# Scanning Electron Microscopy of Antennal Sensilla of *Goniocotes* Species infesting Helmeted Guinea fowl, *Numida meleagris* (Galliformes: Numididae)

Aftab Ahmad<sup>1</sup>, Neelima Gupta<sup>1,\*</sup>, Dileep Kumar Gupta<sup>2</sup>, Arun Kumar Saxena<sup>3</sup>

<sup>1</sup> Centre of Excellence, Department of Animal Science, MJP Rohilkhand University Bareilly 243006, UP, India <sup>2</sup> Department of Zoology, Bareilly College, Bareilly 243005, UP, India <sup>3</sup> Department of Zoology, Government Raza Postgraduate College, Rampur 244 901, UP, India

draftab.lifescience@rediffmail.com, guptagrawal@rediffmail.com, guptaby@gmail.com, akscsir@rediffmail.com

Abstract: Phthirapteran antennae are the principal sites of sensory structures. The antenna of ischnoceran Phthiraptera consists of three components viz. basal scape, a pedicel and the flagellum. The flagellum is further sub divided into three segments: first, second and third flagellomere. SEM study revealed that the second and third flagellomere of *Goniocotes* species infesting Helmeted Guinea fowl, *Numida melagris* carry a placodean sensilla and a sensilla coeloconica. The scape, pedicel and I flagellomere carry 1-3 sensilla chaetica and tectile hair. The nature of sensory organ of both the sexes of adult and different instars of nymphs of *Goniocotes* species appear quite similar. The present report thus furnishes information on the nature of antennal sensilla of three instars of nymphs and the adult of an ischnoceran louse *Goniocotes* species infesting Helmeted Guinea fowl.

[Ahmad A, Gupta N, Gupta DK, Saxena AK. Scanning Electron Microscopy of Antennal Sensilla of *Goniocotes* Species infesting Helmeted Guinea fowl, *Numida meleagris* (Galliformes: Numididae). *Researcher* 2014;6(4):7-11]. (ISSN: 1553-9865). http://www.sciencepub.net/researcher. 3

Key words: lice; Phthiraptera; Mallophaga; Ischnocera.

# 1. Introduction

Phthirapteran ectoparasites (lice) are very small arthropodan creatures which spend their life on different mammalian and avian hosts. Many morphological features of these tiny creatures are not visible under simple microscopic study and hence scanning electron microscopic (SEM) study is required for observing the specific details. Antennal sensillum is also one of the special features located on anterior part of the head of the louse.

The external morphology of different kinds of sensilla found on the surface of the head of phthirapteran ectoparasites vary in shape, size and patterns. It is also a useful tool for taxonomic study. Occasionally, outline drawing of the antenna under low magnification are included in systematic papers which provide only superficial impression of sense organ. Essig (1942) furnished a drawing of the antenna of Menacanthus stramineus in his description of the order for the first time. The occurrence of sensilla in five phthirapteran species was noted (Nuffer, 1954). Clay (1970) demonstrated the presence of saucer like sensilla on last antennal segments of three ischnoceran lice with the help of SEM. The sensilla of antennal flagellum of Craspedorhynchus americanus were demonstrated by (Slifer, 1976). Zlotorzyca and Kassner (1986) recorded the antennal sense organ of 14 phthirapteran species and concluded that its morphology can be used not only for systematics but also for recording the physiological properties. Zlotorzycka and Modrzejewska (1992) further studied

the ultra-structure of antenna of an ischnoceran species, Docophoroides brevis. Cichino and Abrahomobich (1988) recorded the antennal sensilla of adult as well as three instars of nymphs of Boroneilla bergi. The morphology of antennal sensilla of sheep louse, Bovicola bovis (Clark, 1990) and scanning electron microscopy of the pit organs of several species of the same genus (Cruz and Mateo, 1996) have been studied from time to time. The tuft organs were studied on antennal flagellum of Pediculus humanus (Steinbrecht, 1994) but those of P. humanus humanus have comparatively received more attention (Miller, 1969; Szczesna, 1978, 1985; Slifer and Sekhon, 1980). Likewise, the three anopluran species viz., Polyplax serrata, Solenopotes capillatus, Echinophthirius horridus have also been studied from this point of view (Miller, 1970a, b; 1971a, b). Qadri (1936), Dethier (1957), Zacharuk (1985), Mclver (1987), Perez et al. (1995) and Cruz (1995) contributed to physiology and nature of blood sucking insects. Smith (2001) made attempts to review the work done on the subject.

Other significant contributors on scanning electron microscopy of antennal sensilla include workers like Clarke (1990) (*Damalinia ovis*); Baker and Chandrapatya (1992) (*Haematomyzus elephantis*); Steinbrecht (1994) (*Pediculus humanus corporis*); Green and Turner (2001) (louse fly); Cruz and Mateo (1996) (*Bovicola*), 1998 (*Damalinia*), 2001(*Damalinia*) and 2009 (*P. humanus* and *Haematopinus apri*); Turner (2003) (*Damalinia crenelata*); Turner (2004) (*Haematopinus bufali*) and Agarwal et al. (2011) (Upupicola upupae) conducted scanning electron microscopy of antennal sensilla. Information on the sensory equipment of the antennal flagellum of several species of *Damalinia* was provided by Cruz and Mateo (1998). The distribution of sensilla on the antennal flagellum of five poultry lice was noted by Bhatnagar et al. (2004). Arya et al. (2010) compared the nature of antennal sensilla of selected *Brueelia* species. Recently, Arya and Singh (2012) described the antennal sensilla of head of poultry shaft louse, *Menopon gallinae*. The present report furnishes information on the nature of antennal sensilla of three nymphal instars and the adult of an ischnoceran louse *Gonicotes* species infesting Helmeted Guinea fowl.

#### 2. Materials and Methods

Adult lice and three instars of nymphs were collected from Helmeted Guinea fowl from District Bareilly, India through survey work. The lice were stored in 70% alcohol, cleaned in distilled water (two changes) and ether (three changes), dehydrated, mounted on metal specimen stub using double sided tape, coated with gold palladium in Neo Coater 100-240V and observed under SEM (Neo JCM-6000). The samples were then observed under SEM at varying magnifications and selected areas were photographed. Some specimens were treated with Osmium tetra oxide (2%) for achieving better results.

# 3. Results

The antennae of female *Goniocotes* species consist of a scape, a pedicel and three terminal flagellomeres (Figure 1A). The scape is a small rod like structure (30  $\mu$ m) whereas, pedicel is elongated and cylindrical (46.5  $\mu$ m). The Ist and IInd flagellomeres resemble pedicel (26  $\mu$ m and 24  $\mu$ m respectively). The terminal flagellomere is comparatively longer (45.1  $\mu$ m). The scape, pedicel and first flagellomere bear 1-3 spines (sensilla chaetica) and tectile hair.

The IInd and IIIrd flagellomere bear a sensilla placodea and sensilla coeloconica. The former is saucer shaped with central raised area surrounded by varying number of radiating ridges, separated by narrow grooves. The apical end of IIIrd flagellomere bears 10-12 sensilla (Figure 1B). One of them appears whip like and tappers to a fine point. The remaining structures appear to be tectile pegs of varying sizes ( $6.01\mu$ m- 21.6  $\mu$ m). Most of the tectile pegs possess broader bases and blunt terminated tips. The presence of aperture at the tips could not be ascertained.

The antennal sensilla of the male *Goniocotes* species are not sexually dimorphic (Figure 1C). The distribution of placodean sensilla and tectile pegs

appears similar (Figure 1D). As far as the nature of antennal sensilla of nymphal instars is concerned the apex of terminal flagellomere of all the three nymphal instars carries 10-12 tectile pegs, as in case of adult (Figure1A-I). Likewise, side wall of IIIrd flagellomere bears sensilla placodea, as found in adults.



- A. Whole antennae of male (X 550)
- B. Enlarged view of III flagellomere (X 2400)
- C. Whole antennae of female (X 700)
- D. Enlarged view of III flagellomere (X 2000)
- E. Whole antenna of first instar (X 1100)
- F. Enlarged view of III flagellomere (X 7500)
- G. Whole antenna of second instar (X 650)
- H. Enlarged view of apical end of IIIrd flagellomere (X 2700)
- I. Whole antenna of third instar (X 800)

Figure 1. A-I. Scanning Electron Micrographs of antennae of *Gonicotes* species

#### Abbreviations

FLI-Flagellomere I; FLII-Flagellomere II; FLIII-Flagellomere III; PED- Pedicel; SCH-Sensilla chaetica; SP-Scape; SLP-Sensilla Placodea; TH-Tectile hair.

## 4. Discussion

The primary function of insect antennae appears to be sensory. The antennae of most ischnoceran lice comprise of 5 annuli (scape, pedicel and three flagellomeres) but in some species (eg: *Coloceras* species), the three flagellomeres fuse to form a single structure. A scrutiny of literature reveals that there is considerable superficial diversity in the form of antennal sense organ, even within genera.

Bhatnager et al. (2004) studied the antennal sensilla of four ischnoceran species (*Lipeurus heterographus*, *L. caponis*, *L. lawrensis tropicalis* and *Goniocotes gallinae*) and one amblyceran species (*Menopon gallinae*) poultry lice by SEM. In this study, the IInd and IIIrd flagellomere of ischnoceran species bear sensilla coeloconica and the pore organ. The apex of terminal flagellomeres of four species carries a cluster of eleven tectile hairs. The IIIrd flagellomere of amblyceran lice bears a diverse assemblage of spines, cones and pits.

Surman and Suneel (2012) described the antennal sensilla of head of poultry shaft louse, *Menopon gallinae*. He observed a small, ovoid scape and pedicel (broad cup-like structure having narrower base) in antennal sensilla under SEM. In addition to sensory setae, sense organ was present on terminal segment. Tuft organ contains 6/7 small peg like structures. Pit organ was also visible at the sub-terminal area of fourth segment. Presence of any structure resembling coeloconic chaemo-receptor was not observed on any flagellar sub-segment of *M. gallinae*.

According to Clay (1970) the scape, pedicel and Ist flagellomere bear tectile sensilla while IIIrd flagellomere carries two plates and pit sensilla. Furthermore, 10-12 tectile sensilla occur at the tips of IIIrd flagellomere. In case of Goniocotes species, a IIIrd flagellomere appears to carry one plate and one pore organ. Clay (1970) further observed two sensilla coeloconica on terminal segment of species having four segmented antenna in a member of Menoponidae. She further stated that Boopidae and Ricinidae also have similar arrangements but Laemobothridae have terminal three sensilla on segments and Trimenoponidae and Gyropydae exhibit diversity in the form of antennal organs. However, Zlotorzycka and Kassner (1986) also reported that the terminal segments of several ischnocerans bear two pore organs and one sensilla coeloconica. In Goniocotes species, presence of any such sensilla coeloconicum was not observed. Miller (1969, 1971a) also recorded the presence of pore organ in certain species. He further noted that anopluran pore organ remains surrounded by a ring of radiating grooves. However, the presence of sensory setae and tuft organ (consisting of thin / or thick walled) chemo-receptors appears to be a common feature of most of phthirapterans investigated so far. The number, size and location of these structures on different components of antenna seem to be variable (Miller 1969, 1970a, b, 1971b; Clay 1969; Clay 1970; Slifer 1976; Szczesna 1978, 1984; Zlotorzycka and Kassner 1986; Kassner and Zlotorzycka 1987; Zlotorzycka and Modrzejewska 1992).

The present study further indicates that the nature and distribution of antennal sensilla was quite similar in the two sexes of *Goniocotes* species. Moreover, the nature and distribution of sensory equipments in I, II and IIIrd instar nymph was also quite similar. Cicchino and Abrohamovich (1988) also noted that the nature of antennal sensilla of different nymphal instars of nymphs and adults of *Vernoniella bergi* are quite similar. Thus, the present paper supports the observation of Cicchino and Abrohamovich (1988) as there is no major difference in the sensory equipments of different nymphal stages and adults of the ischnoceran phthiraptera, *Goniocotes*.

### Acknowledgement:

One of us, Aftab Ahmad (Dr D.S. Kothari Post Doctoral Fellow) expresses thanks to the University Grants Commission, New Delhi for providing funds under Dr. D.S. Kothari Post Doctoral Fellowship Scheme of U.G.C. (Reference: F-4-2/2006 (BSR)/133-398/2011 (BSR).

# **Correspondence to:**

Prof. Neelima Gupta Centre of Excellence Laboratory Department of Animal Science MJP Rohilkhand University Bareilly- 243006, UP, India Telephone: +91-9412376677 Emails: guptagrawal@rediffmail.com; guptaby@gmail.com

### References

- 1. Essig EO. *College Entomology*. 1942; P. 197. The Macmillan Company, New York.
- Neuffer G. Die Mallophaga geshaut andihru Differenxierungen. Zool. J. Anat 1954; 73: 450-519.
- 3. Clay T. The Amblycera (Phthiraptera: Insecta). Bulletin of the British Museum (Natural History). Entomol 1970; 25(3): 73-98.
- 4. Slifer EH. Sense organs on the antennal flagellum of a bird louse (Mallophaga). J. New York Entomol. Soc 1976; 84:159-165.
- 5. Zlotorzycka J, Kassner J. Antennal sensilla in Mallophaga in scanning electron microscope.

Folia Histochem. et Cytobiol 1986; 24(4):324-325.

- Zlotorzycka J, Modrezejewska M. Morphologic features, with particular regard to surface ultra-structure of *Docophoroides brevis* (Docopherididae, Mallophage). Wiad. Parazytol 1992; 38(1-2): 43 – 50.
- Cicchino AC, Abrohamovich AL. Contribution to the knowledge of cephalic sensilla and wateruptake system of adult and nymphs of *Vernoniella bergi* (Kellogg, 1906) (Insecta: Phthiraptera: Ischnocera). Microsc. Electr. Biol. Cell 1988; 12 (2): 121-148.
- Clark AR. External morphology of the antennae of *Damilinia ovis* (Phthiraptera: Trichodectidae). J. Morphol 1990; 203(2): 203-209.
- 9. Cruz MDS, Mateo MPM. Antennal sense organs of Phthiraptera (Insecta) scanning electron microscopy of the 'Pit Organs' of several species of *Bovicola*. Micron 1996; 27(1): 11-15.
- 10. Steinbrecht R A. The tuft organs of the human body louse, Pediculus humanus corporis cryofixation study of a thermo-hygrosensitive sensillum. Tissue and Cell1994; 26(2):259-275.
- 11. Miller FH. Antennal tuft organs of *Pediculus humanus* Linn. and *Phthirus pubis* (Linn.). J. New York Entomol. Soc 1969; 77:85-89.
- Szcsesna Z. Sense organs on the legs of *Pediculus* humanus humanus L. (Anoplura: Pediculidae) nymphs and imagines. Polskie Pismo Entomol 1978; 48(4):593-600.
- 13. Szczesna Z. The sense organs on antennae of *Pediculus humanus* (Anoplura, Pediculidae) in post-embryonic development stages. Acta Parasitol. Polonica 1984; 29(30-43):395-404.
- Slifer E H, Sekhon S S. Sense organs on the antennal flagellum of the human louse, Pediculus humanus (Anoplura). J. Morpho 1980; 164:161-166.
- Miller FH. Scanning electron microscopy of antennal structures of *Polyplax serrata* (Burmeister) (Anoplura: Hoplopleuridae). J. New York Entomol. Soc 1970a; 78(3): 33-37.
- Miller FH. Scanning electron microscopy of Solenopotes capillatus Enderlein (Anoplura: Linognathidae). J. New York Entomol. Soc 1970b; 78(3): 139-145.
- 17. Miller FH. Scanning electron microscopy of *Echinophthirius horridus* (Von Olfers), *Antarctophthirus callorhini* (Osbern), and *Proechinophthirius fluctus* (Ferris) with emphasis on the antennal structures (Anoplura: Echinophthiridae). J. Parasitol. 1971a; 57(3): 668-674.
- 18. Miller FH. Scanning electron microscopy of antennal structures of five *Haematopinus*

(Anoplura: Haematopinidae). New York Entomol. Soc 1971b; 79: 19-26.

- Qadri MAH. Some new Mallophaga from North-Indian birds. Zeit. für Parasitenkd 1936; 8(6):638-644.
- 20. Dethier VG. Parasitological Reviews. The sensory physiology of blood-sucking arthropods. Exp. Parasitol 1957; 6: 68-122.
- Zacharuk R. Antennae and sensilla. In: Kerkut, G.A. Gilbert, L.I. (Ed.). Comprehensive insect physiology. 1985; V. 6. Oxford Pergaman Press. p. 1-69.
- Mclver SB. Sensilla of haematophagous insects sensitive to vertebrate host-associated stimuli. Int. J. Trop. Insect Sci 1987; 8: 627-635.
- 23. Perez JM, Granados JE, Rutz I. The morphology of *Laemobothrion maximum* (Phthiraptera: Laemobothriidae). Parasitologica. 1995; 57: 45-51.
- Cruz MDS. Antennal sense organs of Phthiraptera (Insecta): Scanning electron microscopy of several species of Anoplura. Micron 1995; 26(1):7-14.
- 25. Smith VS. Avian louse phylogeny (Phthiraptera: Ischnocera): a cladistic study based on morphology. Zool. J. Linn. Soc 2001; 132pp 81-144.
- Baker GT, Chandrapatya A. Sensilla on the mouthparts and antennae of the elephant louse, *Haematomyzus elephantis* Piaget (Phthiraptera: Haematomyzidae). J. Morphol 1992; 214:333-340.
- Green ED, Turner ML. The micromorphological specialization of the claw of the lousefly. J. South Afr. Vet. Ass 2001; 73(2): 124-158.
- 28. Cruz MDS, Mateo MPM. Sensory equipment of the antennal flagellum of several species of *Damalinia*. Micron 1998; 29 (6): 431-438.
- 29. Cruz MD Mateo MP. Structures of the preantennal region of several species of Damalinia (Phthiraptera: Trichodectidae). J. Med. Entomol 2001; 38(6):802-808.
- Cruz MDS, Mateo MMP.Scanning electron microscopy of legs of two species of sucking lice (Anoplura : Phthiraptera). Micron 2009; 40(3): 401 – 408.
- 31. Turner ML. The micromorphology of the blesbuck louse *Damalinia crenelata* as observed under the scanning electron microscope. Koedoe 2003; 46(1): 65-71.
- 32. Turner ML. Labuschagne C, Green ED. The micromorphology of the African buffalo louse *Haematopinus bufali* as observed under the scanning electron microscope. *Koedoe* 2004; 47(2): 83-90.

- Agarwal GP, Ahmad A, Rashmi A, Arya G, Bansal N, Saxena AK. Bio-ecology of the louse, Upupicola upupae, infesting the Common Hoopoe, Upupa epops. J. Insect Sci 2011; 11: Article 26.
- 34. Bhatnager S, Khan V, Ahmad A, Beg S, Singh SK, Singh DK. Antennal sensilla of five poultry lice (Phthiraptera). Proceeding of Zoology and Human Welfare (Allahabad U.P, India) 2004; 211-216.
- 35. Arya G, Ahmad A, Bansal N, Saxena R, Saxena AK. Nature of placodean sensilla of four ischnoceran Phthiraptera. Entomon 2010; 35 (3): 199-202.
- Arya S, Singh S K. Antennal sensilla of head of poultry shaft louse, *Menopon gallinae* (Phthiraptera, Insecta, Menoponidae, Amblycera. JANS 2012; 4 (2): 196-199.

4/8/2014

- Surman A, Suneel K S. Antennal sensilla of head of poultry shaft louse, *Menopon gallinae* (Phthiraptera, Insecta, Menoponidae, Amblycera). JANS 2012; 4 (2): 196-199.
- 38. Clay T. A key to the genera of the Menoponidae (Amblycera : Mallophaga : Insecta). Bulletin of the British Museum (Natural History). Entomol 1969; 24: 3-26.
- 39. Kassner J, Zlotorzycka J. Problems in preparing the antennal sensilla of insects for scanning studies. Wiad. Parazytol 1987; 33(1):93-97.
- 40. Cicchino AC, Abrohamovich AL. Contribution to the knowledge of cephalic sensilla and wateruptake system of adult and nymphs of *Vernoniella bergi* (Kellogg, 1906) (Insecta: Phthiraptera: Ischnocera). Microsc. Electr. Biol. Cell 1988; 12 (2): 121-148.