komeilizadeh H, Shariatpanahi M S, Jassbi A R, Masoudi S. Comparative study of essential oils of three *Achillea* species from Iran. *Journal of Essential Oil Research* 1998; 10: 207-209.
22. SAS. SAS® 9.1 Qualification Tools User's Guide. SAS Institute Inc., Cary, NC, USA, 2003.
23. Schlichting C D, Levin D A. Phenotypic phytotoxicity of annual phlox test of some hypotheses. *American Journal of Botany* 1984; 71: 252–260.
24. Schlichting C D. Phenotypic plasticity in plants. *Plant Species Biology* 2002; 17: 85–88.
25. Sokmen A, Sokmen M, Daferera D, Polissiou M, Candan F, Unlu M, Akpulat H A. The in vitro antioxidant and antimicrobial activities of the essential oil and methanol extracts of *Achillea biebersteinii* Afan. (Asteraceae). *Phytotherapy Research* 2004; 18: 451–456.
26. Sultan S E. Phenotypic plasticity for fitness components in *Polygonum* species of contrasting ecological breadth. *Ecology* 2001; 82: 328–343.
27. Tulig M C, Clark C. Morphological variation in *Mimulus* section *Diplacus* (Scrophulariaceae). *American Journal of Botany* 2000; 87: (Suppl. 6) 182.
28. Yang, J. Intraspecific variation in plant and the exploring methods. *Journal of Wuhan Botanical Research* 1991; 9 (2): 185–195.
29. Yeater K M, Bollero G A, Bullock D J, Rayburn A L, Zas S R. Assessment of genetic variation in hairy vetch using canonical discriminate analysis. *Crop Science* 2004; 44: 185–189.
30. Zargari A. Medicinal Plants. Tehran University Publication, Tehran, 1996.

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