

Effect Of Crude Ethanol Extract Of *Nauclea Latifolia* On Some Clinical Isolates Of Food Importance And Its Toxicological Potentials

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Abstract: The leaves of *Nauclea latifolia* used in traditional medicine for the treatment of diarrhea were extracted in hot and cold ethanol with the aim of determining their antibacterial activities and toxicological potentials. Soxhlet extraction method was used for the hot ethanol extraction while for the cold ethanol ground samples were soaked in ethanol for 48h. The Agar diffusion method was used for the antibacterial assay at different concentrations on *Escherichia coli*, *Staphylococcus aureus*, *Pseudomonas aeruginosa* and *Salmonella typhi*. Albino rats were used for toxicological studies by injecting varying doses of the extracts through the intraperitoneal route for 14 days. The growth of *S. typhi* was not inhibited by the extract. However the hot ethanol extract had minimum inhibitory concentrations (MICs) of 3.24mg/ml, 3.28mg/ml and 4.82mg/ml on *E. coli*, *S aureus* and *P. aeruginosa* respectively, while the cold ethanol extract produced MICs of 4.74mg/ml, 5.14mg/ml and 5.61 mg/ml respectively on the isolates. Hematological analyses revealed that RBC, PCV and Hb values decreased with increase in doses of extract while ESR and WBC values increased. MCHC values were lower than that obtained from the control. However, statistical analyses revealed that there were significant differences in the values obtained for RBC, PCV and Hb at the lower doses and the controls. The extracts contained alkaloids, tannins and saponins. The antibacterial assay justifies the use of the plant in palm wine preservation and possible use in preservation of other foods, and the treatment of diarrhoea.

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Key words: Antibacterial; toxicological; preservation; ethanol extracts; *Nauclea latifolia*; clinical isolates.

Introduction

Throughout recorded history, spices and herbs have been used for flavouring foods and beverages, and for medicinal purposes. Also a wide range of natural products from plants can be useful in extending shelf life of foods, reducing or eliminating pathogenic bacteria, and increasing overall quality of food products (Droughon, 2004).

There are estimations that as much as 250,000-500,000 plant species are on Earth (Boris, 1996) and thousands of compounds have been isolated from these plants which are claimed to possess antimicrobial or medicinal properties (Schultes, 1978; Cowan, 1999). Although numerous studies

have been done in vitro to evaluate the antimicrobial activity of botanicals, only a few have been done with food products (Doughon, 2004; Hancock and Harrison, 2002; Tsigarida et al., 2000; Gill et al., 2002; Diaz et al., 2002; Lemay et al., 2002). In Nigeria also some ethnobotanical texts are available which describe the species that possess antimicrobial and medicinal properties (Iwu, 1993; Soforowa, 1982; Igoli et al., 2005). Some have been found to inhibit the growth of some food and clinical isolates (Akujobi et al., 2004; Esimone et al., 1998; Ogbulie et al., 2004; Ogueke et al., 2006; Ogbulie et al., 2007; Ogueke et al., 2007a; Ogueke et al., 2007b). Amongst these plants that possess medicinal properties is *Nauclea latifolia*.

60 *Nauclea latifolia* belongs to the family Rubiaceae
61 (Ntiejumokwu and Kolawole, 1991) and is a
62 common plant in the tropical forests of South
63 Eastern Nigeria. It is commonly used by palm wine
64 tappers as a preparative ("nche") for palm wine. In
65 traditional medicine the leaves and the stem bark are
66 used in the treatment of diarrhea (Igoli et al., 2005). It
67 is called "African quinine in Northern Nigeria, a cold
68 infusion of the bark is taken as a diuretic and
69 anthelmintic. The Fulanis in Nigeria uses the leaf
70 extract to regularly deworm animals (Adebowale
71 1993). The ethanol extract have been found to
72 decrease the level of parasitaemia in a dose-
73 dependent manner in mice experimentally infected
74 with a *Trypanosoma brucei* (Morah, 1998). Different
75 indolo-quinolizidine alkaloids and glycol-alkaloids
76 have been isolated from the root bark. The former has
77 been identified and named angustine, angustoline.
78 The glyco alkaloids have been identified as
79 cadambine 3-a-dihydro cadambine (Hottellier et al,
80 1975).

81 There is therefore need to authenticate the
82 antibacterial effects of the crude plant extract,
83 especially on the diarrheal causing microorganisms,
84 and determine their toxicological potentials, at least
85 on laboratory animals. This is a prelude to
86 determining their preservative potentials in a food
87 system. Thus the objectives of this, work were to
88 authenticate the antibacterial properties on some
89 clinical isolates of food importance and determine the
90 toxicological potentials on albino rats.

91 **Materials and Methods**

92 Plant collection and identification: Fresh leaves
93 of *N. latifolia* were obtained from Obinze, Owerri
94 West Local Government Area of Imo State, Nigeria
95 in August, 2009. The plant was certified by Dr. I. I.
96 Ibeawuchi of the Department of Crop Science
97 Technology, Federal University of Technology,
98 Owerri. Specimen voucher was deposited in the
99 herbarium of the Department of Crop Science
100 Technology, Federal University of Technology,
101 Owerri, Nigeria.

102 **Sample preparation and extraction procedure**

103 The fresh leaves were air dried and ground into
104 fine powder using a mechanical grinder. Two
105 methods, the hot ethanol extraction and cold ethanol
106 extraction techniques were employed for the
107 extraction of the plant's active principles.

108 For the hot ethanol extraction technique, 20g of
109 the ground plant material were wrapped in Whatman

110 filter paper, each wrap containing 2.0g. They were
111 put in the timble of a Soxhlet apparatus. Then 250 ml
112 of 95 % ethanol was put in a round bottom flask and
113 this was used to mount the Soxhlet apparatus. The
114 round bottom flask was heated and extraction of the
115 plant material was stopped after seven refluxes. The
116 solution was then evaporated to dryness using a
117 rotary evaporator (model type 34/2; Corning Ltd,
118 England). Yield of 17.95% was obtained in relation
119 to the powdered material.

120 For the cold ethanol extraction technique 20g of
121 the ground plant sample was weighed into 250 ml of
122 ethanol (95%) in a conical flask. It was covered,
123 shaken every 30 mins, for 6h and thereafter allowed
124 to stand for 48h for extraction. The solution at the
125 end of extraction was shaken and filtered using
126 Whatman filter paper. The filtrate was subsequently
127 evaporated to dryness using a rotary evaporator.
128 Yield of 17.60% was obtained in relation to the
129 powdered plant material.

130 The extracts obtained were thereafter stored at
131 10°C in amber coloured bottles until required.

132 **Preparation of stock solutions of extracts**

133 The method of Akujobi et al., (2004) was adopted
134 for the preparation of stock solutions. The crude
135 extracts obtained were diluted with 20% dimethyl
136 sulphoxide (DMSO) solution to obtain 50mg/ml,
137 25mg/ml, 12.5mg/ml, 6.25mg/ml and 3.2mg/ml
138 concentrations. They were stored at 10°C in amber
139 coloured bottles until required.

140 **Test microorganisms and their sources**

141 The bacterial isolates used for the study were
142 clinical isolates of food importance. These were
143 *Escherichia coli* (5 isolates), *Staphylococcus aureus*
144 (5 isolates), *Salmonella typhi* (7 isolates) and
145 *Pseudomonas aeruginosa* (3 isolates). They were
146 obtained from the Microbiology Laboratory of
147 Federal Medical Centre (FMC) Owerri, Nigeria.
148 They were re-identified, sub-cultured on Nutrient
149 agar slants for *Escherichia coli* and *Pseudomonas*
150 *aeruginosa*, Baird Parker agar slant for
151 *Staphylococcus aureus*, Deoxycholate citrate agar
152 slant for *Salmonella typhi* and stored at 4°C until
153 required.

154 **Evaluation of antibacterial activity**

155 The well in agar diffusion method as described by
156 Esimone et al.,(1998) and Osadebe and Ukwueze
157 (2004) was adopted for the study. Standardized
158 Nutrient broth cultures of the test isolates containing

159 approximately 10^7 cells/ml organisms were used.
160 0.1ml of the broth cultures were introduced into
161 sterile Petri dishes and 15mls of molten Nutrient agar
162 poured into the Petri dishes. The contents were
163 thoroughly mixed and allowed to solidify. Three
164 holes each measuring 5.0mm in diameter were made
165 in each of the solid agar plates using a sterile cork
166 borer. 0.04ml of the different concentrations of plant
167 extracts were transferred into the holes using a
168 Pasteur pipette. Two Petri dishes containing a
169 particular bacterium were used for each concentration
170 of the extracts. The plants were thereafter allowed to
171 stand for one hour for pre-diffusion of the extracts
172 (Esimone et al., 1998) and were subsequently
173 incubated at 37° C for 24 h.

174 After incubation, the plates were collected and
175 the zones of growth inhibition were measured. The
176 minimum inhibitory concentrations (MICs) of the
177 extracts were determined by plotting a graph of the
178 log of concentrations used (x - axis) against the
179 squares of the zones of growth inhibition (y- axis). A
180 regression line was then drawn through the points
181 and the antilogarithm of the value at the intercept on
182 the x- axis gave the MIC values (Osadebe and
183 Ukwueze, 2004; Esimone et al., 1998).

184 Evaluation of haematological toxicity on albino 185 rats

186 Initial LD₅₀ studies were carried out to
187 determine the maximum dose of extracts that will not
188 produce any death on the rats. Based on the LD₅₀
189 studies, four groups of albino rats (male) each
190 comprising three rats, randomly selected and
191 weighing 132.5g was used. Doses of 30.2 mg/kg
192 body weight, 60.4mg/kg body weight, 120.8 mg/kg
193 body weight and 241.6 mg/kg body weight were
194 injected into each group through the intraperitoneal
195 route (Iyaniwura et al., 1991, EFPIA/ECVAM, 2001)
196 on daily basis for 14 days. The control group was
197 injected with the diluent (20% DMSO solution).
198 Food and water were provided adlibitum.

199 On the 15th day, the animals were collected and
200 blood samples drawn from the sublingual vein
201 according to the method described by Zeller et al.,
202 (1998). This method has been found suitable for
203 laboratory animal's well being as stated in EFPIA/
204 ECVAM (2001). 3.0ml of blood sample was
205 immediately transferred to ethylene di-amine tetra
206 acetic acid (EDTA) treated bottles for hematological
207 assay. They were analyzed within 3h of collection for
208 total erythrocyte (RBC), leukocyte (WBC) counts,
209 packed cell volume (PCV), haemoglobin (Hb)
210 contents, serum glutamate pyruvic transaminase

211 (SGPT) and serum glutamic oxaloacetic transaminase
212 (SGOT) according to the methods described by
213 Okeudo et al., (2003). ESR was determined
214 according to the method described by Okeudo et al.
215 (2003) and Iheukwumere et al. (2002). Varied
216 haematological indices were calculated from the
217 results obtained. These included mean corpuscular
218 volume (MCV), mean corpuscular haemoglobin
219 (MCH), and mean corpuscular haemoglobin
220 concentration (MCHC).

221 Preliminary phytochemical analysis of extracts

222 These were carried out according to the methods
223 described by Trease and Evans (1989) for
224 determination of alkaloids, tannins, saponins,
225 flavonoids, cardiac glycosides and cyanogenic
226 glycosides.

227 Analysis of data

228 Statistical analyses were conducted on the data
229 obtained from the study using Analysis of Variance
230 (ANOVA). The means were separated using Fisher's
231 Least Significant Difference (Sanders, 1990).

232 Results

233 The results obtained from the study showed that
234 in general the hot ethanol extracts produced greater
235 inhibitory effect on the isolates than the cold ethanol
236 extracts.

237 Table 1 shows the antibacterial activity of the
238 crude hot ethanol extract of the plant on the isolates.
239 *E. coli* and *S. aureus* were inhibited by all the
240 concentrations applied while *S. typhi* was not affected
241 by any of the concentrations. The 3.2 mg/ml
242 concentration of the extract did not inhibit the growth
243 of *P. aeruginosa* while the other concentrations had
244 inhibitory effects on the bacterium.

245 Table 2 shows the antibacterial effect of the cold
246 ethanol extracts on the isolates. All the isolates were
247 not inhibited by the 3.2mg/ml concentration of the
248 extract. *S. typhi* was not affected by all the
249 concentrations of the extract.

250 Table 3 shows the MICs of the extracts on the
251 bacterial isolates. The lowest MIC was produced by
252 the hot ethanol extract on *E. coli* with MIC value
253 3.24 mg/ml. The highest MIC value was obtained
254 from the cold ethanol extract with a value of 5.61
255 mg/ml on *P. aeruginosa*. In general the hot ethanol
256 extract produced lower MIC values than the cold
257 ethanol extract.

258 Table 4 shows the results of the haematological
259 analyses of the blood samples obtained from rats
260 injected with different doses of the extracts. It was
261 observed that increase in doses of extract resulted in
262 decrease in the RBC, PCV and Hb values of the
263 blood samples. However there was no significant
264 difference in these values with those obtained from
265 the control up to 60.4 mg/kg body weight. ESR and
266 WBC values increased with increase in doses
267 applied.

268 Statistical analysis revealed that there was no
269 significant difference in values and the control at 30.2

270 mg/kg body weight dose for ESR and WBC
271 respectively. However, for MCV, MCH and MCHC,
272 there was no ordered pattern in the values as doses
273 administered increased. Values obtained for MCV
274 and MCH were higher than that obtained for the
275 control while the values of MCHC were lower than
276 the control.

277 Table 5 shows the result of the preliminary
278 phytochemical analyses of the extracts. Tannins,
279 alkaloids and saponins were identified in the extracts.

280

281

282 Results

283 Table 1: *Antibacterial activity of crude hot ethanol extract of *N. latifolia* on isolates

Concentration of Extract (mg/ml)	Mean diameter zone of inhibition (mm)			
	<i>E. Coli</i>	<i>S. aureus</i>	<i>P. aeruginosa</i>	<i>S. typhi</i>
3.2	8.7	6.4	-	-
6.25	13.8	11.6	7.5	-
12.5	19.7	16.3	14.8	-
25.0	26.6	22.3	21.9	-
50.0	32.2	29.9	25.5	-

284 *Results are average of the triplicate determinations of the isolates

285

286 Table 2:* Antibacterial activity of crude cold ethanol extract of *N. latifolia* on isolates.

Concentration of Extract (mg/ml)	Mean diameter zone of inhibition (mm)			
	<i>E. Coli</i>	<i>S. aureus</i>	<i>P. aeruginosa</i>	<i>S. typhi</i>
3.2	-	-	-	-
6.25	7.2	6.3	5.9	-
12.5	10.5	9.7	9.6	-
25.0	14.6	13.5	13.8	-
50.0	17.9	17.1	17.9	-

287 *Results are average of the triplicate determinations of the isolates

288

289 Table 3: The minimum inhibitory concentration (MIC) of *N. Latifolia* extracts on isolates.

Minimum inhibitory Concentration (mg/ ml)					
Type of extract	<i>E. coli</i>	<i>S. aureus</i>	<i>P. aeruginosa</i>	<i>S.aureus</i>	
Hot ethanol	3.24 ^a	3.28 ^a	4.82 ^b		NIL
Cold ethanol	4.74 ^a	5.14 ^b	5.61 ^b		NIL

290 a,b....values on the same row with the same superscript are not significantly different [P=0.05]

291 Table 4: Results of the haematological toxicity tests on rats injected with varying doses of *N. latifolia* ethanol extract.
292

Doses of extracts administered [mg/kg body weight]						
Parameters	Control	30.2	60.4	120.8	241.6	
RBC (x10 ⁶ cells/mm ³)	5.32 ^a	4.78 ^{a,b}	4.49 ^{a,b}	4.01 ^b	3.93 ^b	
PCV (%)	36.3 ^a	35.4 ^a	35.0 ^a	33.6 ^b	32.6 ^b	
ESR (mm/hr)	3.50 ^a	3.60 ^a	4.14 ^b	5.03 ^c	5.51 ^c	
MCV (cubic microns)	68.23 ^a	74.06 ^b	77.95 ^c	83.79 ^d	82.95 ^d	
Hb (g /100ml)	9.8 ^a	9.5 ^a	9.2 ^{a,b}	9.0 ^b	8.4 ^a	
MCHC(%)	27.00 ^a	26.84 ^a	26.29 ^b	26.78 ^a	26.69 ^a	
WBC(X10 ³ cells / mm ³)	4.77 ^a	4.96 ^{a,b}	5.02 ^{a,b}	5.39 ^{a,b}	5.73 ^b	

293 a, b..... values on the same row with the same superscript are not significantly different (P = 0.05)

294

295 Table 5: Phytochemical analysis of crude hot and cold ethanol extracts of *N. latifolia*.

Type of extract	Alkaloids	Tanins	Saponins	Flavonoids	Cardiac glycosides	Cyanogenic glycosides
Hot Ethanol	+	+	+	-	-	-
Cold ethanol	+	+	+	-	-	-
+ =	Present	- =	Absent			

296

297

298

299 Discussion

300 The results obtained from this study showed that
301 the extracts inhibited the growth of the isolates
302 except *S. typhi*. However, the hot ethanol extract had
303 greater inhibitory effect on the isolates than the cold
304 ethanol extract. That the extract inhibited the growth
305 of the isolates is an indication that they contain
306 substance(s) that are active against bacteria. Other
307 workers have also shown that extracts of plants
308 inhibit the growth of various bacteria (Akujobi et al.,
309 2004, Esimone et al., 1998, Nweze et al., 2004
310 Osadebe and Ukwueze, 2004, Ntiejumokwu and
311 Kolawole, 1991). That the extract did not inhibit the
312 growth of *S. typhi* may be due to the fact that the
313 bacterium possesses mechanisms for detoxifying or
314 removing the active principles. Some bacteria such as
315 *S. aureus* detoxify penicillin by converting it to
316 penicillanic acid (Braude, 1982). The observed
317 antibacterial activities of the extracts may be due to
318 tannins, alkaloids and saponins, either singly or in
319 combination, identified in the extracts. Some workers
320 (Hottellier *et al*, 1975) have identified indolo-
321 quinolizidine alkaloids and glycol-alkaloids (
322 cadambine 3-a-dihydro cadambine) in the root barks.
323 These groups of compounds have been identified to
324 possess antibacterial properties (Draughon, 2004).

325 That the hot ethanol extract produced greater
326 inhibitory effect than the cold ethanol extract shows
327 that the extraction method employed affects the yield
328 of the active principles even when the same solvent is
329 used. Probably application of vigorous heat in an
330 enclosed system such as is found when using the
331 Soxhlet apparatus increases the rate and yield of
332 extraction of the active principles. It could therefore
333 be advised that such method be employed for
334 extraction of plant active principles. Some workers
335 (Ogbulie et al., 2004) have also shown that Soxhlet
336 extraction method is more efficient in the extraction
337 of plant active principles than other methods of
338 extraction.

339 The results obtained from the determination of
340 the MICs indicate that the extracts, especially the hot
341 ethanol extracts, could inhibit these isolates at low
342 concentrations. This is most evident in the MIC of
343 the hot ethanol extract on *E. coli*. Such extracts that
344 could produce such low MIC on microorganisms
345 could be of great importance in food preservation
346 especially as they are natural, where they could be
347 used in place of the common synthetic preservative
348 substances. This will reduce the usual health risks
349 associated with the use of these substances. Several
350 works are in literature showing that plant extracts can
351 be used for preservation of certain foods (Hancock

352 and Harrison, 2002; Tsigarida et al., 2000; Cate et al.,
353 2000; Elgayyar et al., 2001 Diaz et al., 2002; Lemay
354 et al., 2002; Pszozola, 2002; Leuschner and
355 Zamparini, 2002; Gill et al., 2002). Thus upon
356 purification to remove other compounds in the
357 extract, better results could be achieved. Some
358 workers have shown that further purification of
359 extracts could enhance their antibacterial properties
360 (Okoli and Iroegbu, 2004).

361 The results are also of significance in the health
362 care delivery system; since they could be used as
363 alternatives to orthodox antibiotics in the treatment of
364 infections caused by these organisms, especially as
365 these organisms frequently develop resistance to the
366 orthodox antibiotics (Singleton, 1999). The level of
367 growth inhibition exhibited by the extract especially
368 on *E. coli* justifies the use of the plant by traditional
369 medical practitioners in the treatment of diarrhea
370 (Igoli et al., 2004) and its use in the preservation of
371 palm wine by palm wine tappers. *E. coli* is known to
372 be a major cause of various diarrhoeagenic infections
373 (Adams and Moss 1999) in the developing countries.

374 Results from the haematological analyses of the
375 blood samples revealed that RBC, PCV and Hb
376 values decreased with increase in dose of extract
377 administered. The consistent decrease in the RBC
378 count with increasing level of extract administered is
379 an indication that in its present crude form the extract
380 may be destroying circulating erythrocytes or may
381 have impaired the blood forming (erythropoietic)
382 centres of the rats. However, statistical analysis
383 revealed that up to 60.4 mg/kg body weight, the
384 values were not significantly different from the
385 values obtained from the control, indicating that at
386 those lower doses the extract may not adversely
387 affect the erythrocytes. Some other workers (Aniagu
388 et al., 2005) however, have shown that some other
389 plant extracts could remarkably increase the level of
390 red blood cells.

391 Since PCV is an indication of the ratio of the red
392 cell component to the total blood volume (Smith et
393 al., 1974; Fischbach, 1980), it would seem that
394 increase in dose of the extract decreased the red cell
395 component, thus indicating that the effect of the
396 extract was on the circulating red cells. However,
397 statistical analysis also revealed that up to 60.4 mg/kg
398 body weight, the values were not significantly
399 different from the value obtained from the control.

400 The consistent decrease in Hb with increase in
401 dose of extract is an indication that the extract
402 destroyed the haemoglobin in the red blood cells or
403 they impaired the uptake and utilization of iron by the

404 rats. Sokunbi and Egbunike (2000) and Iheukwumere
405 et al., (2000) have suggested that some plant extracts
406 affect iron uptake and utilization by animals. It could
407 also be that the extract affected the uptake and
408 utilization of copper, an element necessary for the
409 utilization of iron in the production of haemoglobin
410 (Smith et al., 1974). Statistical analysis, however,
411 showed that up to 60.4 mg/kg body weight the values
412 were not significantly different from that obtained
413 from the control.

414 Results obtained for MCV and MCHC show that
415 the extract induced hypochromic macrocytic anaemia
416 in these rats. Smith et al. (1974) stated that the
417 outpouring of less mature erythrocytes (reticulocytes)
418 in response to the destruction of circulating red blood
419 cells usually results in increased MCV and also
420 usually contain lower percentage of Hb per
421 erythrocyte.

422 For the white blood cells (WBC) the values
423 increased with increase in doses of extracts. It could
424 be that the presence of the extract stimulated the
425 production of more WBC, probably to fight an
426 intruder (the extract). This however, is normal.
427 Statistical analysis revealed that there was no
428 significant difference between the values obtained at
429 different doses and the control.

430 The results obtained from the study are of
431 significance as the plant is used in traditional
432 medicine for the treatment of diarrhea. More studies
433 should be conducted to determine the effectiveness of
434 the extract upon refinement on preservation of food,
435 and the administration of the extract on the vital
436 organs of the body.

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