

Bacterial Isolates from Calves Slaughtered at Abattoir Suffering from Respiratory Problems in Sharkia Governorate

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Abstract: This study was carried out on 300 samples from slaughtered calves (150 heart blood and 150 pieces pneumonic lungs), at different ages and different breed at Sharkia Governorate and subjected to bacteriological examination. Results showed that, 53.3% of blood samples were positive for bacterial examination as 81.2%, 18.8% harbour single and mixed isolates respectively, meanwhile 86.7% of lung samples were positive for bacterial isolation of which 78.5% single isolate and 21.5% mixed isolates. The total number of isolates were 210, of which 90 *Pasteurella multocida* type A (35 heart blood and 40 pneumonic lung and 15 mixed infection), 41 (5 heart blood and 22 lung tissue and 14 mixed infection), 79 *Mannhaemia haemolytica* type A species (25 heart blood, 40 lung and 14 mixed infection). Sensitivity test was carried out using different types of antibiotic most isolated bacteria were highly sensitive to Enoxofloxacin, Tetracycline and florofenicol. The total bacterial count of 10% of samples had count ranging from $10 - 10^8$; while the most samples showed count $10^8 - 10^{11}$.

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1. Introduction

Calf pneumonia is caused by many pathogens and several contributory factors such as warm wet weather, mixing age groups, poor ventilation, stress and high densities (*Hartel et al., 2004*).

Pneumonia is a major cause of economic losses because of decreased production, high levels of mortality and morbidity and increased veterinary and labour costs (*Genaba, et al., 1995*), moreover, calves experienced pneumonia at early age might have severe depression in the production capabilities in the future (*Sayed and Zaitun, 2009*).

Pasteurellosis broadly refers to any of the disease condition caused by species of the genus *Pasteurella* (*Davies et al. 2003*). *Mannheimia haemolytica* formerly known as *Pasteurella haemolytica* (*Christensen et al., 2003*), *Mannheimia haemolytica* is the etiological agent of both bovine and ovine pneumonic pasteurellosis (*Falada, 2002*). *Mannheimia haemolytica* and *Pasteurella multocida* are often associated with bovine respiratory disease in cattle; these two bacteria are considered as part of the normal bacterial flora found in the upper respiratory tract of most cattle but are not considered as normal flora of the lungs (*MaxIrsik, 2010*).

Respiratory troubles to all domestic animals attributed mixed infection with different bacteria isolates (*Mycoplasma bovis*, *Pasteurella* species, *Streptococcus* species, *Staphylococcus aureus*, *Kelbesella* species and *E. coli*) and their toxins (*Yehia, 2000*).

The aim of this work planned to isolation and identification of bacteria with clinical signs of pneumonia in slaughtered calves, determine the effect of antibiotic on the isolated bacteria to reach specific treatment, total bacterial count in lung and.

2. Material and Methods

A total of 300 samples were collected from slaughtered calves of different ages and different breeds, from different abattoirs at Sharkia Governorate, at period from March 2012 to June 2012.

30 white mice, each was 3 weeks age and about 18 – 22 gm body weight were used for purification of *Pasteurella* isolates.

The samples were 150 swabs from heart blood, the samples collected by sterile cotton swabs and transferred immediately to laboratory for bacteriological examination; and 150 lungs were collected from congested lungs area of slaughtered calves, in sterile plastic bages and transported on ice box for bacteriological examination. Each heart blood swabs and congested lungs were cultured into nutrient broth for 24 hours at 37°C and then loopful was taken and sub-cultured into each of the different following media: nutrient agar, 7% sheep blood agar and MacConkey's agar.

The inoculated plates were incubated at 37°C for at least 24 - 72 hours. The growing colonies were picked up and purified by inoculation on nutrient broth and then sub-cultured on selective media and the isolates were identified morphologically by Gram's stain and colonial appearance according to *Finegold and Martin*

(1982), the pure colonies were identified biochemically according to *Koneman, et al. (1983)* and *Krieg and Holt (1984)*.

On the other hand coccobacilli were inoculated in laboratory mice for pathogenicity test and virulence of some isolated strains of pasteurilla were determined according to *Wilson and Miles (1975)*, the serological identification of pasteurilla organisms using pasteurilla antisera (Vet. Serum and Vaccine Research Institute, Abbasia, Cairo) were carried out (*Carter and Rappy, 1962*) for capsular typing with indirect haemagglutination

test and somatic typing as carried (*Heddleston et al., 1972*) using gel diffusion precipitation test.

Purification of *Pasteurella haemolytica* in mice carried out according to *Wessman (1964)*.

Sensitivity test:

The susceptibility of the most predominant pathogenic isolates to different chemotherapeutic agents was tested by using disc diffusion method according to *Finegold and Martin (1982)* and *Cruickshank et al., 1975*.

Total colony count: were examined for 30 specimens according to **American Public Health Association "APHA" (2001)**.

3. Results

Table (1): Results of bacterial examination of examined blood and tissue samples (n = 300)

Sources and type of samples	Total number of samples	Positive		Negative	
		No.	%	No.	%
Heart blood	150	80	53.3	70	46.7
Lung tissue	150	130	86.7	20	13.3
Total	300	210	70	90	30

Table (2): Incidence of bacteria isolated bacterial strains from heart blood and pneumonic lungs of slaughtered calves

Types of samples	Single infection		Mixed infection	
	No.	%	No.	%
Heart blood	65	81.2	15	18.8
Pneumonic lungs	102	78.5	28	21.5
Total	167	79.4	43	20.6

No.= number %= percentage

Table (3): The prevalence of pathogenic bacteria isolated from examined slaughtered calves samples

Type of infection	Type of bacteria	Heart blood		Pneumonic lungs	
		No.	%	No.	%
Single infection	<i>E. coli</i>	5	7.7	22	21.6
	<i>P. multocida</i> (m) serotype A*	35	53.8	40	39.2
	<i>P. (Mannheimia) haemolytica</i> serotype A**	25	38.5	40	39.2
Mixed infection	<i>E. coli</i> with <i>P. multocida</i> serotype A*	7	46.7	15	53.6
	<i>P. multocida</i> type A with <i>Mannheimia haemolytica</i> serotype A	8	53.3	13	46.4

**P. multocida*: Indole +ve

***P. (Mannheimia) haemolytica*: growth on MacCkonky – fermentation arginose sugar

Table (4): Passive mouse for purification of Pasteurella isolates (0.1 ml x 10⁸ S/C)

Group	No. of mouse	1 st day		2 nd day		3 rd day		7 th day	
		Dead	Live	Dead	Live	Dead	Live	Dead	Live
Control	15	0	15	0	15	0	15	0	15
Infected mouse	15	12	3	2	1	1	0	0	0

Table (5): Results of antibiotic sensitivity test to some pathogenic representative bacteria isolates

<i>antibiotic disc</i>	<i>Bacteria isolates</i>		<i>P. multocida</i> Type A (82)		<i>E. coli</i> (30)		<i>P. (Mannheimia) haemolytica</i> type A (75)	
	No.	%	No.	%	No.	%	No.	%
Enrofloxacin 10ug	71	86	27	90	65	87		
Gentamycin 10ug	35	42	25	83	60	80		
Tetracycline 30ug	74	90	26	87	67	89		
Erythromycin 10ug	44	53	11	37	38	51		
Amoxicillin 30ug	59	72	00	00	27	36		
Florofincol 10ug	68	83	17	57	63	84		
Penicillin 100iu	25	30	00	00	10	13		

Table (6): frequency distribution of examined samples based on their total colony count

<i>Specimens</i>	<i>Intervals</i>	<i>Frequency</i>	
		No.	%
Lung	$10^6 - 10^8$	3	10
Lung	$10^8 - 10^{10}$	12	40
Lung	$10^{10} - 10^{11}$	15	50

4. Discussion

Commensal bacteria present in the respiratory system may cause diseases when the animals subjected to stress factors (*Palatary and Newhall, 1985*). In the present study 300 samples (150 heart blood and 150 lung tissue) from slaughtered calves were examined and results obtained revealed that 53.3% and 86.7% respectively were pathogenic bacteria from heart blood and lungs tissue (Table 1), such high incidence of isolation was also reported by *Barbour et al. (1997)* and *Aba-Alkhail and El-Naenaey (2003)*.

Due to respiratory defence mechanism depressed due to immunosuppression associated with virus (*MaxIrisk, 2010*). Prevalence of pneumonia among slaughtered cattle had the highest number of bacteria (*Adamu, and Ameh, 2007*).

Results in table (2) indicate high percentage of isolates refer single infection in heart blood and lung tissue were 81.2% and 78.5% respectively, while mixed isolates in heart blood and lungs tissue were 18.8% and 21.5% respectively. Nearly results agreement with *Genedy et al., (2008)*.

There are several bacteria capable of inducing pneumonia, but the most common organisms are *Pasteurella multocida*, *Pasteurella hemolytica*, and *Hemophilus somnus* (*William and Rebhun, 1982*).

Tables (3 & 4) showed bacteria isolated *E. coli* from heart blood and pneumonic lung were 7.7% and 21.6% or mixed with *Pasteurella*

multocida serotype A were 46.7% and 53.6% respectively. These results reported by *Quinn et al. (1994)*; *Sayed (1996)* and *Sedeek and Thabat (2001)*; also isolated *P. multocida serotype A* from heart blood and pneumonic lungs were 53.8% and 39.2% or mixed with *Mannheimia haemolytica serotype A* were 53.3% and 46.4% respectively. At the same time isolated *Mannheimia haemolytica serotype A* as pure colonies from heart blood and pneumonic lungs were 38.5% and 39.2% respectively. These results were in agreement with *El-Sangary et al. (2008)* and *Genedy et al. (2008)*.

Antibiotic treatment of bacterial pneumonia must be sufficient in duration and, most crucially, early enough to prevent lesions forming that may resist both therapy and regeneration of normal lung parenchyma (*Ingrid Lorenz et al., 2011*).

In table (5), sensitivity test of the antibiotic was carried on 3 different types of bacterial isolates which represent the main causative agents of respiratory manifestation in slaughtered calves, the most of the bacterial isolates were highly sensitive to Enrofloxacin and florofincol and less sensitive to Erythromycin, the results the previous findings of *Roberson, et al. (1994)* and *Singer et al. (1998)*.

Obtained result recorded in table (6) showed that 10% of samples had count ranging from $10^6 - 10^8$, while most samples showed count ranging from $10^8 - 10^{11}$. These results are nearly similar to that obtained by *Vanderline et al. (1999)* and *Paulsen et al. (2006)*.

In conclusion; stress factors as bad hygiene and rise in environmental temperature and increase in

humidity in Egypt lead to activation non pathogenic bacteria which are normal inhabitant of the upper respiratory tract, increase in number and become pathogenic and cause respiratory manifestation characterized by bronchopneumonia which is fibrinous and pleurisy and death occurs due to toxemia and anoxia.

References

1. **Aba-Alkhalil, A. and El-Naenaeey, E.Y. (2003):** Studies on aerobic bacteria causing respiratory infection cattle with special references to zoonotic importance. Suez Canal Vet. Med. J. 2: 157 - 171
2. **Adamu J.Y. and Ameh, J.A. (2007):** Prevalence of pneumonia among slaughtered cattle, goats and sheep in Maiduguri abattoir, Maiduguri, Nigeria. Sahel Journal of Veterinary Science, 6 (1): 112 - 116.
3. **American Public Health Association "APHA" (2001):** Department of Agriculture and Food under the auspices of the European Union Sponsored. P. 179 – 183 Copyright International Association for Food Protection.
4. **Barbour, E.K.; Nabbut, N.H.; Hamadeh, S.K. and Al-Nakhli, H.M. (1997):** Bacterial identity and characteristic in healthy and unhealthy respiratory tract of sheep and calves. Vet. Res. Commun. 21 (6): 421 – 430.
5. **Carter, G.R. and Rappy, D.E. (1962):** Formalized erythrocytes in haemagglutination test or typing *Pasteurella multocida*. British Vet. J., 119: 73 – 77.
6. **Christensen, H.M.; Bisgard, J.; Larsen and Olsen, J.E. (2003):** PCR-detection of *Hemophilus paragallinarum*, *Hemophilus somnus*, *Mannheimia haemolytica*, *Mannheimia* species, *Pasteurella trehalosa* and *Pasteurella multocida*. Methods. Mol. Biol., 216: 257 – 274.
7. **Cruickshank, R.; Duguid, J.R.; Marmion, B.P. and Swain, R.H. (1975):** Microbiology, 13th Ed. Churchill Livingstone, Edinburgh and New York.
8. **Davies, L.R.; MacCorquodale, S. Baillie, and Caffrey, B. (2003):** Characterization and comparison of *Pasteurella multocida* strains associated with porcine pneumonia and atrophic rhinitis. J. Med. Microb., 52: 59 – 67.
9. **El-Sangary, F.; Magda, M. Mohamed; Mokhtar, A.A. and Sahar, E. Saba (2008):** Trial for the use of serum protein electrophoresis as a diagnostic tool for some diseases in buffalo calves. Special Issue for 5th Scientific Conference Suez Canal Veterinary Medicine Journal. XIII (2): 511 – 528.
10. **Falada, S. (2002):** Further *Pasteurella* isolated from the Republic of Zambia: A brief report. Trop. Vet., 20: 130 – 131.
11. **Finegold, S.M. and Martin, W.J. (1982):** Diagnostic Microbiology 6th Ed. C.V. Mosby Co. St. Louis, Toronto, London.
12. **Genaba, R.M.; Bigras-Poluin, D. Belanger and Couture, Y. (1995):** Description of cow-calf productivity in northwestern Quebec and path models for calf mortality and growth. Prev. Vet. Med., 24: 31 – 42.
13. **Genedy, A.M.; Selim, M.A. and Magada, M.M. (2008):** Bacteriological and Virological findings in pneumonic slaughtered calves at Sharkia Governorate. Zag. Vet. J. 36 (2): 79 - 89.
14. **Hartel, H.S.; Nikunen, E.; Neuvonen, R.; Tanskanen and Kivela, S.L. (2004):** Viral and bacterial pathogens in bovine respiratory disease in Finland. Acta Vet. Scand. 45:193 – 200.
15. **Heddleston, K.L.; Gallgher, J.E. and Roberts, P.A. (1972):** Fowl cholera diffusion precipitation test for serotyping of *P. multocida*. Avian Diseases, 16: 925 – 936.
16. **Ingrid Lorenz, Bernadette Earley, John Gilmore, Ian Hogan and Emer Kennedand Simon J More (2011):** Calf health from birth to weaning. III. housing and management of calf pneumonia. Irish Veterinary Journal, 64:14.
17. **Koneman, E.W.; Allen, S.D.; Dozell, V.R. and Summers, H.M. (1983):** Colour Atlas and Text Book of Diagnostic Microbiology, 2nd Edn., Edwards Arnold, London.
18. **Krieg, N.R. and Holt, J.G. (1984):** Bergey's Manual of Systematic Microbiology Vol. 1 William and Baltimore, London.
19. **MaxIrisk, DVM, MAB (2010):** Bovine respiratory disease associated with *Mannheimia haemolytica* or *Pasteurella multocida*. Topic Vet. Med., IriskMax, Cattle diseases volum163.
20. **Palatary, J. and Newhall, J. (1985):** Pneumonia in newly weaned calves. Am.Vet. Assoc., 133: 353 – 359.
21. **Paulsen, P.; Schopf, E. and Sumulders, F.J.M. (2006):** Estimation of total aerobic and E. coli in minced meat. Journal of Food Protection. 29 (10): 2500 – 2503.
22. **Quinn, P.J.; Carter, M.E.; Markey, B.K. and Carter, G.R. (1994):** Clinical Veterinary Microbiology Wolfe Virginia. U.S.A.
23. **Roberson, J.R.; Fox, L.K.; Hamcock, D.D.; Gay, J.M. and Beesser, T.B. (1994):** Etiology of *Staphylococcus aureus* isolated from various sites on dairy farms. J. Dairy Science, 77 (11): 3354 -3364.
24. **Sayed, A.M. (1996):** Some Bacteriological studies on sheep pneumonia at Assuit

- Governorate. Assuit Vet. Med. J. 36 (71): 68 – 73.
25. **Sayed, S.M. and Zaitoun, M.A. (2009):** Aerobic bacterial pathogens of pneumonic feedlot buffalo-calves in Assuit governorate, Egypt, Ass. Univ. Bull Enviro. RES.12: 55 - 62.
 26. **Sedeek, S.R. and Thabat, A.M. (2001):** Some studies on bacterial causes of pneumonia in cattle in Assuit Governorate. Assuit Vet. Med. J. 45 (90): 243 – 255.
 27. **Singer, R.S.; Case, J.T.; Carpenter, T.E.; Walker, R.L. and Hirsh, D.C. (1998):** Assessment of spatial and temporal clustering of ampicillin and tetracycline resistant strains of *Pasteurella multocida* and *Pasteurella haemolytica* isolated from cattle in California. J. Am. Vet. Med. Assoc., 212 (7): 1001 – 1005.
 28. **Vanderline, P.B.; Shay, B. and Murray, J. (1999):** Microbiological status of Australian sheep meat. Journal of Food protection. 62 (4): 380 – 385.
 29. **Wessman, G.E. (1964):** Enterluration of smooth and non-smooth variant in the dissociation of *P. Haemolytica*. J. Bacteriology, 88: 356 – 360.
 30. **William, Y. and Rebhun, C. (1982):** A review of infectious bovine rhinotracheitis, shipping fever pneumonia and viral-bacterial synergism in respiratory disease of cattle. Can. J. Comp. Med., 46 (3): 225 – 263.
 31. **Wilson, G.S. and Miles, A.A. (1975):** Topley and Wilson principles of Bacteriology, virology and Immunology. Vol. 16th Ed., Edward, Arnold, London.
 32. **Yehia, R.M. (2000):** Respiratory problems in calves. MVSc, Thesis, Faculty of Vet. Med., Zagazig, University.

2013/12/3